

HETEROCYCLES, Vol. 91, No. 7, 2015, pp. 1417 - 1422. © 2015 The Japan Institute of Heterocyclic Chemistry
Received, 30th March, 2015, Accepted, 26th May, 2015, Published online, 4th June, 2015
DOI: 10.3987/COM-15-13216

STRUCTURE AND ABSOLUTE CONFIGURATION OF THE 11-NORIRIDOID “CHAPINGOLIDE”

Matilde Villa-García,^a María Amparo Borja-De-La-Rosa,^b Holber Zuleta-Prada,^a Rubén A. Toscano,^c and Benito Reyes-Trejo^{a*}

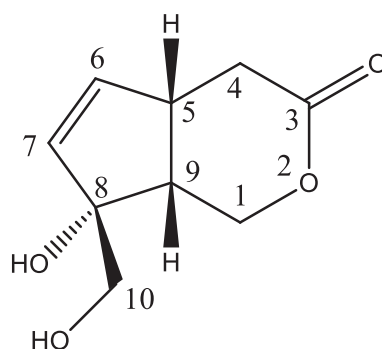
^aLaboratorio de Productos Naturales, Área de Química, Departamento de Preparatoria Agrícola, Universidad Autónoma Chapingo, Apartado 74, Oficina de Correos Chapingo, Km 38.5 Carretera México-Texcoco, Chapingo, México 56230, México, ^bDepartamento de Productos Forestales. División de Ciencias Forestales, Universidad Autónoma Chapingo, Km 38.5 Carretera México-Texcoco, Chapingo, México 56230, México, and ^cInstituto de Química, Universidad Nacional Autónoma de México, Circuito, Exterior, Ciudad Universitaria, Coyoacán 04510 México, D.F, México. Correspondence email: benijovi@yahoo.com.mx, Tel.: +55-513-31108 (ext. 5760)

Abstract – The natural product chapingolide {systematic name: (4a*S*, 7*R*, 7a*R*)-7-hydroxy-7-(hydroxymethyl)-4,4a,7,7a-tetrahydrocyclopenta[*c*]pyran-3(1*H*)-one}, C₉H₁₂O₄, (**1**), is a new 11-*nor*-iridoid isolated from *Calatola mollis* Standl. The colourless compound has a bicyclic structure with an *endo* double bond in the five-membered ring. The absolute configuration was established by mean of the anomalous dispersion of the oxygen scatters present in the structure.

Calatola is a genus of trees in family Icacinaceae. To date, about seven *Calatola* species are recorded in America, and most of them are found in rain forest regions.¹ Approximately, four species of *Calatola* plants were recorded in Mexico¹ of which, *Calatola mollis* Standl is a tree up to 20 m tall. The seeds exhibited potently vomitive-purgative activity.² At present *Calatola mollis* species had been poorly investigated with only botanical descriptions reported to date.¹ Even thought, specimens of this species have been misidentified as *Calatola costaricensis*,^{1,3} and hence a phytochemical study of plants of *Calatola* genus will be useful to establish differences in secondary metabolites composition as a valuable systematic character. The chapingolide (**1**), a *nor*-iridoid, was obtained from the methanolic extract of the leaves of *Calatola mollis* Standl, as a crystalline colourless compound. Its bicyclic structure is closely related to the structures discussed in Valladares and Rios,⁴ and Franzyk and Stermitz.⁵ The main differences reside in the oxidation

level in the *nor*-iridoid skeleton compared with the previous structures, as well as an *endo* double bond in **1**.^{6,7} In order to ascertain unambiguously the relative position of the hydroxyl groups and the double bond in the structure, as well as to establish the absolute configurations of the asymmetric centres, we analyze herein the thus far unreported crystal structure of chapingolide, isolated from *Calatola mollis* Standl. Icacinaceae leaves.

In order to assess the relative frequency of *nor*-iridoids, a search of the Cambridge Structural Database (CSD, Version 5.34)⁸ was undertaken. Surprisingly few relevant structures were found. There were only eleven structures with a similar 11-noriridoid system (*cis*-2-oxabicyclo[4.3.0]nonane) and, similarly, only three structures with *cis*-2-oxabicyclo[4.3.0]nonan-3-one system and fused five membered rings with substitution equivalent to **1**.^{4,5,9} The bond lengths and angles found for **1** (Table 1) closely match the comparable parameters in these literature structures. In order to complete the full characterization of chapingolide and to elucidate its absolute configuration, its crystal structure has been determined.



chapingolide (**1**)

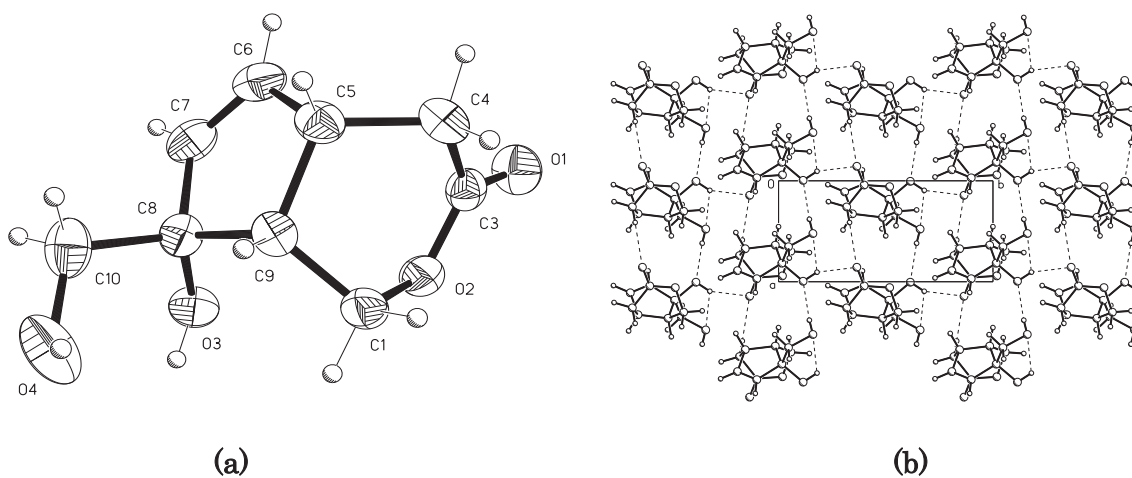
Figure 1a shows an ellipsoid plot of chapingolide. The molecule is made up of a cyclopentene ring fused to a six-membered lactone ring fused showing, a *cis* A/B ring junction, C6=C7 double bond and a *syn* orientation among H-5, H-9 and CH₂-10. In accordance with the biosynthetic origin of the iridoids¹⁰ the *cis* A/B ring junction is α , and **1** corresponds to 8*R*,10-dihydroxy-*cis*-2-oxabicyclo[4.3.0]nonan-6-en-3-one.

The Cremer & Pople parameters¹¹ for each ring are as follows: For pyran ring, $Q = 0.672(2) \text{ \AA}$, $\theta = 90.53(17)^\circ$, and $\varphi = 234.26(18)^\circ$, and for cyclopentene ring, $Q = 0.090(2) \text{ \AA}$, $\varphi = 133.4(15)^\circ$, indicating “intermediate conformations”.

In order to assess in a more quantitative way the real significance of the verbal description for “intermediate conformations”, a quantitative conformational analysis was performed by using the program RICON¹² showing that the pyran ring displays an “intermediate conformation” between boat (80%) and twist (19%) conformation, while the cyclopentene ring poses an “intermediate conformation” between twist (61%) and envelope (39%) conformations. The dihedral angle between cyclopentene and pyran rings is $81.99(12)^\circ$.

Table 1. Geometric parameters (Å, °) for chapingolide

O(1)-C(3)	1.207(3)	C(1)-O(2)	1.459(3)	C(1)-C(9)	1.505(3)
O(2)-C(3)	1.338(3)	C(3)-C(4)	1.496(3)	C(4)-C(5)	1.542(3)
C(5)-C(6)	1.497(3)	C(5)-C(9)	1.556(3)	C(6)-C(7)	1.312(4)
C(7)-C(8)	1.503(3)	C(8)-O(3)	1.433(3)	C(8)-C(10)	1.525(3)
C(8)-C(9)	1.569(2)	C(10)-O(4)	1.412(4)		
O(2)-C(1)-C(9)	112.47(16)	C(3)-O(2)-C(1)	116.51(16)	O(1)-C(3)-O(2)	118.9(2)
O(1)-C(3)-C(4)	124.9(2)	O(2)-C(3)-C(4)	116.22(19)	C(3)-C(4)-C(5)	111.31(17)
C(6)-C(5)-C(4)	112.18(19)	C(6)-C(5)-C(9)	103.86(17)	C(4)-C(5)-C(9)	111.74(17)
C(7)-C(6)-C(5)	113.4(2)	C(6)-C(7)-C(8)	112.82(19)	O(3)-C(8)-C(7)	109.05(16)
O(3)-C(8)-C(10)	108.11(18)	C(7)-C(8)-C(10)	111.39(19)	O(3)-C(8)-C(9)	113.54(16)
C(7)-C(8)-C(9)	103.45(17)	C(10)-C(8)-C(9)	111.28(16)	C(1)-C(9)-C(5)	110.52(17)
C(1)-C(9)-C(8)	114.86(17)	C(5)-C(9)-C(8)	105.65(16)	O(4)-C(10)-C(8)	111.6(2)
C(9)-C(1)-O(2)-C(3)	56.6(2)	C(1)-O(2)-C(3)-C(4)	-8.0(2)	O(2)-C(3)-C(4)-C(5)	-44.4(2)
C(3)-C(4)-C(5)-C(9)	48.6(2)	C(9)-C(5)-C(6)-C(7)	3.8(2)	C(5)-C(6)-C(7)-C(8)	2.1(3)
C(6)-C(7)-C(8)-C(9)	-7.0(2)	O(2)-C(1)-C(9)-C(5)	-46.8(2)	C(4)-C(5)-C(9)-C(1)	-4.0(2)
C(7)-C(8)-C(9)-C(5)	8.8(2)				

**Figure 1.** (a) The molecular structure of (1), ellipsoids drawn at the 50% probability level. (b) Helical chain in the crystal structure of (1), viewed along [001].

As the compound consists only of C, H and O atoms, to establish the absolute configuration of **1** we subject the crystals to a crystallographic analysis by the anomalous X-ray scattering from the oxygen atoms¹³ using Cu K α radiation and by carefully measuring a large number of Bijvoet differences (812 Friedel pairs, 99%). This yielded a refined Flack x parameter¹⁴ and standard uncertainty (s.u.) of 0.18 (25) for the “hole-in-one” fit. We note that the obtained s.u. value is above the suggested upper confidence limit of 0.10 for the determination of the absolute structure of an enantiopure compound.¹³ However, additional confirmation was obtained from the examination of Bayesian statistics on 812 Bijvoet pairs¹⁵ carried out using the program PLATON¹⁶ and suggested that the chapingolide structure is enantiopure: the probability P2 (true) = P3 (true) = 1.000 with P3(racemate-twin) = 0.9×10^{-12} , P3 (false) = 0.2×10^{-59} , G = 0.80(10), and the Hooft parameter $y = 0.10$ (5) for the C8 *R* enantiomer (5*S*, 9*R*). The SHELXL program¹⁷ was used to refine the structure and it reported a Flack parameter of 0.176(252), a G parameter of 1.03(12) and a Hooft parameter of 0.090(49) (Parsons’ method).¹⁸

Regarding the supra-molecular structure of (**1**), there is one significant intermolecular interaction, involving the hydroxyl O3—H3 group as donor and carbonylic atom O1 as acceptor (**Table 2**); connecting neighbouring molecules into a C(8) chain structure¹⁹ running along **b** and built up around the twofold screw axis. These [010] chains are connecting by weak O3—H3...O1 and C4—H4C...O1 interactions. This is observable in Figure 1b and quantitatively assessed by the rather low packing index of 72.7.²⁰ In addition, hydroxyl O3—H3 displays an intramolecular hydrogen bond with the hydroxyl oxygen atom O4, making a 3-center (bifurcate) hydrogen bond (sum of 3 angles about H3 = 359(4) °).

Table 2. Hydrogen bond geometry (Å, °) for chapingolide

$D-H \cdots A$	$D-H$	$H \cdots A$	$D \cdots A$	$D-H \cdots A$
C4—H4B...O3 ⁱ	0.97	2.65	3.362(3)	130
C4—H4C...O1 ⁱⁱ	0.97	2.32	3.221(3)	154
O3—H3...O1 ⁱⁱⁱ	0.84(4)	2.29(3)	2.988(2)	141(3)
O3—H3...O4	0.84(4)	2.34(3)	2.789(3)	114(3)
O4—H4...O3 ⁱⁱ	0.79(5)	2.15(5)	2.939(3)	175(5)

Symmetry codes: (i) $-x, y-1/2, -z+2$; (ii) $x+1, y, z$; (iii) $-x, y+1/2, -z+2$.

EXPERIMENTAL

Optical Rotation was measured on a Perkin-Elmer 343 polarimeter. IR spectra were determined with an Agilent Cary FTIR-ATR 630 spectrophotometer. The ¹³C and ¹H NMR spectra were recorded on an Agilent 400 MR DD2 spectrometer (SantaClara, CA, USA) operating at 100 MHz for ¹³C and 400 MHz for ¹H. The ¹³C and ¹H chemical shifts were measured in CDCl₃ relative to TMS as an internal standard.

HRFAB MS spectrum was measured on a JEOL the MStation JMS-700.

Isolation

Methanol extract (30 g) from the leaves of *C. mollis* (2 kg) was chromatographed over silica gel (200 g) starting with CH₂Cl₂ and increasing the polarity with MeOH. Fractions eluted with CH₂Cl₂/MeOH (95:5) provided 50 mg of a crystalline colourless solid which was submitted to X-ray diffraction study and identified as chapingolide (**1**). For related structures, see: Ban *et al.* (2013); Tada *et al.* (1998).^{6,7}

Chapingolide (1): colourless solid; mp 129-131 °C (CH₂Cl₂/MeOH (95:5)); [α]_D²⁰ +89.4 (*c* 0.17, MeOH); IR (neat) 3439, 1711, 1014 cm⁻¹; ¹H NMR δ 2.56 (dd, *J* = 14, 4 Hz, 1H), 2.68 (dt, *J* = 9, 4 Hz, 1H), 2.74 (dd, *J* = 14, 7 Hz, 1H), 3.35 (m, 1H), 3.50 (d, *J* = 11 Hz, 1H), 3.61 (d, *J* = 11 Hz, 1H), 4.28 (dd, *J* = 14, 4 Hz, 1H), 4.57 (dd, *J* = 14, 3.5 Hz, 1H), 5.82 (br d, *J* = 6 Hz, 1H), 5.85 (br d, *J* = 6 Hz, 1H); ¹³C NMR δ 34.35, 40.56, 41.26, 66.87, 69.27, 86.32, 135.51, 136.00, 172.29; HRFAB MS *m/z* 185.08131 (M+H)⁺ Anal. Calcd for C₉H₁₃O₄.

X-RAY STRUCTURE DETERMINATION OF 1

A colorless prism crystal of C₉H₁₂O₄ having approximate dimensions of 0.41 × 0.17 × 0.04 mm was mounted on a glass fiber. All measurements were made on a Bruker Venture κ -diffractometer using Cu-K α radiation monochromated by multilayer mirror. Cell constants and an orientation matrix for data collection corresponded to a primitive monoclinic cell with dimensions: *a* = 5.1034(7) Å, *b* = 10.8665(15) Å, *c* = 7.6481(10) Å, β = 92.595(6) °, *V* = 423.7(1) Å³. The crystal belongs to the space group *P*2₁. Of the 17267 reflections that were collected, 1729 were unique (*R*_{int} = 0.030); equivalent reflections were merged. Data were collected and processed using APEX2 (Bruker). The structure was solved by direct methods and expanded using Fourier techniques. Deposition number CCDC-1056178 for compound No. 1. Free copies of the data can be obtained via <http://www.ccdc.cam.ac.uk/conts/retrieving.html> (or from the Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge, CB2 1EZ, UK; Fax: +44 1223 336033; e-mail: deposit@ccdc.cam.ac.uk).

ACKNOWLEDGEMENTS

We are grateful to Dr. Enrique Guizar Nolasco for collecting the plant material and Dr. Patricia Vera Calleti, Área de Biología, Departamento de Preparatoria Agrícola, Universidad Autónoma Chapingo, for the identification of the plant material and CONACyT (México) for stimulating support.

REFERENCES

1. P. Vera-Caletti and T. Wendt, *Acta Botánica Mexicana*, 2001, **54**, 39.
2. M. Martínez, 'Las Plantas Medicinales de México', 4th edition, Ediciones Botas, México D.F., México, 1959.
3. The Red List of Mexican Cloud Forest Trees, ICACINACEAE, ed. by M. González-Espinosa, J. A. Meave, G. G. Lorea-Hernández, G. Ibarra-Manríquez, and A. C. Newton, Fauna & Flora International, Cambridge, UK, 2011, p. 38.
4. M. G. Valladares and M. Y. Ríos, *J. Nat. Prod.*, 2007, **70**, 100.
5. H. Franzyk and F. R. Stermitz, *J. Nat. Prod.*, 1999, **62**, 1646.
6. N. K. Ban, V. H. Giang, T. M. Linh, L. Q. Lien, N. T. Ngoc, N. H. Nam, N. X. Cuong, P. V. Kiem, C. V. Minh, and D. T. Do Thi Thao, *Phytochem. Lett.*, 2013, **6**, 267.
7. M. Tada, S. Inoue, T. Miki, S. Onogi, J. Kaminaga, J. Hiraoka, K. Kitano, and K. Chiba, *Chem. Pharm. Bull.*, 1998, **46**, 1451.
8. F. H. Allen, *Acta Cryst.*, 2002, **B58**, 380.
9. J.-G. Huang, L.-J. Zhou, and H.-H. Xu, *Helv. Chim. Acta*, 2011, **94**, 327.
10. V. Plouvier and J. Favre-Bonvin, *Phytochemistry*, 1971, **10**, 1697.
11. D. Cremer and J. A. Pople, *J. Am. Chem. Soc.*, 1975, **97**, 1354.
12. A. Y. Zotov, V. A. Palyulin, and N. S. Zefirov, *J. Chem. Inf. Comput. Sci.*, 1997, **37**, 766.
13. H. D. Flack and G. Bernardinelli, *J. Appl. Cryst.*, 2000, **33**, 1143.
14. H. D. Flack, *Acta Cryst.*, 1983, **A39**, 876.
15. R. W. W. Hooft, L. H. Straver, and A. L. Spek, *J. Appl. Cryst.*, 2008, **41**, 96.
16. A. L. Spek, *Acta Cryst.*, 2009, **D65**, 148.
17. G. M. Sheldrick, *Acta Cryst.*, 2008, **A64**, 112.
18. S. Parsons, H. D. Flack, and T. Wagner, *Acta Cryst.*, 2013, **B69**, 249.
19. J. Bernstein, R. E. Davis, L. Shimoni, and N.-L. Chang, *Angew. Chem., Int. Ed. Engl.*, 1995, **34**, 1555.
20. A. I. Kitaigorodsky, 'Molecular Crystals and Molecules', Academic Press, London, 1973.