

SYNTHESIS AND ACTIVITY OF A TETRAHYDROFOLATE INHIBITOR OF THE ENZYME

N¹⁰-FORMYL-H₄-FOLATE-MET-tRNA^{fMet} TRANSFORMYLASE

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Abstract - The N-formylmethionine-tRNA^{fMet} is involved in the initiation of protein biosynthesis in procaryotic organisms and is formed via the transfer of a formyl group from N¹⁰-formyltetrahydrofolic acid to the amino group of methionine in Met-tRNA^{fMet}. Based on a proposed intermediate for this formylation step, a transition state analog was designed and synthesized and was shown in vitro to be an excellent inhibitor of the transformylase reaction.

The enzyme Met-tRNA^{fMet} transformylase from E. coli has been described¹ and shown to catalyze the transfer of a formyl group² from N¹⁰-formyltetrahydrofolic acid (2) to the free amino group of methionine in Met-tRNA^{fMet} (1), (Scheme I). The resulting N-formylmethionine-tRNA^{fMet} (4) is involved in the initiation of protein biosynthesis in procaryotic organisms.³ The significance of this transformylation process for bacterial gene expression makes the inhibition of this process an interesting target for the potential development of antibacterial agents.

The transfer of the formyl group from 2 to 1 is unique for bacterial protein synthesis but it should be recognized that both 1 and 2 have functions in mammalian biochemical transformations⁴ although the transformylase enzyme is lacking from eukaryotic cell extracts.⁵ Therefore to reduce the possibility of toxicity, any analog designed to inhibit this process should closely resemble the transition state involved in the formyl transfer reaction catalyzed by the transformylase. Although a stable fMet-tRNA^{fMet} enzyme has been isolated, there is no evidence of a formyl transfer to the enzyme yielding an acid stable formylated enzyme.⁶ Our results suggest that a formyl transfer may in fact take place directly from compound 2 to 1 implying the intermediacy of a structure

The diagram illustrates a chemical reaction scheme involving aminoacyl-tRNA species and a transacylation reaction.

Structure 1: An aminoacyl-tRNA. The tRNA is linked to a phosphate group (tRNA-O-P(=O)(O⁻)₂), which is connected to a ribose sugar. The ribose is linked via an ester bond to the 3'-carbon of a threonine derivative. The threonine side chain is $\text{CH}(\text{OH})\text{CH}_2\text{CH}_2\text{CH}_3$. The amino group (NH_2) is attached to the α -carbon of this threonine derivative.

Structure 2: A complex molecule consisting of a 4-aminopyrimidin-2(1H)-one ring system linked via a methylene group to a benzene ring. The benzene ring is further linked to a side chain containing a carboxylic acid group (COOH) and a methylene group (CH_2) attached to a carboxylic acid group (CH_2COOH).

Reaction: An arrow labeled "transacylation" points from Structure 1 to Structure 3.

Structure 3: A complex molecule shown within brackets. It features a threonine derivative side chain ($\text{CH}(\text{OH})\text{CH}_2\text{CH}_2\text{CH}_3$) linked to a nitrogen atom. This nitrogen atom is part of a larger structure that includes a benzene ring and a side chain with a carboxylic acid group (COOH) and a methylene group (CH_2) attached to a carboxylic acid group (CH_2COOH).

Structure 4: A modified aminoacyl-tRNA. The tRNA is linked to a phosphate group (tRNA-O-P(=O)(O⁻)₂), which is connected to a ribose sugar. The ribose is linked via an ester bond to the 3'-carbon of a threonine derivative. The threonine side chain is $\text{CH}(\text{OH})\text{CH}_2\text{CH}_2\text{CH}_3$. The amino group (NH_2) is attached to the α -carbon of this threonine derivative.

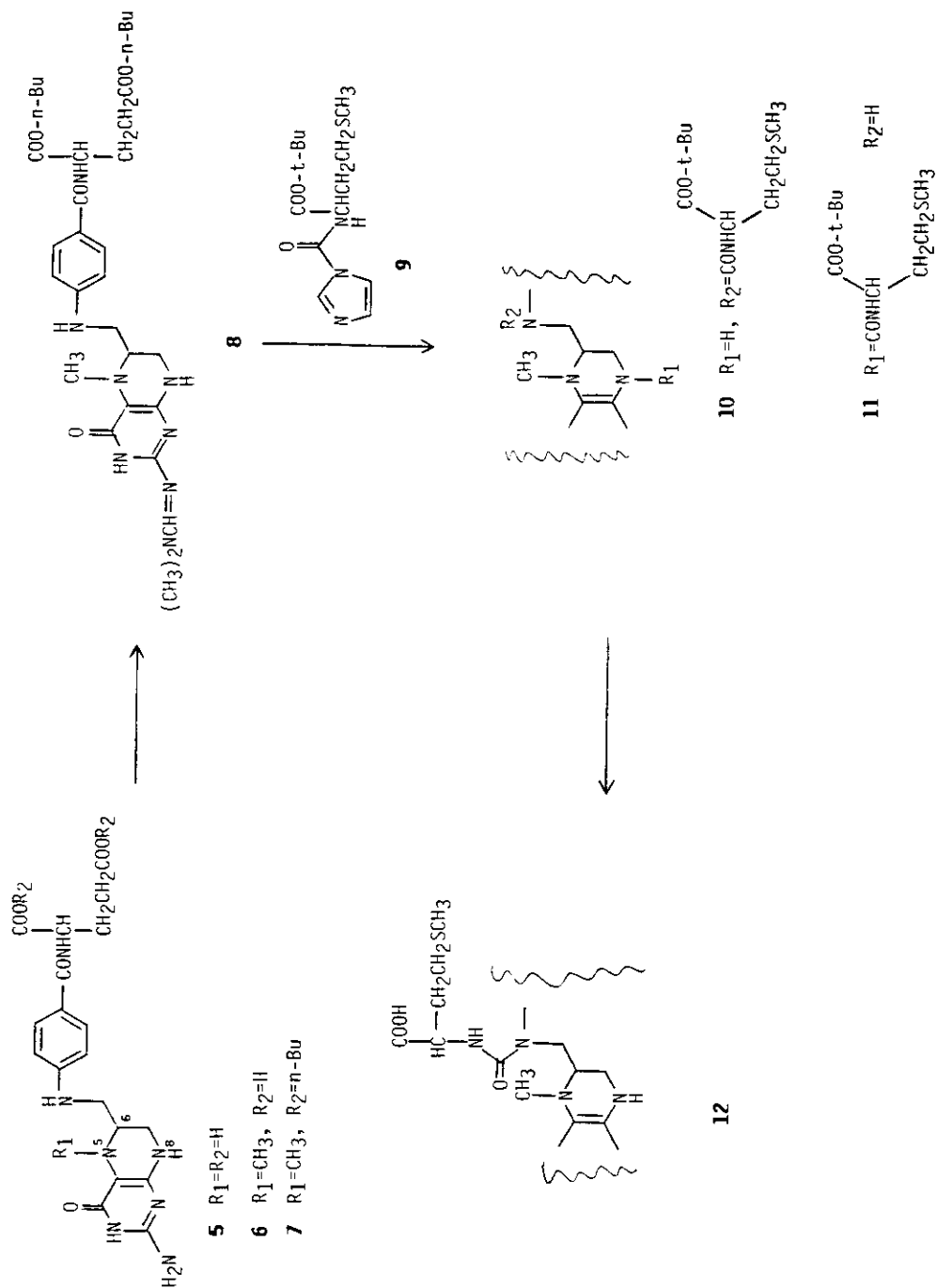
such as **3**⁷ in the bacterial system. With this in mind, we envisaged a compound such as **15** as a possible stable transition state analog, which, although lacking a tetrahedral equivalent to the α -amino alcohol of intermediate **3**, might still function as an inhibitor of this one carbon transfer reaction.

The preparation of several N^5 - and N^5,N^{10} -disubstituted tetrahydrofolic acid analogs have been described,⁸ whereas only one unstable N^{10} -substituted analog has been reported.⁹ Because tetrahydrofolic acid **5**¹⁰ is known to undergo rapid air oxidation,¹¹ we decided to choose the more stable N^5 -methyltetrahydrofolic acid (**6**)¹² (racemic at C6) as our starting material for the preparation of the target intermediate molecule **12** (Scheme II). Because the N^5 -methyl-dl-L-H₄-folate has previously been shown to be an effective inhibitor of the transformylation reaction (although noncompetitive with the formyl donor),¹³ it was anticipated that substitution at N^5 with a methyl group would not interfere with the inhibition properties of the target compound **15**. Due to the insolubility of **6**¹⁴ the acid was first converted to its di-n-butyl ester **7** (1.5N HCl-nBuOH, 80°C, 16 h) in 29% overall yield from folic acid. Treatment of **7** with *N,N*-dimethylormamide dimethylacetal¹⁴ (DMF, 25°C, 4 h, argon) proceeded smoothly to yield the more soluble derivative **8**¹⁵ in 83% yield (mp 104-108°C) after florisil column chromatography.

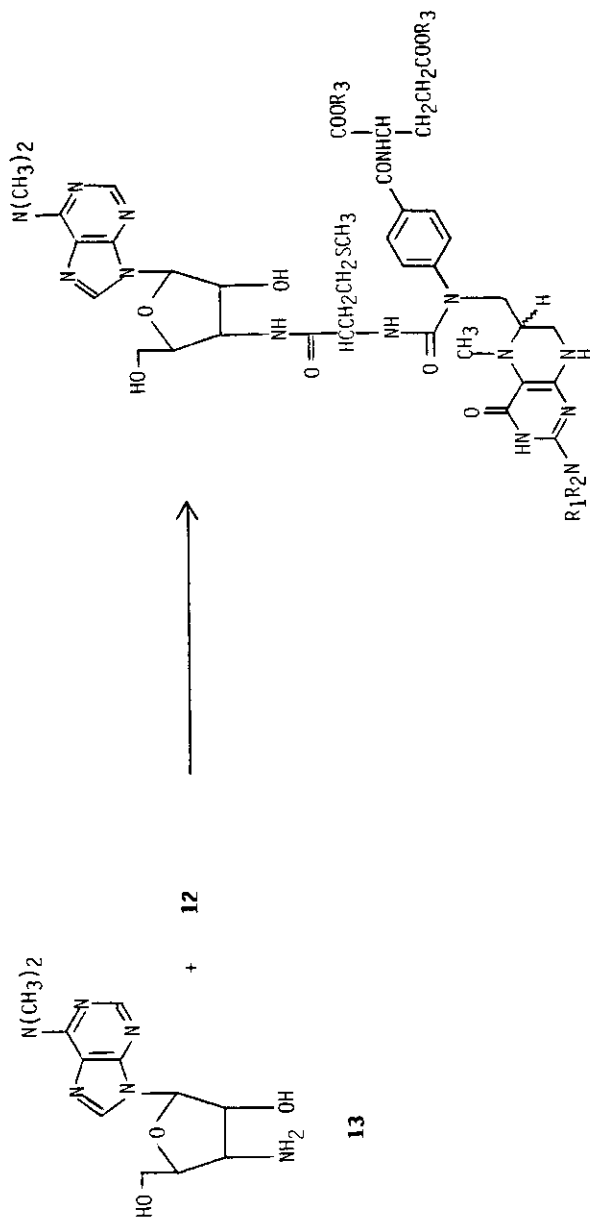
Treatment of the N^5 -methyltetrahydrofolate **8** with the methionyl synthon **9**¹⁶ (3 eq., DME, 85°C, 18 h, argon) produced the N^{10} -methionyl derivative **10** (64% yield) and a minor amount of the N^8 -methionyl analog **11**¹⁵ (11%) (separated by silica gel column chromatography). The *t*-butyl ester of compound **10** was then selectively cleaved (CF₃COOH, 25°C, 1 h) to produce the desired carboxylic acid derivative **12** (90% yield, as mono TFA salt). The final phase of the synthesis was accomplished (Scheme III) by coupling the acid **12** with the readily available puromycin aminonucleoside¹⁷ **13** in the presence of *N*-ethoxycarbonyl-2-ethoxy-1,2-dihydroquinoline¹⁸ (EEDQ) (DMF, 40°C, 16 h, argon). Thus the tetrahydrofolate transition state analog¹⁵ **14** was obtained in 32% yield after purification by silica gel column chromatography. It has been demonstrated that the antibiotic puromycin is capable of inhibiting protein synthesis due to the fact that it can successfully compete with the aminoacyl-tRNA for the ribosome.¹⁹ The puromycin aminonucleoside **13** was, for this reason, considered an appropriate substitute for adenosine and would allow for the selective acylation of the C3 amino group necessary for the construction of the potential transformylase inhibitor **15**.

The water soluble trisodium salt **15** was prepared from the ester **14**, (6 eq. 0.35N aqueous NaOH in 1:1 DME/H₂O, 25°C, 20 h) purified via silica gel chromatography (eluent 3:1 propanol/water) and lyophilized to give a white solid homogeneous by tlc (cellulose, 1:2 1% aqueous sodium bicarbonate/propanol). The structure of **15** rests on the method of synthesis, the ¹Hnmr (in D₂O/CD₃OH) δ 2.07 (s, 3H, SCH₃), 2.53 (s, 3H, NCH₃), 3.44 [s, N(CH₃)₂], 3.2-3.7 [m, NCH; NCH₂, overlapping N(CH₃)₂]

Scheme 11



Scheme 111



3.78, 4.02 (br., AB, $J_{\text{gem}}=12$ Hz, CH_2OH) 4.44 (m, 2H), 4.65-4.7 (m) 6.11 (br, s, 1H, NCHOC), 7.49-7.92 (AA'BB', 4H), 8.18 (s, 1H), 8.34 (s, 1H), the uv_{max} (H_2O) 215 nm ($\text{sh } \epsilon$ 42,300), 275 (30, 100) and amino acid analysis: Found Met (1.0), Glu(1.1).

The fact that **15** can serve as an inhibitor of the transformylase was demonstrated in vitro. In these experiments transformylase activity was assayed essentially as described by Dickerman et al.^{1b} Compound **15** was found to be an excellent inhibitor (~70% inhibition of transformylase activity at a concentration of 10^{-6} M) while the reference compound Ca leucovorin had an inhibitory activity of ~50% at 1×10^{-4} M. More detailed enzymatic studies will be required to determine the mode of inhibition by compound **15** and whether it is truly functioning as a transition state inhibitor.

ACKNOWLEDGEMENT

We are indebted to Professor E. C. Taylor for his advice and consultation.

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Received, 19th May, 1986