

HETEROCYCLES, Vol. 79, 2009, pp. 765 - 771. © The Japan Institute of Heterocyclic Chemistry  
Received, 29th September, 2008, Accepted, 27th October, 2008, Published online, 31st October, 2008.  
DOI: 10.3987/COM-08-S(D)41

## NAUCLEAMIDE F, A NEW MONOTERPENE INDOLE ALKALOID FROM *NAUCLEA LATIFOLIA*

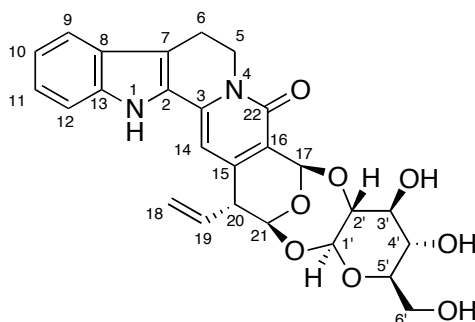
Yuka Kakuguchi, Haruaki Ishiyama, Takaaki Kubota, and Jun'ichi  
Kobayashi\*

Graduate School of Pharmaceutical Sciences, Hokkaido University, Sapporo  
060-0812, Japan. E-mail: jkobay@pharm.hokudai.ac.jp

**Abstract** - A new monoterpene indole alkaloid, naucleamide F (**1**), has been isolated from the bark and wood of *Nauclea latifolia*, and the structure and stereochemistry were elucidated on the basis of the spectral data. Naucleamide F (**1**) is a new monoterpene indole alkaloid consisting of a tetrahydro- $\beta$ -carboline ring fused to a pyridone ring, and a 1,3,5-trioxepane ring fused to a dihydropyran ring and a glucose unit.

### INTRODUCTION

A number of monoterpene indole alkaloids with biological activities have been isolated from *Nauclea* species (Rubiaceae).<sup>1-4</sup> In our search for bioactive metabolites from medicinal plants, we previously isolated new monoterpene indole alkaloids, naucleamides A~E<sup>5</sup>, from the bark and wood of *Nauclea latifolia*. Further investigation of extracts from this plant resulted in the isolation of a new monoterpene indole alkaloid, naucleamide F (**1**), consisting of a heptacyclic ring system including a tetrahydro- $\beta$ -carboline ring fused to a pyridone ring, and a 1,3,5-trioxepane ring fused to a dihydropyran ring and a glucose unit. Here we describe the isolation and structure elucidation of **1**.



**1**

## RESULTS AND DISCUSSION

The bark and wood of *Nauclea latifolia* were extracted with MeOH. The MeOH extracts were partitioned between hexane and 90% aqueous MeOH, and then the MeOH layer was subsequently extracted with *n*-BuOH. The *n*-BuOH-soluble materials were purified by a silica gel column (CHCl<sub>3</sub>-MeOH, 1:0 → 85:15) followed by a C<sub>18</sub> column (MeOH-H<sub>2</sub>O, 60:40) and C<sub>18</sub> HPLC (CH<sub>3</sub>CN-H<sub>2</sub>O, 40:60) to afford naucleamide F (**1**, 0.0003%) together with known related monoterpene indole alkaloids, angustoline<sup>6</sup> (**2**, 0.0004%), compound **3**<sup>7</sup> (0.0004%), and compound **4**<sup>7</sup> (0.0003%).

Table 1. <sup>1</sup>H- and <sup>13</sup>C-NMR Data of Naucleamide F (**1**) in CD<sub>3</sub>OD

Position	<sup>1</sup> H <sup>a</sup>	<sup>13</sup> C <sup>a</sup>
1	-	-
2	-	128.9
3	-	141.0
4	-	-
5a	4.42 (ddd, <i>J</i> = 6.0, 8.4, 14.1)	42.8
5b	4.65 (dt, <i>J</i> = 6.6, 14.1)	
6	3.17 (m) <sup>b</sup>	21.2
7	-	118.4
8	-	127.6
9	7.63 (d, 7.8)	121.3
10	7.13 (t, 7.2)	122.0
11	7.29 (t, 7.8)	126.5
12	7.44 (d, 8.4)	113.7
13	-	141.3
14	6.67 (s)	103.9
15	-	151.4
16	-	116.9
17	6.04 (s)	93.6
18a	5.32 (d, <i>J</i> = 10.8)	120.8
18b	5.38 (d, <i>J</i> = 17.4)	
19	5.86 (ddd, <i>J</i> = 7.8, 10.2, 17.4)	136.2
20	3.50 (d, <i>J</i> = 7.8)	48.8
21	5.51 (s)	96.6
22	-	178.8
1'	5.06 (d, <i>J</i> = 7.2)	99.8
2'	3.22 (d, <i>J</i> = 7.2)	82.2
3'	3.67 (m)	77.0
4'	3.30-3.40 (m)	71.4
5'	3.30-3.40 (m)	79.8
6'a	3.71 (dd, <i>J</i> = 4.2, 12.0)	63.3
6'b	3.88 (dd, <i>J</i> = 4.2, 12.0)	

<sup>a</sup> δ in ppm, <sup>b</sup> 2H

The molecular formula,  $C_{26}H_{26}N_2O_8$ , of nucleamide **1** was established by HR-ESI-MS [ $m/z$  517.1592 ( $M+Na$ )<sup>+</sup>,  $\Delta$  +0.5 mmu]. IR absorptions implied the presence of hydroxy ( $3443\text{ cm}^{-1}$ ) and amide carbonyl ( $1645\text{ cm}^{-1}$ ) functionalities.  $^1\text{H}$  and  $^{13}\text{C}$  NMR data (Table 1) and the HMQC spectrum suggested that **1** possessed one carbonyl, seven  $\text{sp}^2$  quaternary carbons, six  $\text{sp}^2$  methines, one  $\text{sp}^2$  methylene, three  $\text{sp}^3$  methylenes, one  $\text{sp}^3$  methine, four  $\text{sp}^3$  oxymethines, and three acetal methines. Among them, one oxymethylene carbon ( $\delta_{\text{C}}$  63.3), five oxymethine carbons ( $\delta_{\text{C}}$  82.2, 79.8, 77.0, 71.4, and 63.3), and one acetal methine carbon ( $\delta_{\text{C}}$  99.8) were ascribed to a glucopyranose unit (C-1'~C-6').<sup>8</sup> The  $^1\text{H}$ - $^1\text{H}$  COSY and TOCSY spectra of **1** revealed connectivities of four partial structures, C-5 to C-6, C-9 to C-12, C-18 to C-20, and C-1' to C-6'. HMBC cross-peaks of H-5 to C-7 and C-22, H-9 to C-7 and C-13, H-12 to C-8, H-14 to C-2 and C-3, and H-20 to C-14 and C-15 indicated the presence of a tetrahydro- $\beta$ -carboline ring (N-1, C-2, C-3, N-4, and C-5~C-13) fused to a pyridone ring (C-3, N-4, C-14~C-16, and C-22) at C-3 and N-4, which was connected to an  $\text{sp}^3$  methine (C-20). The presence of a 1,3,5-trioxepane ring (C-17, O-17, C-21, O-1', C-1', C-2', and O-2') fused to a dihydropyran ring (C-15~C-17, C-20, C-21, and O-17) at C-17 and C-21, and a glucose unit (C-1'~C-6' and O-5') at C-1' and C-2' was elucidated by HMBC correlations of H-17 to C-16 and C-2', H-21 to C-15 and C-17, and NOESY correlations for H-20 to H-21 and H-21 to H-1'.

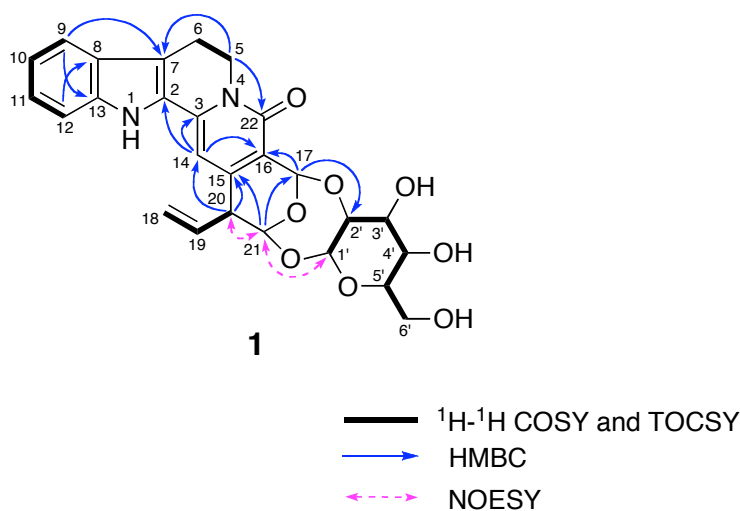


Figure 1. Selected 2D NMR correlations for nucleamide **1**.

The relative stereochemistry of **1** was deduced from NOESY correlations of H-17 to H-19, H-20 to H-21, H-1' to H-21, H-3', and H-5', and a  $J$ -value for H-20/H-21 ( $\sim 0$  Hz) as shown in Figure 2.

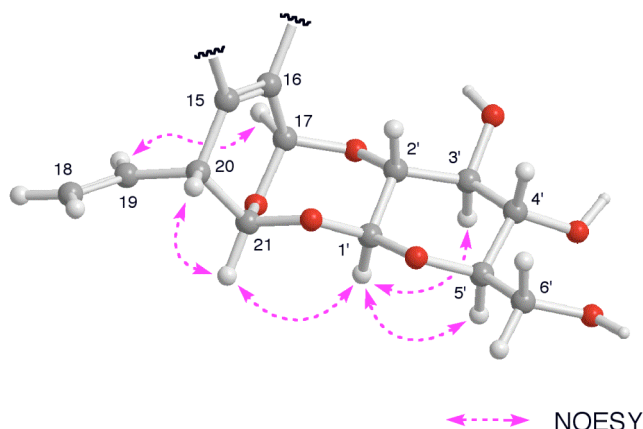


Figure 2. Selected NOESY correlations and relative stereochemistry for a part (C-15~C-21 and C-1'~C-6') of nucleamide F (**1**).

Since the sugar moiety was elucidated to be D-glucopyranose by chiral HPLC analysis of *O*-benzoyl derivatives of the methanolysis products of nucleamide F (**1**),<sup>9</sup> the absolute stereochemistry of nucleamide F (**1**) was assigned as shown in Figure 2.

The absolute stereochemistries of known related monoterpene indole alkaloids **2**~**4**, whose stereochemistries remains unsolved,<sup>6,7</sup> were elucidated as describe below. The absolute configurations at C-3 of **3** and **4** were assigned as both *R* on the basis of the negative Cotton effects at 279 nm ( $\Delta\epsilon$  -0.29) and 253 nm ( $\Delta\epsilon$  -0.72), respectively,<sup>10</sup> while the absolute configurations at C-19 of **2** ~ **4** were elucidated to be *S*, *S*, and *R* on the based of the  $\Delta\delta$  values obtained for (*S*)- and (*R*)-MTPA esters of **2** ~ **4**, respectively<sup>11</sup> (Figure 3).

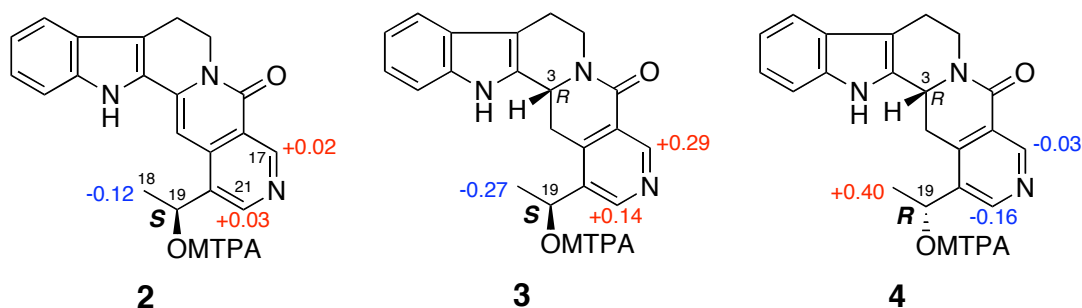


Figure 3.  $\Delta\delta$  values [ $\Delta\delta$  (in ppm)  $\delta_S - \delta_R$ ] obtained for (*S*)- and (*R*)-MTPA esters of compounds **2** ~ **4**.

Naucleamide F (**1**) is a new monoterpene indole alkaloid consisting of a tetrahydro- $\beta$ -carboline ring fused to a pyridone ring, and a 1,3,5-trioxepane ring fused to a dihydropyran ring and a glucose unit. Naucleamide F (**1**) is a rare monoterpene indole alkaloid possessin a glucose unit connected to terpenoid

unit *via* two ether bonds, though an iridoid having a similar unit has been reported from the bark of *Eucommia ulmoides*.<sup>8</sup>

## EXPERIMENTAL

### General Experimental Procedures

Optical rotation was recorded on a JASCO P-1030 polarimeter. IR and UV spectra were recorded on a JASCO FT/IR-5300 spectrophotometers and Shimadzu UV-1600PC, respectively. <sup>1</sup>H, <sup>13</sup>C and 2D NMR spectra were measured on a JEOL JMN-EX400, a JEOL ECA500, and a Bruker AMX-600 spectrometers. The 3.35 and 49.8 ppm resonances of residual CD<sub>3</sub>OD were used as internal references for <sup>1</sup>H and <sup>13</sup>C NMR spectra, respectively. ESI mass spectra were measured on a JEOL JMS-700TZ spectrometer.

### Extraction and Isolation

The bark and wood (300 g) of *Nauclea latifolia* were extracted with MeOH (1.5 L), and the extracts were partitioned between hexane (200 mL x 3) and 90% aqueous MeOH (200 mL). The MeOH layer was partitioned between *n*-BuOH (200 mL x 3) and H<sub>2</sub>O (200 mL). The *n*-BuOH-soluble portions (3.4 g) were subjected to a silica gel column chromatography (CHCl<sub>3</sub>-MeOH, 1:0 → 85:15) to afford fraction **a** (583 mg). Fraction **a** was separated by a C<sub>18</sub> column chromatography (MeOH-H<sub>2</sub>O, 60:40) followed by C<sub>18</sub> HPLC (Capcell Pak RP-18, Shiseido Co. Ltd, 10 x 250 mm; flow rate 2.5 mL/min; UV detection at 210 nm; eluent CH<sub>3</sub>CN/H<sub>2</sub>O, 40:60) to afford naucleamide F (**1**, 0.85 mg, *t*<sub>R</sub> 17 min), angustoline (**2**, 1.3 mg, *t*<sub>R</sub> 42 min), compound 3 (**3**, 1.2 mg, *t*<sub>R</sub> 30 min), and compound 4 (**4**, 0.69 mg, *t*<sub>R</sub> 28 min).

**Naucleamide F (1)**: pale yellow amorphous solid; [ $\alpha$ ]<sub>D</sub><sup>25</sup> +44 (*c* 0.21, MeOH); UV (MeOH)  $\lambda_{\max}$  210 nm (log  $\epsilon$  3.97), 261 (3.44), 289 (3.28), 301 (3.18), and 354 (3.57); IR (KBr) cm<sup>-1</sup>: 3443, 2920, 1645; <sup>1</sup>H- and <sup>13</sup>C-NMR (Table 1); ESI-MS *m/z* 517 (M+Na)<sup>+</sup>; HR-ESI-MS *m/z* 517.1592 (M+Na)<sup>+</sup> (calcd. for C<sub>26</sub>H<sub>26</sub>N<sub>2</sub>O<sub>8</sub>Na, 517.1587).

### Stereochemical assignment of the sugar unit in naucleamide F (1).

Naucleamide F (**1**, 0.1 mg) was treated with 3% HCl/MeOH (300  $\mu$ L) at 110 °C for 1h. After the solvent was removed by nitrogen stream, to the residue was added EtOAc (100  $\mu$ L), and the EtOAc solution was extracted with H<sub>2</sub>O (100  $\mu$ L x 3). The aqueous fraction evaporated in vacuo was treated pyridine (100  $\mu$ L), triethylamine (15  $\mu$ L), and benzoyl chloride (15  $\mu$ L), at rt for 21 h. After addition of MeOH (100  $\mu$ L), the reaction mixture was extracted with *n*-hexane (100  $\mu$ L x 3). The *n*-hexane-soluble fraction was evaporated in vacuo to afford 1-*O*-methyl-2,3,4,6-tetra-*O*-benzoyl derivative of the sugar units of **1**.

Authentic D- and L-glucose were treated with benzoyl chloride as described above to afford 1-*O*-methyl-2,3,4,6-tetra-*O*-benzoyl derivatives of D- and L-glucose, respectively. The 1-*O*-methyl-2,3,4,6-tetra-*O*-benzoyl derivatives were subjected to chiral HPLC analyses using Chiralpak OD-R (Daicel Chemical Industry, Ltd., 4.6 x 250 mm; flow rate 0.5 mL/min; UV detection at 254 nm; eluent MeOH/H<sub>2</sub>O, 95:5). The retention time of 1-*O*-methyl-2,3,4,6-tetra-*O*-benzoyl derivative of methanolysis product of **1** was found to be 18.6 min, while the retention times of authentic 1-*O*-methyl-2,3,4,6-tetra-*O*-benzoyl- $\alpha$ -D-glucopyranose and 1-*O*-methyl-2,3,4,6-tetra-*O*-benzoyl- $\alpha$ -L-glucopyranose were found to be 18.6 and 20.2 min, respectively.

#### Preparation of (*S*)- and (*R*)-MTPA esters of compounds 2~4.

To a solution of **2** (0.1 mg) in CH<sub>2</sub>Cl<sub>2</sub> (100  $\mu$ L) were added (*R*)-MTPACl (0.68 mg), triethylamine (2.0  $\mu$ L), and *N,N*-demethyl-aminopyridine (4.1 mg). The mixture was allowed to stand at rt for 3 h. After evaporation of the solvent, the residue was applied to a silica gel column to give the (*S*)-MTPA ester of **1**. The (*R*)-MTPA ester of **2** and (*S*)- and (*R*)-MTPA esters of **3** and **4** were prepared according to the same procedure as described above.

**(*S*)-MTPA ester of 2:** <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$ 9.40 (H-17), 8.67 (H-21), 6.69 (H-19), 1.82 (H-18); ESIMS *m/z* 548 (M+H)<sup>+</sup>.

**(*R*)-MTPA ester of 2:** <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$ 9.38 (H-17), 8.64 (H-21), 6.40 (H-19), 1.94 (H-18); ESIMS *m/z* 548 (M+H)<sup>+</sup>.

**(*S*)-MTPA ester of 3:** <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$ 9.40 (H-17), 8.67 (H-21), 6.69 (H-19), 1.51 (H-18); ESIMS *m/z* 546 (M+H)<sup>+</sup>.

**(*R*)-MTPA ester of 3:** <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$ 9.11 (H-17), 8.53 (H-21), 6.44 (H-19), 1.78 (H-18); ESIMS *m/z* 546 (M+H)<sup>+</sup>.

**(*S*)-MTPA ester of 4:** <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$ 9.37 (H-17), 8.53 (H-21), 6.69 (H-19), 2.20 (H-18); ESIMS *m/z* 546 (M+H)<sup>+</sup>.

**(*R*)-MTPA ester of 4:** <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$ 9.40 (H-17), 8.69 (H-21), 6.39 (H-19), 1.80 (H-18); ESIMS *m/z* 546 (M+H)<sup>+</sup>.

#### ACKNOWLEDGMENTS

We thank Ms. S. Oka (Center for Instrumental Analysis, Hokkaido University) for measurements of ESIMS. This work was partly supported by a grant from a Grant-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science and Technology of Japan.

## REFERENCES

1. F. Hotellier, P. Delaveau, and J. L. Pousset, *Phytochemistry*, 1975, **14**, 1407.
2. F. Hotellier, P. Delaveau, and J. L. Pousset, *Planta Med.*, 1979, **35**, 242.
3. Z. Zhang, H. N. ElSohly, M. R. Jacob, D. S. Pasco, L. A. Walker, and A. M. Clark, *J. Nat. Prod.*, 2001, **64**, 1001.
4. H. Takayama, O. Ohmori, M. Sakai, M. Funahashi, M. Kitajima, D. Santiarworn, B. Liawruangrath, and N. Aimi, *Heterocycles*, 1998, **49**, 40.
5. H. Shigemori, T. Kagata, H. Ishiyama, F. Morah, A. Ohsaki, and J. Kobayashi, *Chem. Pharm. Bull.*, 2003, **51**, 58.
6. F. Hotellier, P. Delaveau, and J. L. Pousset, *Phytochemistry*, 1975, **14**, 1407.
7. C. A. J. Erdelmeier, U. Regenass, T. Rali, and O. Sticher, *Planta Med.*, 1992, **58**, 43.
8. C. Takamura, T. Hirata, T. Ueda, M. Ono, H. Miyashita, T. Ikeda, and T. Nohara, *J. Nat. Prod.*, 2007, **70**, 1312.
9. J. Kobayashi, T. Kubota, M. Takahashi, M. Ishibashi, M. Tsuda, and H. Naoki, *J. Org. Chem.*, 1999, **64**, 1478.
10. C. M. Lee, W. F. Trager, and A. H. Beckett, *Tetrahedron*, 1967, **23**, 375.
11. I. Ohtani, T. Kusumi, Y. Kashman, and H. Kakisawa, *J. Am. Chem. Soc.*, 1991, **113**, 4092.