

SYNTHESIS AND BIOLOGICAL ACTIVITY OF A TUBERCIDIN ANALOG OF 2-5A

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Previous studies from this laboratory have demonstrated that oligonucleotide binding to the 2-5A-dependent endonuclease (RNase L) of mouse L cells decreases dramatically upon substitution of the adenine rings of 2-5A by uracil, cytosine, or hypoxanthine. To explore the structural requirements for base binding more closely, we prepared the tubercidin analog of 2-5A [ppp5'-(c⁷A)2'p5'-(c⁷A)2'p5'-(c⁷A)] by lead ion-catalyzed polymerization of 5'-phosphoroimidazolidine of c⁷A to give 2',5'-(pc⁷A)₃ which was converted to the triphosphate via pyrophosphate displacement on the phosphoroimidazolidine. This material and intermediates were thoroughly characterized by enzymatic digestions and NMR. The 5'-monophosphate, 2',5'-(pc⁷A)₃, as well as the 5'-triphosphate, 2',5'-pp(pc⁷A)₃, were bound to RNase L of mouse L cells approximately as well as 2-5A itself as judged by displacement of radiolabeled ppp5'A2'p5'A2'p5'A2'p5'A3'p5'Cp from the nuclease. Nonetheless, the 5'-triphosphate, 2',5'-pp(pc⁷A)₃ did not activate RNase L as judged by its ability to inhibit protein synthesis in encephalomyocarditis viral RNA-programmed extracts of L cells under conditions where 2-5A caused a 50% inhibition of translation at 10⁻⁹M. These results suggest that while the N-7 atoms of the adenine rings of 2-5A may not be involved in binding to RNase L, they are critical for the activation of the enzyme.

