

## PERSPECTIVES

# Bisphosphonates and $\gamma\delta$ T-Cells: New Insights into Old Drugs

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### Abstract

Bisphosphonates (BPs) are now well established as front-line treatments for post-menopausal osteoporosis, Paget's disease and tumor-associated osteolysis and hypercalcemia. Since the basic pharmacology and mechanisms of action of these bone-targeting agents have been reviewed in detail elsewhere (1), we summarize new insights into their mode of inhibition of FPP synthase, concepts of their anti-resorptive effects on osteoclasts and, in particular, discuss their recently-discovered effects on  $\gamma\delta$  T-cells and prospects for their future use as immune therapies. *BoneKEy-Osteovision*. 2006 August;3(8):5-14.  
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### Current Concepts of Cellular Uptake and Molecular Actions in Osteoclasts

Because of the ability of BPs to bind avidly to bone mineral, approximately 50% of a circulating dose of BP binds to the skeleton. Recent studies with fluorescently labelled BPs have shown that after release of BPs into the acidic environment of the osteoclastic resorption lacuna, osteoclasts internalize BPs, probably as microprecipitates with  $\text{Ca}^{2+}$ , into membrane-bound vesicles by fluid-phase endocytosis (2;3). In addition to enabling release from the mineral surface, the acidifying activity of  $\text{H}^+$ -ATPases is also essential for the uptake of BPs into the cytosol of osteoclasts (2;3), probably since acidification of endocytic vesicles reduces the negative charge on the phosphonate groups of BPs, allowing dissociation from  $\text{Ca}^{2+}$  ions and subsequent diffusion or transport of BPs across the vesicular membrane (2).

Once internalized by osteoclasts, BPs can be separated into two groups according to their intracellular molecular mechanism of action. The simple BPs that most closely resemble pyrophosphate, such as clodronate and etidronate, are metabolized into methylene-containing analogues of ATP by cytosolic aminoacyl-tRNA synthetase enzymes, with the non-hydrolyzable P-C-P group of the BP replacing P-O-P (i.e. the  $\beta,\gamma$ -phosphate groups) in ATP (4). These AppCp-type metabolites accumulate in the

osteoclast cytosol and induce apoptosis, thereby inhibiting bone resorption (1). Nitrogen-containing bisphosphonates (N-BPs), however, are not metabolized. Rather, N-BPs act by inhibiting farnesyl diphosphate synthase (FPP synthase), a peroxisomal enzyme of the mevalonate pathway (1). The recent generation of X-ray crystal structures of human FPP synthase co-crystallized with N-BPs (5;6), together with detailed kinetic analysis of the recombinant human enzyme, indicates that the interaction with N-BPs is highly complex and characteristic of 'slow-tight binding' inhibition (5). Initially, N-BPs appear to compete directly for binding with the isoprenoid lipid substrates DMAPP or GPP, forming interactions between the nitrogen moiety of the N-BP and a conserved threonine and lysine residue in the enzyme. This is associated with complex conformational changes in the enzyme that promote the binding of the second substrate, IPP, which itself stabilizes the final ternary, "locked" complex of N-BP-bound enzyme, helping to explain why some N-BPs, such as zoledronate, are such potent inhibitors of this enzyme (5;7).

### Inhibition or Activation of Small GTPases?

Inhibition of FPP synthase prevents the synthesis of the isoprenoid lipids FPP and geranylgeranyl diphosphate (GGPP), which are required for the post-translational prenylation of small GTPases such as those

of the Ras, Rho and Rab families. Since prenylation is required for the localization of these GTPases to subcellular membranes, N-BPs inhibit resorption by disrupting the normal localization and function of small GTPases that are essential for osteoclast activity and survival (1). It has been generally assumed that N-BPs, by preventing protein prenylation, disrupt the membrane interaction of small GTPases and thereby inhibit downstream signaling pathways. Surprisingly, we demonstrated recently that the unprenylated forms of Rho, Rac and Cdc42 GTPases that accumulate in osteoclasts and macrophages following treatment with N-BPs are in the active GTP-bound form (8). Although, in the absence of prenylation, unprenylated small GTPases are mislocalized and would therefore not be expected to be able to activate downstream signaling pathways, N-BPs cause sustained activation of p38 downstream of Rac, at least in macrophages (8). These observations suggest that N-BPs, rather than causing the loss of pre-existing, prenylated small GTPases and hence loss of downstream signaling in osteoclasts, may inhibit osteoclast function at least in part by causing the accumulation of the unprenylated form of small GTPases. This accumulation would result in inappropriate activation of some downstream signaling pathways, and/or a dominant negative effect on small GTPase signaling due to the formation of non-productive, cytoplasmic complexes of unprenylated GTPases with effector proteins. Further studies are required to determine which mechanisms occur in osteoclasts and in other cell types that might be affected by N-BPs *in vivo*, such as tumor cells (9).

### The Acute Phase Reaction to Bisphosphonates

The profound anti-resorptive effect of a once-yearly infusion of zoledronic acid (10), or more frequent infusions of ibandronate (11), indicates that the intravenous administration of BPs is an attractive therapeutic alternative to oral administration. The most common adverse-effect of intravenous N-BP administration, which is not generally observed with oral dosing, is the development of a flu-like syndrome called the acute-phase reaction (12) that typically occurs in one-third of patients receiving intravenous N-BPs for the first

time. While this syndrome was first described nearly 20 years ago, the exact molecular mechanism underlying this adverse effect has only recently been elucidated. In a seminal study by Kunzmann *et al.* in 1999, the authors reported that patients that suffered an acute-phase reaction to pamidronate had increased levels of circulating  $\gamma\delta$  T-cells up to 28 days following the initial infusion (13).  $\gamma\delta$  T-cells are non-conventional T-cells that, unlike conventional  $\alpha\beta$  T-cells, can recognize antigen without the need for presentation by MHC-class molecules. In humans,  $\gamma\delta$  T-cells comprise only a minor proportion (1-10%) of CD3<sup>+</sup> T-cells in peripheral blood, and the majority (50-90%) of these  $\gamma\delta$  T-cells belong to the V $\gamma$ 9V $\delta$ 2<sup>+</sup> subset (also termed V $\gamma$ 2V $\delta$ 2<sup>+</sup>). V $\gamma$ 9V $\delta$ 2<sup>+</sup> T-cells are activated by a diverse array of non-peptide molecules, such as pyrophosphomonoesters (IPP, DMAPP) (14) and the synthetic agonist bromohydrin pyrophosphate (BrHPP) (15), alkylamines (16), and N-BPs (13;17). Since N-BPs closely resemble isoprenoid pyrophosphates such as IPP, DMAPP and GPP (which is why N-BPs such as pamidronate and zoledronate inhibit FPP synthase) (5;6;18), this activation of V $\gamma$ 9V $\delta$ 2<sup>+</sup> T-cells by N-BPs was initially thought to occur by a direct agonistic effect of the N-BP on the V $\gamma$ 9V $\delta$ 2<sup>+</sup> T-cell receptor (TCR). However, it is now clear that inhibition of FPP synthase is responsible for the activation of V $\gamma$ 9V $\delta$ 2<sup>+</sup> T-cells by N-BPs.

Following an intravenous infusion of N-BP, the plasma concentration (approximately 10<sup>-6</sup>M for zoledronic acid) (19) is probably sufficiently high to allow the internalization of N-BP into endocytic cells such as monocytes (20). The subsequent inhibition of FPP synthase causes intracellular accumulation of the substrates of this enzyme (IPP and DMAPP) (21-23), which are known agonists of the V $\gamma$ 9V $\delta$ 2<sup>+</sup> TCR (14). This intracellular IPP is then somehow 'presented' to V $\gamma$ 9V $\delta$ 2<sup>+</sup> T-cells in the peripheral circulation (by an unknown mechanism), thereby causing their activation (13;17) and the release of the pro-inflammatory cytokines TNF $\alpha$  and IFN $\gamma$  characteristic of the acute-phase reaction (24-26) (See Figure 1). This hypothesis is supported by the finding that N-BPs must be internalized in order to stimulate V $\gamma$ 9V $\delta$ 2<sup>+</sup> T-cells *in vitro* (21), and that the ability of N-

BPs to activate  $V\gamma 9V\delta 2^+$  T-cells closely matches the potency of the N-BP for inhibiting FPP synthase (22). We have recently shown that dietary alkylamines, which are found in numerous edible plants and plant products, also *indirectly* activate  $V\gamma 9V\delta 2^+$  T-cells through a similar mechanism to N-BPs (i.e. inhibition of FPP synthase and the accumulation of IPP) (27).

Furthermore, statins can completely prevent the stimulatory effect of N-BPs on  $V\gamma 9V\delta 2^+$  T-cells *in vitro* (21-23) by preventing the accumulation of IPP through inhibition of HMG-CoA reductase, an enzyme upstream of IPP synthesis. This raises the intriguing possibility that statins could be used to prevent the acute-phase reaction to N-BPs *in vivo* (22;23).

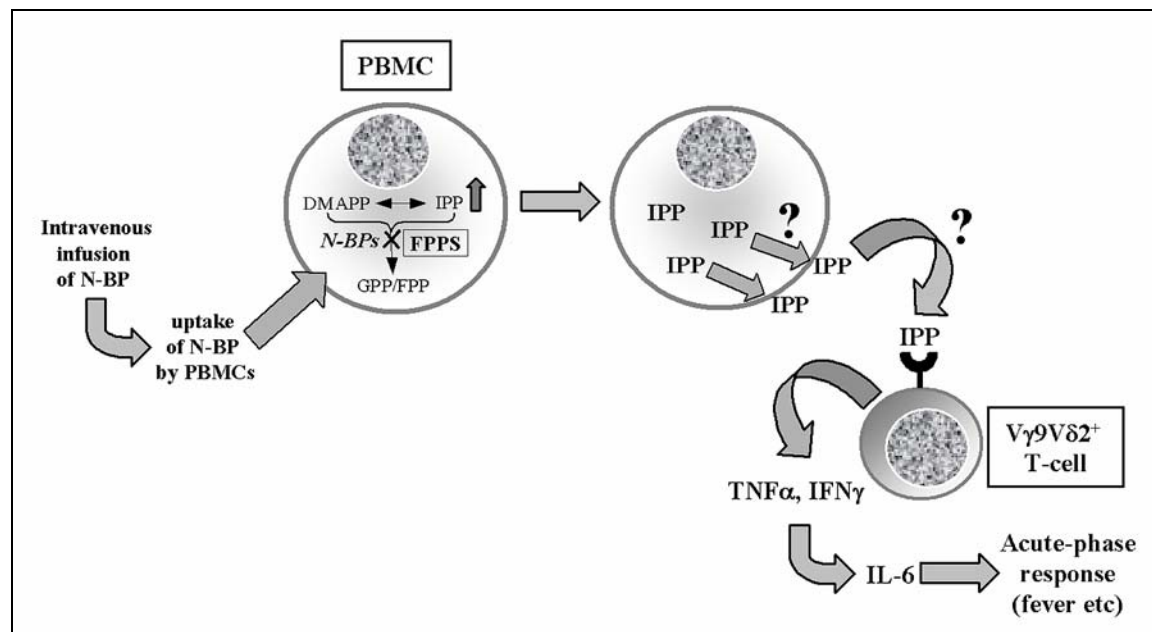


Figure 1. Current model of the molecular mechanism underlying the acute-phase reaction to N-BPs *in vivo*. Following an intravenous administration of N-BP, endocytic cells such as monocytes internalise N-BP, resulting in the inhibition of FPP synthase and the accumulation of IPP/DMAPP. IPP/DMAPP is then presented to  $V\gamma 9V\delta 2^+$  T-cells by an as yet unidentified mechanism, causing their activation and the release of pro-inflammatory cytokines such as  $TNF\alpha$  and  $IFN\gamma$ , characteristic of the acute-phase reaction *in vivo*.

In accord with the above observations that N-BPs activate  $V\gamma 9V\delta 2^+$  T-cells indirectly, a recent report by Hewitt *et al.* suggests that the acute-phase reaction to N-BPs differs from a classical acute-phase response in that the  $V\gamma 9V\delta 2^+$  T-cells, rather than  $CD14^+$  monocytes and macrophages, are the primary source of the pro-inflammatory cytokines  $TNF\alpha$  and IL-6 (23). Thus, although  $CD14^+$  cells are necessary for uptake of N-BP, IPP accumulation and the subsequent presentation of IPP to  $V\gamma 9V\delta 2^+$  T-cells, the activated  $V\gamma 9V\delta 2^+$  T-cells themselves are ultimately responsible for the characteristic cytokine profile observed in the acute-phase reaction to N-BPs *in vivo*.

One poorly understood aspect of the acute-phase reaction to N-BPs is that it usually

only manifests itself following the initial infusion, with subsequent infusions failing to trigger symptoms of comparable severity. Since these subsequent infusions should attain similar plasma concentrations of N-BP *in vivo*, there appears to be some form of inherent regulatory mechanism(s) that limits further  $V\gamma 9V\delta 2^+$  T-cell activation in response to subsequent N-BP infusions. Similar *in vivo*  $V\gamma 9V\delta 2^+$  T-cell activation and subsequent failure to respond has also been observed with repeated treatments of cynomolgus monkeys with the synthetic  $V\gamma 9V\delta 2$ -TCR agonist BrHPP (28). This *in vivo* regulatory mechanism may also explain why only about one-third of patients appear to develop clinical symptoms of an acute-phase reaction, whereas the majority (90-100%) of blood samples taken from healthy

volunteers demonstrate reactive  $V\gamma 9V\delta 2^+$  T-cells when stimulated with clinically relevant concentrations of N-BPs *in vitro* (our unpublished observations and (29;30)). The mechanism responsible for limiting the further stimulatory effects of N-BPs on  $V\gamma 9V\delta 2^+$  T-cells *in vivo* remains to be clarified, but may involve regulatory T-cells ( $T_{regs}$ ), for example,  $CD4^+/CD25^{hi}/FOXP3^+$  self-reactive T-cells capable of suppressing T-cell function. Their defective regulatory nature is thought to underlie a variety of autoimmune diseases, and has been suggested to play a role in tumor growth and infection (reviewed in (31)). However, further investigation is necessary to clarify the exact role of these  $T_{regs}$  in the acute-phase reaction to N-BPs *in vivo*.

### $\gamma\delta$ T-cells in Human Health and Disease

The germline-encoded TCR repertoire of  $\gamma\delta$  T-cells is strikingly small compared to that of conventional  $\alpha\beta$  T-cells, and the usage of the few available gene segments is further restricted within different anatomical sites in the body. In the peripheral blood of healthy adults  $V\gamma 9V\delta 2^+$  T-cells are predominant (50-90% of all  $\gamma\delta$  T-cells), whilst  $V\delta 1^+$  cells co-expressing different  $V\gamma$  genes are primarily localized to epithelial surfaces such as the gut, skin, esophagus, trachea, lungs and genital epithelia (32). In the early 1990s,  $V\gamma 9V\delta 2^+$  T-cells in PBMC cultures were demonstrated to proliferate dramatically *in vitro* following exposure to extracts of *Mycobacterium tuberculosis* (33). Subsequent studies revealed that the most potent ligands for  $V\gamma 9V\delta 2^+$  T-cells are phosphorylated intermediates of the non-mevalonate (1-deoxy-D-xylulose-5-phosphate/DOXP) pathway of isoprenoid biosynthesis, which is utilized by many bacteria including *M. tuberculosis* but not eukaryotic cells (34;35). Therefore,  $\gamma\delta$  T-cells have the innate ability to recognize a diverse array of microbial pathogens containing these phosphorylated isoprenoid intermediates.

In addition to these anti-bacterial effects,  $\gamma\delta$  T-cells, and other immune cells such as NK cells, seem to play an important role in tumor surveillance, since mice deficient in these cell types show increased incidence of tumors (36;37). Whilst the study of  $V\gamma 9V\delta 2^+$

T-cells has been hampered by the lack of a comparable, analogous subset of phosphoantigen-reactive  $\gamma\delta$  T-cells in murine models, clinical evidence is mounting for a similar anti-tumor role for  $\gamma\delta$  T-cells in humans.  $\gamma\delta$  T-cells kill autologous tumor cells and have been isolated from a variety of tumors such as renal cell carcinoma (38), colorectal cancer (39) and lung carcinoma (40). *In vitro*,  $\gamma\delta$  T-cells also demonstrate broad cytotoxic activity towards many different tumour cells types (reviewed in (41)).

$V\gamma 9V\delta 2^+$  T-cells are also capable of anti-viral responses, since treatment of PBMC cultures with IPP induces the release of a wide variety of molecules with anti-viral or immunomodulatory properties such as the cytokines  $IFN\gamma$ ,  $TNF\alpha$ ,  $IL-1\alpha$ ,  $IL-6$  and GM-CSF, and chemokines such as  $MIP-1\alpha$  and  $MIP-1\beta$ , RANTES and SDF-1 (reviewed in (42)). Therefore, through the release of this diverse array of cytokines and chemokines, activation of innate  $\gamma\delta$  T-cells induces further humoral and cellular adaptive immune responses to co-ordinate a sustained anti-viral response to pathogens.

$V\gamma 9V\delta 2^+$  T-cells have been reported to proliferate in response to HIV-infected cells, exert powerful cytotoxic activity against HIV-infected targets (43), and produce HIV-inhibitory  $\beta$ -chemokines and  $\alpha$ -defensins (44;45). Due to the poorly co-ordinated immune response in HIV-infected patients, the *ex vivo* expansion of  $V\gamma 9V\delta 2^+$  T-cells with N-BPs (or synthetic agonists such as BrHPP) presents a tantalizing approach to restoring a more effective immune response in these patients.

### Potential of Bisphosphonates for Immunotherapy

Due to the potent stimulatory effects of N-BPs on  $V\gamma 9V\delta 2^+$  T-cells, this blockbuster class of drugs, developed originally for the treatment of osteoporosis, Paget's disease and metastatic bone disease, may offer a novel approach for selective activation of  $V\gamma 9V\delta 2^+$  T-cells *in vivo*. Since N-BPs are already licensed for clinical use, these drugs are suitable candidates for development as vaccines to boost innate immune responses mediated by  $V\gamma 9V\delta 2^+$  T-cells. Despite their relative scarcity in peripheral blood

(compared to conventional  $CD4^+ \alpha\beta$  T-cells),  $\gamma\delta$  T-cells can orchestrate a co-ordinated immune response to pathogens, mediated through both cytokine release and also by direct cytotoxic effects on target cells (See Figure 2). Upon activation by pamidronate,  $V\gamma9V\delta2^+$  T-cells release  $IFN\gamma$  (17;46), a pro-inflammatory cytokine with multiple anti-tumor effects, including direct inhibition of tumor cell growth, inhibition of angiogenesis and stimulation of macrophage activity (47). Thus,  $IFN\gamma$  may be a key cytokine involved in  $\gamma\delta$  T-cell-mediated anti-tumor responses. Furthermore, *in vivo* treatment of cancer patients with zoledronic acid causes  $V\gamma9V\delta2^+$  T-cells to mature towards an  $IFN\gamma$ -

producing effector phenotype, suggesting that zoledronic acid treatment may lead to more effective anti-tumor responses (48).  $V\gamma9V\delta2^+$  T-cells can recognize and lyse tumor cells pre-treated with N-BPs (21). This effect is due to the intracellular accumulation of IPP in the tumor cells, which in turn activates  $V\gamma9V\delta2^+$  T-cells.  $\gamma\delta$  T-cells have also been found to form stable conjugates with N-BP-pulsed tumor cells (49) and ZOL treatment increases sensitivity of small cell lung cancer and fibrosarcoma cell lines to lysis by  $\gamma\delta$  T-cells, an effect mediated through direct cell to cell contact and perforin-dependent cytotoxicity (50).

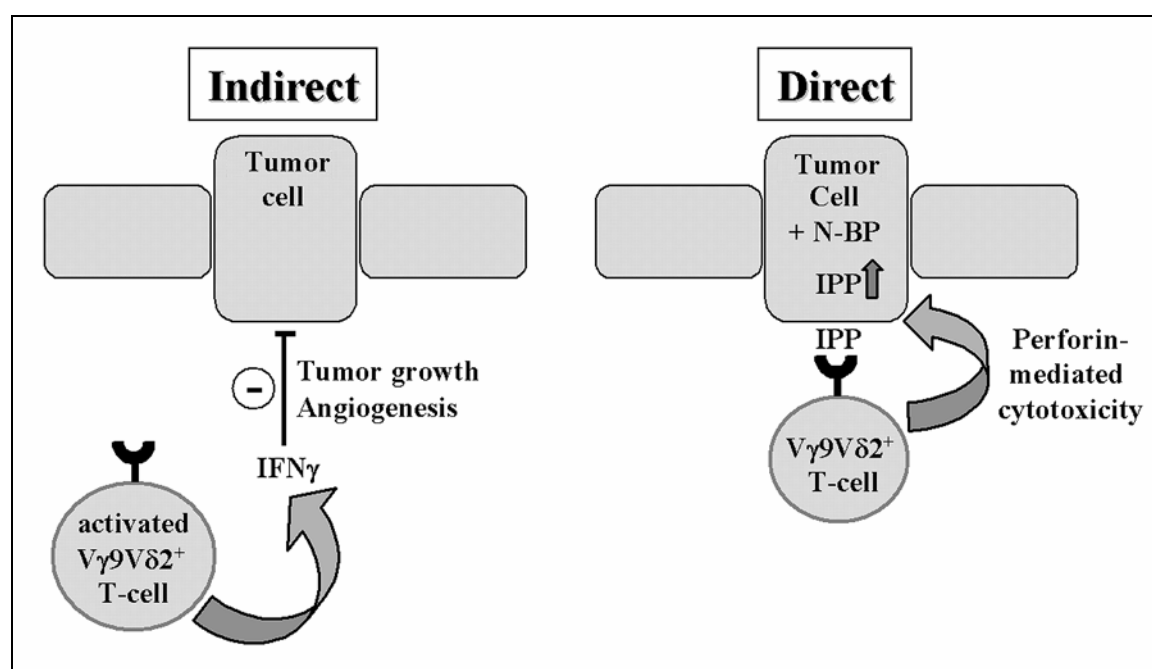


Figure 2. Proposed direct and indirect anti-tumor effects of  $V\gamma9V\delta2^+$  T-cell activation *in vivo*. Following activation,  $V\gamma9V\delta2^+$  T-cells release  $IFN\gamma$  (left panel) that is capable of inhibitory effects on tumor cell growth and angiogenesis. Activated  $V\gamma9V\delta2^+$  T-cells can also recognize IPP on the surface of N-BP-pulsed tumor cells (right panel), and have cytotoxic effects on tumor cells through the release of the cytolytic protein perforin.

The anti-lymphoma activity of  $V\gamma9V\delta2^+$  T-cells (in combination with the T-cell survival factor IL-2) has been investigated in patients with lymphoid malignancies and has demonstrated some efficacy *in vivo*, with  $V\gamma9V\delta2^+$  T-cell expansion being a prerequisite for tumour regression (29). While the evidence for N-BPs themselves having direct effects on tumor cells *in vivo* remains inconclusive, the potent stimulatory effects of N-BPs on  $V\gamma9V\delta2^+$  T-cells provides strong justification for incorporating N-BPs in future treatment regimens for lymphoid

malignancies (reviewed in (51)). The use of N-BPs in this setting will most likely require combination with factors such as IL-2, since  $\gamma\delta$  T-cells themselves do not produce IL-2 in sufficient quantities to maintain a strong proliferative response (52).

Finally, it is tempting to speculate that activation of  $\gamma\delta$  T-cells with anti-tumor activity could explain the striking anti-tumor effects of N-BPs that have recently been described in animal models (9). However, rodents do not appear to possess a

population of  $\gamma\delta$  T-cells analogous to the  $V\gamma 9V\delta 2^+$  T-cells, i.e. those activated by IPP (32), therefore the exact mechanism underlying the anti-tumor effects of N-BPs in mouse models remains to be determined.

## Conclusion

Due to the relative safety, potency and long-term stability of N-BPs such as zoledronic acid, these agents represent ideal candidates for activating  $V\gamma 9V\delta 2^+$  T-cells in the clinical setting, particularly in conditions such as lymphoid malignancies, which are often associated with osteolytic disease. However, inhibition of bone resorption would be a notable side-effect in patients lacking such metastatic bone disease and thus warrants consideration. Although the therapeutic manipulation of  $\gamma\delta$  T-cells is currently still in its infancy, it has enormous potential and could form the basis of numerous anti-bacterial, anti-viral and anti-tumor treatment strategies in the future.

**Conflict of Interest:** Dr. Thompson reports that no conflict of interest exists. Dr. Rogers reports that he receives research funding from Procter & Gamble, Novartis and Roche, and that he is a consultant for Procter & Gamble and Novartis.

## References

1. Rogers MJ. New insights into the molecular mechanisms of action of bisphosphonates. *Curr Pharm Des.* 2003;9(32):2643-58.
2. Thompson K, Rogers MJ, Coxon FP, Crockett JC. Cytosolic entry of bisphosphonate drugs requires acidification of vesicles after fluid-phase endocytosis. *Mol Pharmacol.* 2006 May;69(5):1624-32.
3. Coxon FP, Thompson K, Ebetino FH, Rogers MJ. Resorbing osteoclasts increase the availability of mineral-bound bisphosphonates to non-resorbing cells. *Bone.* 2006 Mar;38(Suppl 1):S45-S46.
4. Frith JC, Monkkonen J, Auriola S, Monkkonen H, Rogers MJ. The molecular mechanism of action of the anti-resorptive and anti-inflammatory drug clodronate: evidence for the formation in vivo of a metabolite that inhibits bone resorption and causes osteoclast and macrophage apoptosis. *Arthritis Rheum.* 2001 Sep;44(9):2201-10.
5. Kavanagh KL, Guo K, Dunford JE, Wu X, Knapp S, Ebetino FH, Rogers MJ, Russell RG, Oppermann U. The molecular mechanism of nitrogen-containing bisphosphonates as antiosteoporosis drugs. *Proc Natl Acad Sci U S A.* 2006 May 16;103(20):7829-34.
6. Rondeau JM, Bitsch F, Bourcier E, Geiser M, Hemmig R, Kroemer M, Lehmann S, Ramage P, Rieffel S, Strauss A, Green JR, Jahnke W. Structural basis for the exceptional in vivo efficacy of bisphosphonate drugs. *ChemMedChem.* 2006 Feb 13;1(2):267-73.
7. Dunford JE, Thompson K, Coxon FP, Luckman SP, Hahn FM, Poulter CD, Ebetino FH, Rogers MJ. Structure-activity relationships for inhibition of farnesyl diphosphate synthase *in vitro* and inhibition of bone resorption *in vivo* by nitrogen-containing bisphosphonates. *J Pharmacol Exp Ther.* 2001 Feb;296(2):235-42.
8. Dunford JE, Rogers MJ, Ebetino FH, Phipps RJ, Coxon FP. Inhibition of protein prenylation by bisphosphonates causes sustained activation of Rac, Cdc42, and Rho GTPases. *J Bone Miner Res.* 2006 May;21(5):684-94.
9. Green JR. Bisphosphonates: preclinical review. *Oncologist.* 2004;9(Suppl 4):3-13.
10. Reid IR, Brown JP, Burckhardt P, Horowitz Z, Richardson P, Trechsel U, Widmer A, Devogelaer JP, Kaufman JM, Jaeger P, Body JJ, Brandi ML, Broell J, Di Micco R, Genazzani AR, Felsenberg D, Happ J, Hooper MJ, Ittner J, Leeb G, Mallmin H, Murray T, Ortolani S, Rubinacci A, Saaf M, Samsioe G, Verbruggen L, Meunier PJ. Intravenous zoledronic acid in postmenopausal women with low bone mineral density. *N Engl J Med.* 2002 Feb 28;346(9):653-61.

11. Delmas PD, Adami S, Strugala C, Stakkestad JA, Reginster JY, Felsenberg D, Christiansen C, Civitelli R, Drezner MK, Recker RR, Bolognese M, Hughes C, Masanaukaite D, Ward P, Sambrook P, Reid DM. Intravenous ibandronate injections in postmenopausal women with osteoporosis: one-year results from the dosing intravenous administration study. *Arthritis Rheum.* 2006 Jun;54(6):1838-46.
12. Adami S, Bhalla AK, Dorizzi R, Montesanti F, Rosini S, Salvagno G, Lo Cascio V. The acute-phase response after bisphosphonate administration. *Calcif Tissue Int.* 1987 Dec;41(6):326-31.
13. Kunzmann V, Bauer E, Wilhelm M. Gamma/delta T-cell stimulation by pamidronate. *N Engl J Med.* 1999 Mar 4;340(9):737-8.
14. Tanaka Y, Morita CT, Tanaka Y, Nieves E, Brenner MB, Bloom BR. Natural and synthetic non-peptide antigens recognized by human gamma delta T cells. *Nature.* 1995 May 11;375(6527):155-8.
15. Espinosa E, Belmont C, Pont F, Luciani B, Poupot R, Romagne F, Brailly H, Bonneville M, Fournie JJ. Chemical synthesis and biological activity of bromohydrin pyrophosphate, a potent stimulator of human gamma delta T cells. *J Biol Chem.* 2001 May 25;276(21):18337-44.
16. Bukowski JF, Morita CT, Brenner MB. Human gamma delta T cells recognize alkylamines derived from microbes, edible plants, and tea: implications for innate immunity. *Immunity.* 1999 Jul;11(1):57-65.
17. Kunzmann V, Bauer E, Feurle J, Weissinger F, Tony HP, Wilhelm M. Stimulation of gammadelta T cells by aminobisphosphonates and induction of antiplasma cell activity in multiple myeloma. *Blood.* 2000 Jul 15;96(2):384-92.
18. Martin MB, Arnold W, Heath HT 3rd, Urbina JA, Oldfield E. Nitrogen-containing bisphosphonates as carbocation transition state analogs for isoprenoid biosynthesis. *Biochem Biophys Res Commun.* 1999 Oct 5;263(3):754-8.
19. Chen T, Berenson J, Vescio R, Swift R, Gilchick A, Goodin S, LoRusso P, Ma P, Ravera C, Deckert F, Schran H, Seaman J, Skerjanec A. Pharmacokinetics and pharmacodynamics of zoledronic acid in cancer patients with bone metastases. *J Clin Pharmacol.* 2002 Nov;42(11):1228-36.
20. Kunzmann V, Engert J, Reimer P, Weissinger F, Herrmann T, Wilhelm M. Crucial role of monocytes in aminobisphosphonate recognition by gamma delta T cells. *Blood.* 2003 Nov 16;102(11):39B.
21. Gober HJ, Kistowska M, Angman L, Jeno P, Mori L, De Libero G. Human T cell receptor gammadelta cells recognize endogenous mevalonate metabolites in tumor cells. *J Exp Med.* 2003 Jan 20;197(2):163-8.
22. Thompson K, Rogers MJ. Statins prevent bisphosphonate-induced gamma,delta-T-cell proliferation and activation in vitro. *J Bone Miner Res.* 2004 Feb;19(2):278-88.
23. Hewitt RE, Lissina A, Green AE, Slay ES, Price DA, Sewell AK. The bisphosphonate acute phase response: rapid and copious production of proinflammatory cytokines by peripheral blood gd T cells in response to aminobisphosphonates is inhibited by statins. *Clin Exp Immunol.* 2005 Jan;139(1):101-11.
24. Schweitzer DH, Oostendorp-van de Ruit M, Van der Pluijm G, Lowik CW, Papapoulos SE. Interleukin-6 and the acute phase response during treatment of patients with Paget's disease with the nitrogen-containing bisphosphonate dimethylamino-hydroxypropylidene bisphosphonate. *J Bone Miner Res.* 1995 Jun;10(6):956-62.
25. Sauty A, Pecherstorfer M, Zimmer-Roth I, Fioroni P, Juillerat L, Markert M,

- Ludwig H, Leuenberger P, Burckhardt P, Thiebaud D. Interleukin-6 and tumor necrosis factor alpha levels after bisphosphonates treatment in vitro and in patients with malignancy. *Bone*. 1996 Feb;18(2):133-9.
26. Thiebaud D, Sauty A, Burckhardt P, Leuenberger P, Sitzler L, Green JR, Kandra A, Zieschang J, Ibarra de Palacios P. An in vitro and in vivo study of cytokines in the acute-phase response associated with bisphosphonates. *Calcif Tissue Int*. 1997 Nov;61(5):386-92.
27. Thompson K, Rojas-Navea J, Rogers MJ. Alkylamines cause Vgamma9Vdelta2 T-cell activation and proliferation by inhibiting the mevalonate pathway. *Blood*. 2006 Jan 15;107(2):651-4.
28. Sicard H, Ingoure S, Luciani B, Serraz C, Fournie JJ, Bonneville M, Tiollier J, Romagne F. In vivo immunomanipulation of V gamma 9V delta 2 T cells with a synthetic phosphoantigen in a preclinical nonhuman primate model. *J Immunol*. 2005 Oct 15;175(8):5471-80.
29. Wilhelm M, Kunzmann V, Eckstein S, Reimer P, Weissinger F, Ruediger T, Tony HP. Gammadelta T cells for immune therapy of patients with lymphoid malignancies. *Blood*. 2003 Jul 1;102(1):200-6.
30. Mariani S, Muraro M, Pantaleoni F, Fiore F, Nuschak B, Peola S, Foglietta M, Palumbo A, Coscia M, Castella B, Bruno B, Bertieri R, Boano L, Boccadoro M, Massaia M. Effector gammadelta T cells and tumor cells as immune targets of zoledronic acid in multiple myeloma. *Leukemia*. 2005 Apr;19(4):664-70.
31. Kronenberg M, Rudensky A. Regulation of immunity by self-reactive T cells. *Nature*. 2005 Jun 2;435(7042):598-604.
32. Hayday AC. [gamma][delta] cells: a right time and a right place for a conserved third way of protection. *Annu Rev Immunol*. 2000;18:975-1026.
33. Kabelitz D, Bender A, Prospero T, Wesselborg S, Janssen O, Pechhold K. The primary response of human gamma/delta + T cells to Mycobacterium tuberculosis is restricted to V gamma 9-bearing cells. *J Exp Med*. 1991 Jun 1;173(6):1331-8.
34. Jomaa H, Feurle J, Luhs K, Kunzmann V, Tony HP, Herderich M, Wilhelm M. Vgamma9/Vdelta2 T cell activation induced by bacterial low molecular mass compounds depends on the 1-deoxy-D-xylulose 5-phosphate pathway of isoprenoid biosynthesis. *FEMS Immunol Med Microbiol*. 1999 Sep;25(4):371-8.
35. Belmont C, Espinosa E, Poupot R, Peyrat MA, Guiraud M, Poquet Y, Bonneville M, Fournie JJ. 3-Formyl-1-butyl pyrophosphate A novel mycobacterial metabolite-activating human gammadelta T cells. *J Biol Chem*. 1999 Nov 5;274(45):32079-84.
36. Smyth MJ, Thia KY, Street SE, Cretney E, Trapani JA, Taniguchi M, Kawano T, Pelikan SB, Crowe NY, Godfrey DI. Differential tumor surveillance by natural killer (NK) and NKT cells. *J Exp Med*. 2000 Feb 21;191(4):661-8.
37. Girardi M, Oppenheim DE, Steele CR, Lewis JM, Glusac E, Filler R, Hobby P, Sutton B, Tigelaar RE, Hayday AC. Regulation of cutaneous malignancy by gammadelta T cells. *Science*. 2001 Oct 19;294(5542):605-9.
38. Choudhary A, Davodeau F, Moreau A, Peyrat MA, Bonneville M, Jotereau F. Selective lysis of autologous tumor cells by recurrent gamma delta tumor-infiltrating lymphocytes from renal carcinoma. *J Immunol*. 1995 Apr 15;154(8):3932-40.
39. Maeurer MJ, Martin D, Walter W, Liu K, Zitvogel L, Halusczyk K, Rabinowich H, Duquesnoy R, Storkus W, Lotze MT. Human intestinal Vdelta1+ lymphocytes recognize tumor cells of epithelial origin. *J Exp Med*. 1996 Apr 1;183(4):1681-96.
40. Ferrarini M, Heltai S, Pupa SM, Mernard S, Zocchi R. Killing of laminin receptor-positive human lung cancers by tumor infiltrating lymphocytes bearing



- gammadelta(+) t-cell receptors. *J Natl Cancer Inst.* 1996 Apr 3;88(7):436-41.
41. Ferrarini M, Ferrero E, Dagna L, Poggi A, Zocchi MR. Human gammadelta T cells: a nonredundant system in the immune-surveillance against cancer. *Trends Immunol.* 2002 Jan;23(1):14-8.
  42. Poccia F, Agrati C, Martini F, Mejia G, Wallace M, Malkovsky M. Vgamma9Vdelta2 T cell-mediated non-cytolytic antiviral mechanisms and their potential for cell-based therapy. *Immunol Lett.* 2005 Aug 15;100(1):14-20.
  43. Poccia F, Cipriani B, Vendetti S, Colizzi V, Poquet Y, Battistini L, Lopez-Botet M, Fournie JJ, Gougeon ML. CD94/NKG2 inhibitory receptor complex modulates both anti-viral and anti-tumoral responses of polyclonal phosphoantigen-reactive V gamma 9V delta 2 T lymphocytes. *J Immunol.* 1997 Dec 15;159(12):6009-17.
  44. Poccia F, Battistini L, Cipriani B, Mancino G, Martini F, Gougeon ML, Colizzi V. Phosphoantigen-reactive Vgamma9Vdelta2 T lymphocytes suppress in vitro human immunodeficiency virus type 1 replication by cell-released antiviral factors including CC chemokines. *J Infect Dis.* 1999 Sep;180(3):858-61.
  45. Agerberth B, Charo J, Werr J, Olsson B, Idali F, Lindbom L, Kiessling R, Jornvall H, Wigzell H, Gudmundsson GH. The human antimicrobial and chemotactic peptides LL-37 and alpha-defensins are expressed by specific lymphocyte and monocyte populations. *Blood.* 2000 Nov 1;96(9):3086-93.
  46. Miyagawa F, Tanaka Y, Yamashita S, Minato N. Essential requirement of antigen presentation by monocyte lineage cells for the activation of primary human gamma delta T cells by aminobisphosphonate antigen. *J Immunol.* 2001 May 1;166(9):5508-14.
  47. Boehm U, Klamp T, Groot M, Howard JC. Cellular responses to interferon-gamma. *Annu Rev Immunol.* 1997;15:749-95.
  48. Dieli F, Gebbia N, Poccia F, Caccamo N, Montesano C, Fulfarò F, Arcara C, Valerio MR, Meraviglia S, Di Sano C, Sireci G, Salerno A. Induction of gammadelta T-lymphocyte effector functions by bisphosphonate zoledronic acid in cancer patients in vivo. *Blood.* 2003 Sep 15;102(6):2310-1.
  49. Kato Y, Tanaka Y, Tanaka H, Yamashita S, Minato N. Requirement of species-specific interactions for the activation of human gamma delta T cells by pamidronate. *J Immunol.* 2003 Apr 1;170(7):3608-13.
  50. Sato K, Kimura S, Segawa H, Yokota A, Matsumoto S, Kuroda J, Nogawa M, Yuasa T, Kiyono Y, Wada H, Maekawa T. Cytotoxic effects of gammadelta T cells expanded ex vivo by a third generation bisphosphonate for cancer immunotherapy. *Int J Cancer.* 2005 Aug 10;116(1):94-9.
  51. Kunzmann V, Wilhelm M. Anti-lymphoma effect of gammadelta T cells. *Leuk Lymphoma.* 2005 May;46(5):671-80.
  52. Pechhold K, Wesch D, Schondelmaier S, Kabelitz D. Primary activation of V gamma 9-expressing gamma delta T cells by Mycobacterium tuberculosis. Requirement for Th1-type CD4 T cell help and inhibition by IL-10. *J Immunol.* 1994 May 15;152(10):4984-92.