

HETEROCYCLES, Vol. 61, 2003, pp. 189 - 196

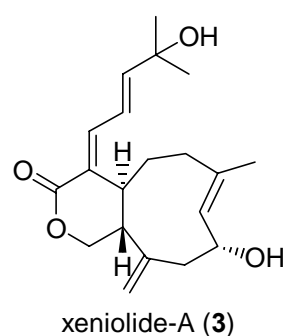
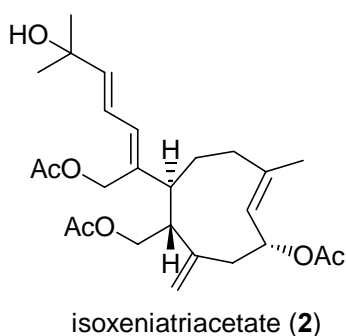
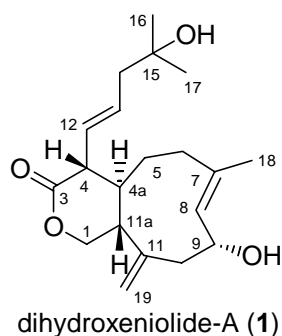
Received, 16th May, 2003, Accepted, 19th June, 2003, Published online, 24th June, 2003

TWO NEW XENICANE DITERPENOIDS FROM OKINAWAN SOFT CORAL OF THE GENUS, *XENIA*Hiroaki Miyaoka, Masakazu Nakano, Kazuo Iguchi,¹ and Yasuji Yamada*School of Pharmacy, Tokyo University of Pharmacy and Life Science,
1432-1 Horinouchi, Hachioji, Tokyo 192-0392, Japan

E-mail: yamaday@ps.toyaku.ac.jp

Abstract - Two new xenicane diterpenoids, dihydroxeniolide-A and isoxeniatriacetate, were isolated from Okinawan soft coral of the genus, *Xenia*. The relative configurations of two new xenicane diterpenoids were determined based on spectroscopic analysis. The absolute configuration of dihydroxeniolide-A was determined using the modified Mosher's method. The absolute configuration of isoxeniatriacetate was determined by chemical conversion.

Soft coral of the genus *Xenia* is a rich source of xenicane diterpenoids which possess unique structural features and biological activity.^{2,3} The isolation and structural determinations of xenicane diterpenoids from the Okinawan soft coral *Xenia* sp. were reported in our previous papers.^{4,5} Subsequent study on chemical constituents of Okinawan soft coral led to the isolation of new xenicane diterpenoids, dihydroxeniolide-A (**1**) and isoxeniatriacetate (**2**), from a soft coral of the genus of *Xenia* along with xeniolide-A⁶ (**3**), xeniolide-B,⁶ and xeniolactol.^{7,8} In the following, the isolation and structural determinations of dihydroxeniolide-A (**1**) and isoxeniatriacetate (**2**) are discussed.



Dihydroxeniolide-A (**1**) was found to have the molecular formula C₂₀H₃₀O₄ based on HREIMS. The IR spectrum of **1** indicated absorptions due to a hydroxy group (3500 cm⁻¹) and δ -lactone (1735 cm⁻¹). All twenty carbons appeared in the ¹³C NMR and DEPT spectra disclosed the presence of three methyls, five sp³ methylenes, one sp² methylene, four sp³ methines, three sp² methines, one sp³ quaternary carbon and three sp² quaternary carbons (Table 1). ¹H and ¹³C NMR spectral correlations were demonstrated by the HMQC spectrum. ¹H and ¹³C NMR spectra showed the presence of one *E*-disubstituted olefin [δ_{H} 5.53 (1H, d, *J* = 7.5 Hz), δ_{C} 130.7 (CH), 132.8 (C)], one *exo* methylene [δ_{H} 4.88 (1H, br s), 5.02 (1H, br s), δ_{C} 115.6 (CH₂), 148.7 (C)], one olefinic methyl [δ_{H} 1.68 (3H, s), δ_{C} 17.3 (CH₃)], two methyls [δ_{H} 1.21 (3H, s), 1.24 (3H, s), δ_{C} 29.0 (CH₃), 29.4 (CH₃)], one oxygenated quaternary carbon [δ_{C} 70.5 (C)], one oxygenated methine adjacent to trisubstituted olefin [δ_{H} 4.72 (1H, br t, *J* = 6.6 Hz), 5.18 (1H, d, *J* = 7.5

Table 1. NMR spectral data for dihydroxeniolide-A (**1**) and isoxeniatricetate(**2**).

| No. | 1 | | 2 | |
|------|----------------------------------|---|--------------------------------------|---|
| | ¹³ C NMR ^a | ¹ H NMR (<i>J</i> in Hz) ^b | ¹³ C NMR ^c | ¹ H NMR (<i>J</i> in Hz) ^d |
| 1 | 67.7 (CH ₂) | 4.06 (dd, 11.5, 2.8) 4.19 (dd, 11.5, 3.9) | 64.9 (CH ₂) | 3.80 (dd, 11.5, 10.4) 4.04 (dd, 11.5, 4.9) |
| 3 | 174.2 (C) | - | 60.9 (CH ₂) | 4.71 (d, 12.6) 4.76 (d, 12.6) |
| 4 | 49.5 (CH) | 2.88 (dd, 9.6, 8.0) | 120.5 (C) | - |
| 4a | 46.0 (CH) | 1.82 (m) | 46.9 (CH) | 2.28 (m) |
| 5 | 34.9 (CH ₂) | 1.46 (m) 1.87 (m) | 33.2 (CH ₂) | 1.47 (m) 1.63 (m) |
| 6 | 39.5 (CH ₂) | 2.17 (dt, 12.6, 3.5) 2.24 (dt, 4.3, 12.6) | 39.7 (CH ₂) | 1.89 (m) 1.99 (m) |
| 7 | 132.8 (C) | - | 138.1 (C) | - |
| 8 | 130.7 (CH) | 5.18 (d, 7.5) | 124.0 (CH) | 5.27 (d, 10.1) |
| 9 | 67.0 (CH) | 4.72 (br t, 6.6) | 72.4 (CH) | 5.52 (m) |
| 10 | 46.0 (CH ₂) | 2.38 (dd, 13.7, 1.2) 2.43 (dd, 13.7, 6.0) | 36.5 (CH ₂) | 1.96 (m) 2.57 (dd, 12.8, 5.2) |
| 11 | 148.7 (C) | - | 142.1 (C) | - |
| 11a | 48.4 (CH) | 1.79 (m) | 52.3 (CH) | 2.46 (dt, 4.9, 10.4) |
| 12 | 130.1 (CH) | 5.53 (dd, 15.5, 8.0) | 131.0 (CH) | 6.08 (d, 11.0) |
| 13 | 131.3 (CH) | 5.62 (dt, 15.5, 7.2) | 121.5 (CH) | 6.44 (dd, 15.2, 11.0) |
| 14 | 46.6 (CH ₂) | 2.27 (d, 7.2) | 143.6 (CH) | 5.88 (d, 15.2) |
| 15 | 70.5 (C) | - | 70.8 (C) | - |
| 16 | 29.0 (CH ₃) | 1.21 (s) | 29.7 (CH ₃) | 1.35 (s) |
| 17 | 29.4 (CH ₃) | 1.24 (s) | 29.7 (CH ₃) | 1.35 (s) |
| 18 | 17.3 (CH ₃) | 1.68 (s) | 19.5 (CH ₃) | 1.69 (s) |
| 19 | 115.6 (CH ₂) | 4.88 (br s) 5.02 (br s) | 120.1 (CH ₂) | 5.00 (br s) 5.02 (br s) |
| 1-Ac | | | 21.3 (CH ₃) 170.6 (C) | 2.06 (s) - |
| 3-Ac | | | 21.0 (CH ₃) 171.0 (C) | 2.09 (s) - |
| 9-Ac | | | 20.9 (CH ₃) 170.6 (C) | 1.94 (s) - |

^a 100 MHz, CDCl₃, ^b 400 MHz, CDCl₃, ^c 125 MHz, CDCl₃, ^d 500 MHz, CDCl₃

Hz), δ_C 67.0 (CH), 130.7 (CH), 132.8 (C)], one oxygenated methylene [δ_H 4.06 (1H, dd, $J = 11.5, 2.8$ Hz), 4.19 (1H, dd, $J = 11.5, 3.9$ Hz), δ_C 67.7 (CH₂)] and lactone carbonyl [δ_C 174.2 (C)]. The above functional groups were extended to the partial structures, -O-CH₂-CH- (C-1 and C-11a), -CH-CH=CH-CH₂- (C-4, C-12, C-13 and C-14), -CH-CH₂-CH₂- (C-4a, C-5 and C-6) and -C(CH₃)=CH-CH(OH)-CH₂- (C-7, C-18, C-8, C-9 and C-10) according to the COSY spectrum. These partial structures and the other functional groups were connected together based on the HMBC spectrum; also observed were following the cross peaks: H-1/C-3; H-4/C-3, C-4a; H-4a/C-11a; H-6/C-7; H-10/C-11; H-11a/C-11; H-14/C-15; Me-16/C-15 and Me-17/C-15. Based on these findings, it was thus possible to construct the xenicane skeleton. The *E* configuration of the carbon-carbon double bond at C-7 position was indicated by ¹³C chemical shift (δ_C 17.3, CH₃) of the olefinic methyl group⁹ at the C-18 position and NOE correlations between H-9 (δ_H 4.72) and Me-18 (δ_H 1.68) (Figure 1).

Relative configurations of all chiral centers in **1** were clarified from the NOESY spectrum (Figure 1). *trans*-Juncture of two rings was indicated by NOE correlations between H-4 (δ_H 2.88) and H-11a (δ_H 1.79) and between H-4a (δ_H 1.82) and H-19 (δ_H 4.88), suggesting H-4 and H-11a to be on the same face of the ring and H-4a and the *exo* methylene moiety to be on the opposite face to H-11a. Conformation of the 9-membered

ring was inferred based on NOE correlations between H-4a and H-8 (δ_H 5.18) and between H-8 and H-19 (δ_H 5.02) and between H-11a and Me-18 (δ_H 1.68). Stereochemistry of the hydroxy group at C-9 was found to show an α configuration by NOE correlation between H-9 (δ_H 4.72) and H-18. The relative configuration of **1** was thus assigned to 4*R**, 4a*S**, 9*R** and 11a*R**. The absolute configuration of **1** was determined by application of the modified Mosher's method.¹⁰ Dihydroxeniolidide-A (**1**) was converted to (*R*)-MTPA ester and (*S*)-MTPA ester, respectively. The ¹H NMR spectrum of each MTPA ester was measured and Figure 2 shows the value of $\Delta\delta$. The signs are positive due to left-sided protons but negative owing to right-sided protons, thus demonstrating the 9*R* configuration. The present results indicate 4*R*, 4a*S*, 9*R* and 11a*R* for **1**.

Isoxeniatriacetate (**2**) was shown to have the molecular formula C₂₆H₃₈O₇ based on HREIMS. The IR spectrum of **2** indicated absorptions due to a hydroxy group (3391 cm⁻¹) and ester carbonyl (1737 cm⁻¹).

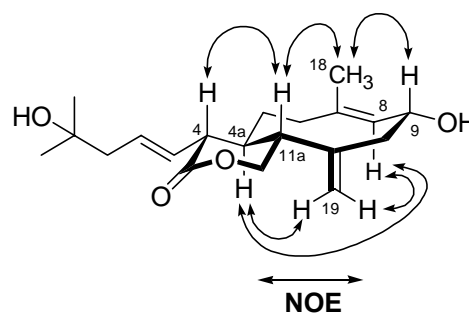
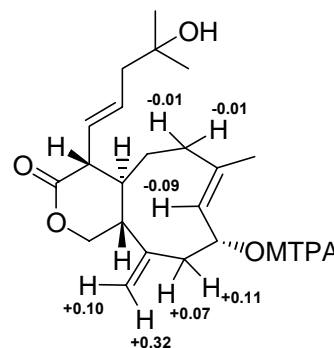


Figure 1. Selected NOE correlations of **1**.



$$\Delta\delta = \delta(S)\text{-MTPA ester} - \delta(R)\text{-MTPA ester}$$

Figure 2. $\Delta\delta$ obtained for the MTPA ester of **1**.

The conjugated diene group (-CH=CH-CH=C-) could be seen from the UV spectrum [λ_{\max} 244 nm (ϵ 29,000)]. All twenty-six carbons appeared in the ^{13}C NMR and DEPT spectra indicated six methyls, five sp^3 methylenes, one sp^2 methylene, three sp^3 methines, four sp^2 methines, one sp^3 quaternary carbon and six sp^2 quaternary carbons to be presented (Table 1). ^1H and ^{13}C NMR correlations were evident from the HMQC spectrum. ^1H and ^{13}C NMR spectra disclosed three acetyl groups [δ_{H} 1.94 (3H, s), 2.06 (3H, s), 2.09 (3H, s), δ_{C} 170.6 (C), 170.6 (C), 171.0 (C)], one conjugated diene group (-CH=CH-CH=C-) [δ_{H} 5.88 (1H, d, $J = 15.2$ Hz), 6.08 (1H, d, $J = 11.0$ Hz), 6.44 (1H, dd, $J = 15.2, 11.0$ Hz) δ_{C} 120.5 (C), 121.5 (CH), 131.0 (CH), 143.6 (CH)], one *exo* methylene [δ_{H} 5.02 (1H, br s), 5.02 (1H, br s), δ_{C} 120.1 (CH₂), 142.1 (C)], one olefinic methyl [δ_{H} 1.69 (3H, s), δ_{C} 19.5 (CH₃)], two methyls [δ_{H} 1.65 (3H, s), 1.35 (3H, s), δ_{C} 29.7 (CH₃), 29.7 (CH₃)], one oxygenated quaternary carbon [δ_{C} 70.5 (C)], one oxygenated methine adjacent to trisubstituted olefin [δ_{H} 5.27 (1H, d, $J = 10.1$ Hz), 5.52 (1H, m), δ_{C} 72.4 (CH), 124.0 (CH), 138.1 (C)] and two oxygenated methylenes [δ_{H} 3.80 (1H, dd, $J = 11.5, 10.4$ Hz), 4.04 (1H, dd, $J = 11.5, 4.9$ Hz), δ_{C} 64.9 (CH₂); δ_{H} 4.71 (1H, d, $J = 12.6$ Hz), 4.76 (1H, d, $J = 12.6$ Hz), δ_{C} 60.9 (CH₂)]. The above functional groups were extended to the partial structures, -O-CH₂-CH-CH-CH₂- (C-1, C-11a, C-4a and C-5), -C=CH-CH=CH- (C-4, C-12, C-13 and C-14) and -C(CH₃)=CH-CH(OAc)-CH₂-C=CH₂ (C-7, C-18, C-8, C-9, C-10, C-11 and C-19) based on the COSY spectrum. These partial structures and the other functional groups were found to be connected based on the HMBC spectrum; also observed were included the cross peaks: H-1/1-Ac; H-3/3-Ac, C-4; H-4a/C-4, C-12; H-6/C-4a, C-5, C-7, C-8, Me-18; H-9/9-Ac; H-11a/C-11 and H-14/C-15, so that the xenicane skeleton could be constructed. The *E* configuration of the carbon-carbon double bond at C-7 position was indicated by ^{13}C chemical shift (δ_{C} 19.5, CH₃) of the olefinic methyl group⁸ at the C-18 position and NOE correlations between H-9 (δ_{H} 5.52) and Me-18 (δ_{H} 1.69). The *Z* configuration of the carbon-carbon double bond at C-4 position was indicated by the NOE correlations between H-3 (δ_{H} 4.71 and 4.76) and H-13 (δ_{H} 6.44) and between H-4a (δ_{H} 2.28) and H-12 (δ_{H} 6.08) (Figure 3).

Relative configurations of all chiral centers in **2** were elucidated based on the NOESY spectrum (Figure 3). NOE correlations between H-11a (δ_{H} 2.46) and H-19 (δ_{H} 5.00), among H-9 (δ_{H} 5.52), H-18 (δ_{H} 1.69) and H-19 (δ_{H} 5.02) and between H-4a (δ_{H} 2.28) and H-8 (δ_{H} 5.27), which suggested H-9 and H-11a to be on the same face of the 9-membered ring and H-4a to be on the opposite face to H-11a. The relative configuration of **2** was thus assigned to 4a*S**, 9*R** and 11a*R**.

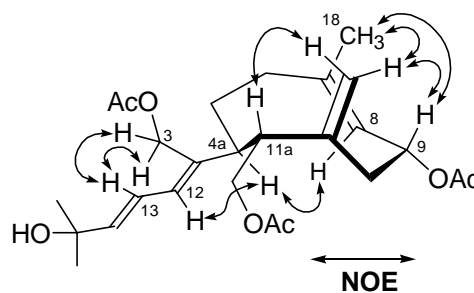
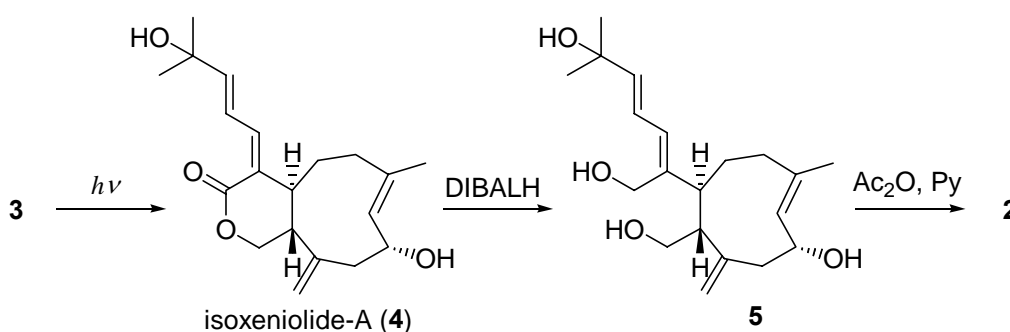


Figure 3. Selected NOE correlations of **2**.

The absolute configuration of isoxeniatriacetate (**2**) was determined by chemical conversion of xeniolide-A (**3**) to isoxeniatriacetate (**2**) (Scheme 1). A solution of xeniolide-A (**3**) in benzene was irradiated with a high pressure mercury lamp to provide isoxeniolide-A (**4**), $[\alpha]_D^{20} +41.0^\circ$ (c 0.40, MeOH), lit. $[\alpha]_D +50^\circ$ (c 0.655, MeOH).¹¹ Isoxeniolide-A (**4**) was reduced with DIBALH in THF at -78°C to give tetraol (**5**), $[\alpha]_D^{27} -184^\circ$ (c 0.90, MeOH₃). Treatment of tetraol (**5**) with acetic anhydride and pyridine afforded triacetate (**2**), $[\alpha]_D^{22} -178^\circ$ (c 0.09, CHCl₃). Spectral data and sign of optical rotation of synthesized **2** were identical to those of natural isoxeniatriacetate (**2**). This conversion demonstrated the absolute configuration of **2** to be 4a*S*, 9*R* and 11a*R*.



Scheme 1. Chemical conversion of xeniolide-A (**3**) to isoxeniatriacetate (**2**).

EXPERIMENTAL

General Experimental Procedures. Optical rotations were measured with a JASCO DIP-360 automatic polarimeter and IR spectra were taken with a Perkin-Elmer FT-IR 1710 spectrophotometer or JASCO A-302 spectrophotometer. ¹H and ¹³C NMR spectra were measured with a Bruker AM-400 or AM-500. Chemical shifts are given on a δ (ppm) scale with tetramethylsilane (TMS) as the internal standard. EIMS and HREIMS spectra were obtained with a VG Auto Spec spectrometer.

Animal Material, Extraction and Isolation. Soft coral *Xenia* sp. was collected from the coral reef of Ishigaki Island (Okinawa, Japan) in May 1992 at a depth of 1-3 m. A voucher specimen (SC-II-1) is deposited at this laboratory, School of Pharmacy, Tokyo University of Pharmacy and Life Science (Tokyo, Japan). Wet specimens (8.0 kg) were immersed in MeOH (5.0 L x 3) at rt for 24 h and MeOH extracts (222 g) were partitioned between EtOAc (1.5 L x 2) and H₂O (1.5 L) to give EtOAc-soluble portions (14.1 g). A part (7.9 g) of the EtOAc-soluble portions was chromatographed on a silica gel column to give the following fractions; fraction 1 (1.4 g) eluted with hexane-EtOAc = 10 : 1 (1.0 L), fraction 2 (1.7 g) eluted with hexane-EtOAc = 3 : 1 (1.0 L), fraction 3 (3.6 g) eluted with EtOAc (1.0 L) and fraction 4 (1.2 g) eluted with MeOH (1.0 L). Fraction 3 was subjected to flash silica gel column chromatography (hexane-EtOAc = 3 : 1), providing fractions 3-1 (1.86 g), 3-2 (880 mg), 3-3 (790 mg) and 3-4 (70 mg). Fraction 3-1, by repeated flash silica gel column chromatography (hexane-EtOAc = 6 :

5 for the first chromatography and hexane-EtOAc = 3 : 2 for the second chromatography) gave fractions 3-1-1 (104 mg), 3-1-2 (73 mg) and 3-1-3 (55 mg). Fraction 3-1-2 subjected to normal phase HPLC (hexane-EtOAc = 2 : 1) gave isoxeniatricetate (**2**) (11.0 mg). From fraction 3-2 *via* to repeated flash silica gel column chromatography (CHCl₃-acetone = 3 : 1) xeniolide-A⁶ (**3**) (634 mg) and fraction 3-2-2 (246 mg) were obtained. Fraction 3-2-2 was subjected to normal phase HPLC (CHCl₃-MeOH = 30 : 1) and flash ODS column chromatography (MeOH-H₂O = 3 : 1) followed by normal phase HPLC (hexane-EtOAc = 2 : 3) to give dihydroxeniolide-A (**1**) (14 mg). Fraction 3-3 was subjected to normal phase HPLC (CHCl₃-MeOH = 30 : 1) and flash ODS column chromatography (MeOH-H₂O = 3 : 1) followed by normal phase HPLC (hexane-EtOAc = 1 : 2) to give xeniolide-B⁶ (9.0 mg). Fraction 3-4 was subjected to flash silica gel column chromatography (CHCl₃-acetone = 2 : 1) to give xenialactol^{7,8} (21 mg).

Dihydroxeniolide-A (1): Colorless oil; $[\alpha]_D^{25} -31.2^\circ$ (*c* 1.21, CHCl₃); IR (CHCl₃) ν_{\max} 3500, 1735, 1640 cm⁻¹; ¹H and ¹³C NMR spectra see Table 1; HMBC correlation (H/C) 3/1, 3/4, 3/4a, 3/12, 4a/3, 4a/4, 4a/5, 4a/6, 4a/11a, 4a/12, 6/4a, 6/5, 6/7, 6/8, 6/18, 8/6, 8/9, 8/18, 9/7, 9/8, 9/10, 10/8, 10/9, 10/11, 10/11a, 10/19, 11a/1, 11a/4, 11a/4a, 11a/10, 11a/11, 11a/19, 12/3, 12/4a, 12/13, 12/14, 13/4, 13/12, 13/14, 13/15, 14/13, 14/15, 14/15, 14/16, 16/14, 16/15, 16/17, 17/14, 17/15, 17/16, 18/6, 18/8, 19/10, 18/11a; NOESY correlation (H/H) 3/11a, 3/12, 4a/8, 9/18, 11a/19, 14/17, 18/19; FABMS *m/z* 335 [M+H]⁺; EIMS *m/z* 316 [M-H₂O]⁺; HREIMS *m/z* 316.2043 (calcd for C₂₀H₂₈O₃, 316.2038).

Isoxeniatricetate (2): Colorless oil; $[\alpha]_D^{27} -196^\circ$ (*c* 0.55, CHCl₃); IR (CHCl₃) ν_{\max} 3391, 1737, 1240 cm⁻¹; UV (MeOH) λ_{\max} 244 nm (ϵ 29,000); ¹H and ¹³C NMR spectra see Table 1; HMBC correlation (H/C) 3/1, 3/4, 3/4a, 3/12, 4a/3, 4a/4, 4a/5, 4a/6, 4a/11a, 4a/12, 6/4a, 6/5, 6/7, 6/8, 6/18, 8/6, 8/9, 8/18, 9/7, 9/8, 9/10, 10/8, 10/9, 10/11, 10/11a, 10/19, 11a/1, 11a/4, 11a/4a, 11a/10, 11a/11, 11a/19, 12/3, 12/4a, 12/13, 12/14, 13/4, 13/12, 13/14, 13/15, 14/13, 14/15, 14/15, 14/16, 16/14, 16/15, 16/17, 17/14, 17/15, 17/16, 18/6, 18/8, 19/10, 18/11a; NOESY correlation (H/H) 3/11a, 3/12, 4a/8, 9/18, 11a/19, 14/17, 18/19; EIMS *m/z* 462 [M]⁺; HREIMS *m/z* 462.2599 (calcd for C₂₆H₃₈O₇, 462.2618).

(R)-MTPA ester of dihydroxeniolide-A (1). To a solution of dihydroxeniolide-A (**1**) (1.0 mg, 3.0 μ mol) in CHCl₃ (0.2 mL) were added DMAP (1.0 mg) and (R)-MTPA-Cl (2.0 mg, 7.9 μ mol). After stirring at rt for 12 h, the reaction mixture was diluted with EtOAc and washed with H₂O and saturated NaCl aqueous solution. The organic layer was dried over MgSO₄ and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (hexane-EtOAc = 2 : 3) to give (R)-MTPA ester (1.0 mg, 61 %) as a colorless oil: $[\alpha]_D^{25} -21.7^\circ$ (*c* 0.12, CHCl₃); IR (CHCl₃) ν_{\max} 3550, 1740 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 7.54 (2H, m), 7.41 (3H, m), 5.89 (1H, br t, *J* = 6.5 Hz), 5.61 (1H, dt, *J* = 15.5, 7.3 Hz), 5.50 (1H, dd, *J* = 15.5, 8.2 Hz), 5.29 (1H, d, *J* = 7.1 Hz), 4.76 (1H, s), 4.53

(1H, s), 4.17 (1H, dd, $J = 11.5, 3.7$ Hz), 4.01 (1H, dd, $J = 11.5, 2.5$ Hz), 3.59 (3H, s), 2.86 (1H, dd, $J = 9.9, 8.2$ Hz), 2.45 (1H, dd, $J = 14.9, 5.9$ Hz), 2.39 (1H, dd, $J = 14.9, 1.5$ Hz), 2.26 (2H, d, $J = 7.1$ Hz), 2.21 (1H, dt, $J = 12.6, 2.9$ Hz), 2.07 (1H, dt, $J = 3.9, 12.6$ Hz), 1.90 (1H, m), 1.79 (1H, m), 1.77 (3H, s), 1.74 (1H, m), 1.46 (1H, m), 1.24 (3H, s), 1.21 (3H, s); EIMS m/z 492 $[M-C_3H_6O]^+$; HREIMS m/z 492.2119 (calcd for $C_{27}H_{31}O_5F_3$, 492.2124).

(S)-MTPA ester of dihydroxeniolide-A (1). To a solution of dihydroxeniolide-A (**1**) (1.0 mg, 3.0 μ mol) in $CHCl_3$ (0.2 mL) were added DMAP (1.0 mg) and (S)-MTPA-Cl (2.0 mg, 7.9 μ mol). After stirring at rt for 12 h, the reaction mixture was diluted with EtOAc and washed with H_2O and saturated NaCl aqueous solution. The organic layer was dried over $MgSO_4$ and concentrated under reduced presser. The residue was purified by silica gel column chromatography (hexane-EtOAc = 2 : 3) to give (S)-MTPA ester (1.0 mg, 61 %) as a colorless oil: $[\alpha]_D^{25} -103^\circ$ (c 0.15, $CHCl_3$); IR ($CHCl_3$) ν_{max} 3550, 1740 cm^{-1} ; 1H NMR ($CDCl_3$, 400 MHz) δ 7.54 (2H, m), 7.42 (3H, m), 5.87 (1H, m), 5.61 (1H, dt, $J = 15.5, 7.1$ Hz), 5.52 (1H, dd, $J = 15.5, 8.1$ Hz), 5.20 (1H, d, $J = 8.1$ Hz), 4.86 (1H, s), 4.85 (1H, s), 4.18 (1H, dd, $J = 11.5, 3.5$ Hz), 4.04 (1H, dd, $J = 11.5, 2.6$ Hz), 3.53 (3H, s), 2.87 (1H, dd, $J = 9.7, 8.1$ Hz), 2.51 (1H, s), 2.50 (1H, s), 2.27 (2H, d, $J = 7.1$ Hz), 2.20 (1H, dt, $J = 12.7, 3.2$ Hz), 2.06 (1H, dt, $J = 4.3, 12.7$ Hz), 1.90 (1H, dt, $J = 14.1, 3.6$ Hz), 1.80 (1H, m), 1.77 (1H, m), 1.76 (3H, s), 1.55 (1H, m), 1.24 (3H, s), 1.21 (3H, s); EIMS m/z 550 $[M]^+$; HREIMS m/z 550.2558 (calcd for $C_{30}H_{37}O_6F_3$, 550.2542).

Photoisomerization of xeniolide-A (3) to isoxeniolide-A (4). A solution of xeniolide-A (**3**) (45.0 mg, 136 μ mol) in benzene (20 mL) was irradiated with a 100W high-pressure mercury lamp at rt for 5 h. The reaction mixture was concentrated under reduce presser. The residue was purified by HPLC (hexane-EtOAc = 2 : 3) to give isoxeniolide-A¹¹ (**4**) (39.5 mg, 88 %) as colorless crystals: $[\alpha]_D^{20} +41.0^\circ$ (c 0.40, MeOH); mp 170-172°C; IR ($CHCl_3$) ν_{max} 3460, 1725, 1640 cm^{-1} ; UV (MeOH) λ_{max} 263 nm (ϵ 16,100); 1H NMR ($CDCl_3$, 400 MHz) δ 6.85 (1H, dd, $J = 15.5, 11.1$ Hz), 6.36 (1H, d, $J = 11.1$ Hz), 6.07 (1H, d, $J = 15.5$ Hz), 5.21 (1H, d, $J = 7.3$ Hz), 5.08 (1H, s), 4.96 (1H, s), 4.77 (1H, m), 4.08 (1H, dd, $J = 11.4, 5.9$ Hz), 3.63 (1H, dd, $J = 12.3, 11.4$ Hz), 2.60 (1H, m), 2.46 (1H, dd, $J = 13.7, 6.1$ Hz), 2.39 (1H, d, $J = 13.7$ Hz), 2.19 (2H, m), 2.01 (1H, m), 1.70 (3H, s), 1.64 (2H, m), 1.37 (3H, s), 1.37 (3H, s); ^{13}C NMR ($CDCl_3$, 100 MHz) δ 169.0 (s), 147.8 (s), 147.8 (s), 136.0 (d), 132.7 (s), 132.6 (s), 130.6 (d), 122.4 (d), 114.8 (t), 71.0 (t), 70.8 (s), 67.0 (d), 50.9 (d), 49.9 (d), 45.5 (t), 39.8 (t), 37.7 (t), 29.4 (q), 29.3 (q), 17.3 (q); EIMS m/z 273 $[M-C(CH_3)_2OH]^+$; HREIMS m/z 273.1465 (calcd for $C_{17}H_{21}O_3$, 273.1491).

DIBALH reduction of isoxeniolide-A (4) to tetraol (5). To a cold (-78°C) solution of isoxeniolide-A (**4**) (18.0 mg, 54 μ mol) in THF (2.0 mL) was added DIBALH (600 μ L, 558 μ mol, 0.93M in hexane). The mixture was stirred for 10 min, treated with MeOH (0.1 mL), diluted with Et_2O , treated with saturated aqueous NaCl solution and stirred at rt for 2 h. The organic layer was dried over $MgSO_4$ and

concentrated under reduced pressure. The residue was purified by silica gel column chromatography (hexane-acetone = 1 : 2) to afford tetraol (**5**) (10.0 mg, 55 %) as a colorless oil: $[\alpha]_{\text{D}}^{27} -184^{\circ}$ (*c* 0.90, MeOH); IR (CHCl₃) ν_{max} 3430, 1635 cm⁻¹; UV (MeOH) λ_{max} 243 nm (ϵ 19,700); ¹H NMR (CDCl₃, 400 MHz) δ 6.56 (1H, dd, *J* = 15.2, 11.0 Hz), 5.96 (1H, d, *J* = 11.0 Hz), 5.80 (1H, d, *J* = 15.2 Hz), 5.29 (1H, d, *J* = 9.9 Hz), 5.01 (1H, s), 4.94 (1H, s), 4.62 (1H, dt, *J* = 5.4, 9.9 Hz), 4.25(2H, m), 3.49 (2H, m), 2.58 (1H, dd, *J* = 12.6, 5.1 Hz), 2.38 (1H, m), 2.19 (2H, m), 2.03 (1H, m), 1.95 (1H, m), 1.89 (1H, m), 1.63 (3H, s), 1.45 (2H, m), 1.34 (3H, s), 1.34 (3H, s); ¹³C NMR (CDCl₃, 100 MHz) δ 144.6 (d), 144.2 (s), 141.9 (s), 136.2 (s), 128.5 (d), 128.1 (d), 122.0 (d), 119.1 (t), 70.9 (s), 70.4 (d), 63.8 (t), 59.5 (t), 56.9 (d), 56.9 (d), 40.6 (t), 39.9 (t), 33.8 (t), 30.0 (q), 29.7 (q), 19.2 (q); EIMS *m/z* 287 [M-H₂O-CH₂OH]⁺.

Acetylation of tetraol 5 to isoxeniatriacetate (2). To a solution of tetraol (**5**) (1.8 mg, 5.0 μ mol) in pyridine (200 μ L) was added acetic anhydride (200 μ L), followed by stirring at rt for 15 min. The reaction mixture was concentrated under reduced pressure and the residue was purified by silica gel column chromatography (hexane-EtOAc = 2 : 1) to give isoxeniatriacetate (**2**) (1.0 mg, 43 %) as a colorless oil: $[\alpha]_{\text{D}}^{22} -178^{\circ}$ (*c* 0.09, CHCl₃).

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