

COMMENTARIES

Take Two Aspirin: for Osteoporosis?

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Commentary on: Yamaza T, Miura Y, Bi Y, Liu Y, Akiyama K, Sonoyama W, Patal V, Gutkind S, Young M, Gronthos S, Le A, Wang CY, Chen W, Shi S. Pharmacologic stem cell based intervention as a new approach to osteoporosis treatment in rodents. *PLoS ONE*. 2008 Jul 9;3(7):e2615

Post-menopausal osteoporosis is characterized by increased bone turnover, with bone resorption exceeding formation, leading to loss of bone mass. Estrogen deficiency in rodents is associated with increased expression of inflammatory cytokines by lymphocytes, which is likely to contribute to bone loss. The objectives of the study by Yamaza *et al.* (1) were to investigate the role of inflammatory T cells in bone marrow mesenchymal stem cell apoptosis and in ovariectomy (OVX)-induced bone loss in mice and to examine whether aspirin could prevent bone loss and the effects of activated T cells.

This complex study comes from 5 different research centers in the United States, Japan and Australia and has two provocative messages that deserve further investigation (1). The first is an extension of prior studies on the role of T cells in the pathogenesis of osteoporosis (2), while the second is a new finding that pretreatment with aspirin may mitigate bone loss in OVX mice by a T-cell dependent mechanism. Prior studies of the role of lymphocytes in the pathogenesis of post-menopausal osteoporosis have largely focused on bone resorption (2). The first part of the manuscript describes experiments with activated T cells co-cultured with bone marrow mesenchymal stem cells (BMMSCs). Activated T cells are shown to increase apoptosis of BMMSCs through the Fas/FasL pathway, suggesting that activated T cells may decrease bone formation. The

authors also examined the effect of OVX on bone mass of immunocompromised (bg-nu/nu-xid) mice that do not have T cells. They show that these mice do not lose bone mass with OVX as measured by DXA. However, DXA predominantly measures changes in cortical bone mass, and it was previously reported that T-cell deficient mice maintain their cortical bone mass (3). There are also conflicting reports on trabecular bone loss after OVX in T-cell deficient mice, perhaps due to different experimental models (3;4). In the study by Yamaza *et al.* (1) the inflammatory T-cells increased osteoclasts associated with trabeculae, suggesting that there may be loss of trabecular bone, but this loss should be confirmed with μ CT or histomorphometric studies.

The authors then examined the effects on bone mass in OVX immunocompromised mice of adding back a population of pro-inflammatory T cells ($CD4^+CD25^-CD45RB^{+hi}$), which were previously shown to induce inflammatory bowel disease in immunocompromised mice. The $CD4^+CD25^-CD45RB^{+hi}$ T cells caused bone loss in OVX immunocompromised mice while the reciprocal $CD4^+CD25^-CD45RB^{-/low}$ T cells did not. It is likely that the bone loss induced in the OVX immunocompromised mice after T cell transfer resulted from production of cytokines and cellular immune mediators of inflammatory disease in these mice. Since the authors did not examine the effects of

transferring CD4⁺CD25⁻CD45RB⁺ T cells into sham-operated mice, it is unclear if the role of these inflammatory T cells is specific for bone loss after OVX. It is possible that sham-operated mice would lose equal amounts of bone after transfer of CD4⁺CD25⁻CD45RB⁺ T cells.

The second series of experiments with aspirin are novel. OVX mice were pretreated for 2 months with aspirin before surgery and continued on aspirin for another month after OVX. These aspirin-treated OVX mice were compared with sham-operated and OVX mice without any aspirin treatment. Aspirin-treated OVX mice had less trabecular bone loss, measured by μ CT, compared to OVX only mice. However, since there were no sham-operated controls treated with aspirin, it is not clear whether the effects observed with aspirin treatment are specific to OVX. It is possible that aspirin has effects in sham-operated mice.

BMMSCs were isolated from OVX mice and aspirin-treated OVX mice and implanted into immunocompromised mice. Transplantation into immunocompromised mice of BMMSCs from aspirin-treated OVX mice formed more bone than BMMSCs from OVX mice. The authors demonstrated that aspirin can inhibit Fas-stimulated death of human BMMSCs and decrease viability of activated T cells. Using cultured human BMMSCs, they showed that aspirin can increase *in vitro* mineralization and slightly increase telomerase activity and telomere length. Human BMMSCs expanded *in vitro* and treated with aspirin before being transplanted into immunocompromised mice formed more bone compared with BMMSCs not treated with aspirin.

In addition to the effects on osteoblastic cells, there were fewer osteoclastic cells in OVX mice treated with aspirin compared with OVX alone. The decrease in osteoclastic cells in aspirin-treated OVX mice was associated with decreased serum receptor activator of nuclear factor κ B ligand (RANKL), a required factor for osteoclast formation, and increased osteoprotegerin (OPG), an inhibitor of RANKL, compared

with OVX alone. Again, it is not clear if the effects of aspirin are specific for OVX-induced changes in osteoclasts or serum RANKL and OPG because sham-operated mice were not also given aspirin. Osteoclast formation can be stimulated *in vitro* by treating osteoclastic precursors, isolated from bone marrow or spleens, with macrophage-colony stimulating factor (M-CSF) and RANKL or by stimulating production of these factors with hormones or cytokines in osteoblastic cells. 1,25-OH vitamin D stimulated osteoclastic cell formation in co-cultures of primary osteoblasts with either bone marrow cells or spleen cells, which was inhibited by aspirin treatment. Aspirin treatment also decreased osteoclastic cell formation in spleen cells treated with M-CSF and RANKL. These results suggest that aspirin may decrease osteoclastogenesis both by regulating osteoblastic support of osteoclast formation and by inhibiting osteoclast formation directly.

This work suggests the potential of aspirin as an anti-osteoporosis drug, but questions remain. While the authors did show possible effects of aspirin on cell survival and BMMSC differentiation and telomere length, the direct mechanism of aspirin action was not identified. One of the major effects of aspirin is to inhibit prostaglandin (PG) production. Aspirin is an acetylated salicylate and acetylates a critical serine in the arachidonic acid binding site of cyclooxygenase (COX) 1 and 2, inhibiting the synthesis of PGs (5;6). Unlike other COX inhibitors (non-steroidal anti-inflammatory drugs, NSAIDs), this reaction produces irreversible inhibition of the enzymes. The role of PGs on bone maintenance is complicated, having effects on both bone formation and resorption (7). The *in vitro* effects of aspirin to inhibit osteoclast formation are similar to those seen with deficiency or inhibition of COX-2 (8). However, the *in vitro* effects of aspirin to stimulate osteoblastic mineralization are the opposite of the effects of COX-2 deficiency or inhibition (9). Exogenous PGE₂ increases both bone formation and bone resorption *in vivo* (10-15). Whether there is a net gain or

loss of bone appears to depend on the experimental model (16;17). Genetic deletion of COX-2, which is the predominant source of PGs in bone, results in a complex phenotype, including delayed fracture healing and a high bone turnover state associated with high serum parathyroid hormone (PTH) (18;19). Preliminary results from our laboratory have suggested that there is a greater loss in bone mass following OVX of COX-2 knockout mice, compared with wild type mice, however, these results may be confounded by high serum PTH in the knockout mice (20). Thus, if aspirin is increasing osteogenesis and decreasing resorption, these effects are likely to be not simply due to the inhibition of COX.

Aspirin and other salicylates have been described as having diverse actions, independent of effects on cyclooxygenase (21;22). These actions of salicylates may limit bone loss directly by acting on bone cells or indirectly by limiting inflammation or oxidative stress. Salicylates may inhibit nuclear factor κ B, which is required for osteoclast formation stimulated by RANKL (23;24). Inhibition of nuclear factor κ B by aspirin could contribute to the decreased osteoclast cells observed following aspirin treatment. Aspirin may also act by inhibiting cell adhesion and reducing oxidative stress (25-27). On the other hand, inhibitory actions of salicylates on the function of inflammatory cells could also prevent bone loss (21;22). Activated lymphocytes secrete pro-inflammatory cytokines that decrease osteoblastic function, while enhancing osteoclast formation, resulting in decreased bone mass (28). Thus, it is possible that the pleiotropic actions of aspirin may increase formation and decrease resorption of bone independently of the effects of aspirin to inhibit PGs. Use of non-acetylated salicylates and COX-deficient mouse models will help to clarify the mechanism of aspirin in limiting bone loss associated with estrogen deficiency.

Human epidemiologic studies suggest an association of aspirin with increased bone mineral density (BMD), although this is not

consistently observed (29-32). A recent cross-sectional study of community-dwelling men and women older than 65 years assessed the association of aspirin and COX-2 selective inhibitors on BMD (32). In this study, men taking COX-2 inhibitors had lower BMD compared with men not taking any NSAID. Men using aspirin in addition to COX-2 inhibitors had a further reduction of BMD, compared with COX-2 inhibitor use alone. On the other hand, COX-2 inhibitors were associated with higher BMD in women not taking estrogen. Aspirin and COX-2 inhibitor use was associated with higher BMD than COX-2 inhibitor use alone in these women. Since low dose aspirin is widely used and relatively safe, prospective studies of its effects on the rapid bone loss that occurs at menopause, or even earlier during perimenopause, would be feasible and of great clinical interest.

Conflict of Interest: None reported.

Peer Review: This article has been peer-reviewed.

References

1. Yamaza T, Miura Y, Bi Y, Liu Y, Akiyama K, Sonoyama W, Patel V, Gutkind S, Young M, Gronthos S, Le A, Wang CY, Chen W, Shi S. Pharmacologic stem cell based intervention as a new approach to osteoporosis treatment in rodents. *PLoS ONE*. 2008 Jul 9;3(7):e2615.
2. Weitzmann MN, Pacifici R. The role of T lymphocytes in bone metabolism. *Immunol Rev*. 2005 Dec;208:154-68.
3. Lee SK, Kadono Y, Okada F, Jacquin C, Koczon-Jaremko B, Gronowicz G, Adams DJ, Aguila HL, Choi Y, Lorenzo JA. T lymphocyte-deficient mice lose trabecular bone mass with ovariectomy. *J Bone Miner Res*. 2006 Nov;21(11):1704-12.
4. Gao Y, Qian WP, Dark K, Toraldo G, Lin AS, Guldberg RE, Flavell RA, Weitzmann MN, Pacifici R. Estrogen prevents bone loss through transforming

- growth factor beta signaling in T cells. *Proc Natl Acad Sci U S A*. 2004 Nov 23;101(47):16618-23.
5. Roth GJ, Stanford N, Majerus PW. Acetylation of prostaglandin synthase by aspirin. *Proc Natl Acad Sci U S A*. 1975 Aug;72(8):3073-6.
 6. DeWitt DL, el-Harith EA, Kraemer SA, Andrews MJ, Yao EF, Armstrong RL, Smith WL. The aspirin and heme-binding sites of ovine and murine prostaglandin endoperoxide synthases. *J Biol Chem*. 1990 Mar 25;265(9):5192-8.
 7. Pilbeam C, Choudhary S, Blackwell K, Raisz L. Prostaglandins and bone metabolism. In: Bilezikian JP, Raisz LG, Martin TJ, eds. *Principles of Bone Biology*. 3rd ed. San Diego: Elsevier/Academic Press; 2008:1235-71.
 8. Okada Y, Lorenzo JA, Freeman AM, Tomita M, Morham SG, Raisz LG, Pilbeam CC. Prostaglandin G/H synthase-2 is required for maximal formation of osteoclast-like cells in culture. *J Clin Invest*. 2000 Mar;105(6):823-32.
 9. Choudhary S, Halbout P, Alander C, Raisz L, Pilbeam C. Strontium ranelate promotes osteoblastic differentiation and mineralization of murine bone marrow stromal cells: involvement of prostaglandins. *J Bone Miner Res*. 2007 Jul;22(7):1002-10.
 10. Jee WS, Ma YF. The in vivo anabolic actions of prostaglandins in bone. *Bone*. 1997 Oct;21(4):297-304.
 11. Tang LY, Cullen DM, Yee JA, Jee WS, Kimmel DB. Prostaglandin E2 increases the skeletal response to mechanical loading. *J Bone Miner Res*. 1997 Feb;12(2):276-82.
 12. Norrdin RW, Shih MS. Systemic effects of prostaglandin E2 on vertebral trabecular remodeling in beagles used in a healing study. *Calcif Tissue Int*. 1988 Jun;42(6):363-8.
 13. Suponitzky I, Weinreb M. Differential effects of systemic prostaglandin E2 on bone mass in rat long bones and calvariae. *J Endocrinol*. 1998 Jan;156(1):51-7.
 14. Faye-Petersen OM, Johnson WH Jr, Carlo WA, Hedlund GL, Pacifico AD, Blair HC. Prostaglandin E1-induced hyperostosis: clinicopathologic correlations and possible pathogenetic mechanisms. *Pediatr Pathol Lab Med*. 1996 May-Jun;16(3):489-507.
 15. Velaphi S, Cilliers A, Beckh-Arnold E, Mokhachane M, Mphahlele R, Pettifor J. Cortical hyperostosis in an infant on prolonged prostaglandin infusion: case report and literature review. *J Perinatol*. 2004 Apr;24(4):263-5.
 16. Tian XY, Zhang Q, Zhao R, Setterberg RB, Zeng QQ, Iturria SJ, Ma YF, Jee WS. Continuous PGE2 leads to net bone loss while intermittent PGE2 leads to net bone gain in lumbar vertebral bodies of adult female rats. *Bone*. 2008 May;42(5):914-20.
 17. Gao Q, Xu M, Zhan P, Alander CB, Pilbeam CC, Raisz LG. Demonstration of an anabolic effect of prostaglandin E2 on bone in CD-1 mice. *J Bone Miner Res*. 2007 Sep;22(Suppl 1):S167.
 18. Xu M, Choudhary S, Goltzman D, Ledgard F, Adams D, Gronowicz G, Koczon-Jaremko B, Raisz L, Pilbeam C. Do cyclooxygenase-2 knockout mice have primary hyperparathyroidism? *Endocrinology*. 2005 Apr;146(4):1843-53.
 19. Zhang X, Schwarz EM, Young DA, Puzas JE, Rosier RN, O'Keefe RJ. Cyclooxygenase-2 regulates mesenchymal cell differentiation into the osteoblast lineage and is critically

- involved in bone repair. *J Clin Invest*. 2002 Jun;109(11):1405-15.
20. Xu M, Gao Q, Voznesensky OS, Choudhary S, Adams DJ, Raisz LG, Pilbeam CC. Cyclooxygenase-2 knockout does not prevent bone loss after ovariectomy. *J Bone Miner Res*. 2006 Sep;21(Suppl 1):S413.
21. Tegeder I, Pfeilschifter J, Geisslinger G. Cyclooxygenase-independent actions of cyclooxygenase inhibitors. *FASEB J*. 2001 Oct;15(12):2057-72.
22. Amann R, Peskar BA. Anti-inflammatory effects of aspirin and sodium salicylate. *Eur J Pharmacol*. 2002 Jun 28;447(1):1-9.
23. Franzoso G, Carlson L, Xing L, Poljak L, Shores EW, Brown KD, Leonardi A, Tran T, Boyce BF, Siebenlist U. Requirement for NF-kappaB in osteoclast and B-cell development. *Genes Dev*. 1997 Dec 15;11(24):3482-96.
24. Iotsova V, Caamaño J, Loy J, Yang Y, Lewin A, Bravo R. Osteopetrosis in mice lacking NF-kappaB1 and NF-kappaB2. *Nat Med*. 1997 Nov;3(11):1285-9.
25. Pillinger MH, Capodici C, Rosenthal P, Kheterpal N, Hanft S, Philips MR, Weissmann G. Modes of action of aspirin-like drugs: salicylates inhibit erk activation and integrin-dependent neutrophil adhesion. *Proc Natl Acad Sci U S A*. 1998 Nov 24;95(24):14540-5.
26. Ristimäe T, Zilmer M, Zilmer K, Kairane C, Kullisaar T, Teesalu R. Effect of low-dose aspirin on the markers of oxidative stress. *Cardiovasc Drugs Ther*. 1999 Nov;13(6):485-90.
27. Wu R, Lamontagne D, de Champlain J. Antioxidative properties of acetylsalicylic acid on vascular tissues from normotensive and spontaneously hypertensive rats. *Circulation*. 2002 Jan 22;105(3):387-92.
28. Lorenzo J, Horowitz M, Choi Y. Osteoimmunology: interactions of the bone and immune system. *Endocr Rev*. 2008 Jun;29(4):403-40.
29. Bauer DC, Orwoll ES, Fox KM, Vogt TM, Lane NE, Hochberg MC, Stone K, Nevitt MC. Aspirin and NSAID use in older women: effect on bone mineral density and fracture risk. Study of Osteoporotic Fractures Research Group. *J Bone Miner Res*. 1996 Jan;11(1):29-35.
30. Carbone LD, Tylavsky FA, Cauley JA, Harris TB, Lang TF, Bauer DC, Barrow KD, Kritchevsky SB. Association between bone mineral density and the use of nonsteroidal anti-inflammatory drugs and aspirin: impact of cyclooxygenase selectivity. *J Bone Miner Res*. 2003 Oct;18(10):1795-802.
31. Lane NE, Bauer DC, Nevitt MC, Pressman AR, Cummings SR. Aspirin and nonsteroidal antiinflammatory drug use in elderly women: effects on a marker of bone resorption. The Study of Osteoporotic Fractures Research Group. *J Rheumatol*. 1997 Jun;24(6):1132-6.
32. Richards JB, Joseph L, Schwartzman K, Kreiger N, Tenenhouse A, Goltzman D; Canadian Multicentre Osteoporosis Study Group. The effect of cyclooxygenase-2 inhibitors on bone mineral density: results from the Canadian Multicentre Osteoporosis Study. *Osteoporos Int*. 2006;17(9):1410-9.