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## A NEW MONOTERPENOID INDOLE ALKALOID FROM *UNCARIA RHYNCHOPHYLLA*

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**Abstract** – A new alkaloid, uncanidine A (**1**) and 14 known alkaloids (**2-15**) were isolated from the hook-bearing branches of *Uncaria rhynchophylla*. Their structures were elucidated by spectroscopic analyses, including 1D-, 2D-NMR, HR-ESI-MS, ECD, as well as comparison with the data reported in the literature. Noteworthy, uncanidine A (**1**) was a novel *Uncaria* alkaloid which possessed a 6/5/6/6/5 hexacyclic ring system. In addition, uncanidine A (**1**) was found to inhibit the generation of NO from RAW 264.7 cells stimulated by LPS.

Monoterpenoid indole alkaloids (MIAs) are well-known for their significant biological effects. The liana *Uncaria rhynchophylla* (Rubiaceae), widely distributed in South China, especially in the Guangdong, Fujian and Yunnan Provinces, is a rich source of MIAs known as *Uncaria* alkaloids.<sup>1</sup> The dried branches of *Uncaria rhynchophylla* (Miq.) Miq. ex Havil. is one of the most important traditional Chinese herbs, named “Gouteng” in Chinese, which has long been used in traditional Chinese medicine to relieve hypertension, epilepsy, headaches and dizziness.<sup>2-4</sup> From this plant, around 300 chemical constituents have been reported, covering alkaloids, flavonoids, triterpenoids, *etc.*, of which MIAs are regarded as the predominantly bioactive constituents.<sup>3,5-7</sup> Modern pharmacological studies have shown that *Uncaria* alkaloids has biological activities such as lowering blood pressure, anti-convulsant, antiarrhythmia, sedative, and antithrombotic.<sup>3,7-9</sup> In a search for structurally diverse and bioactive alkaloids, we carried out a phytochemical investigation on the total alkaloid fraction of the hook-bearing branches of *Uncaria rhynchophylla*. A new alkaloid uncanidine A (**1**) and fourteen known ones were obtained by various



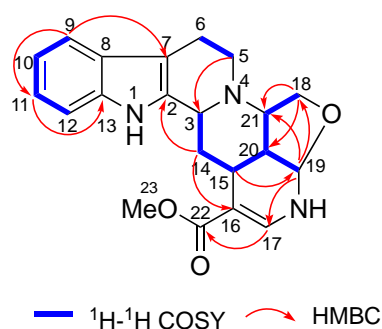
oxygenated olefinic proton [ $\delta_{\text{H}}$  7.52 (1H, s)], and seven methylene and methine proton signals possibly associated with heteroatoms [ $\delta_{\text{H}}$  4.74 (1H, d,  $J = 5.2$  Hz), 4.36 (1H, dd,  $J = 9.0, 2.8$  Hz), 3.57 (1H, dd,  $J = 9.0, 2.8$  Hz), 4.16 (1H, m), 3.98 (1H, ddd,  $J = 9.7, 7.3, 2.5$  Hz), 3.17 (1H, m), 2.89 (1H, m)] were observed. The  $^{13}\text{C}$  NMR spectrum of **1** showed 21 carbon signals, one indole moiety including eight carbon signals ( $\delta_{\text{C}}$  138.1, 107.4, 128.4, 118.4, 119.7, 121.7, 111.9, 138.1), one ester carbonyl ( $\delta_{\text{C}}$  170.5), one double bond ( $\delta_{\text{C}}$  143.0, 101.9), and five carbon signals possibly attached to heteroatom ( $\delta_{\text{C}}$  63.8, 63.4, 52.4, 50.8, 51.3). Comparison of the MS and NMR data (Table 1) of **1** with those of the known compounds ajmalicine and akuammigine<sup>19</sup> revealed that their NMR signals were similar except for the absence of the terminal methyl signals, and the existence of signals for a methylene [ $\delta_{\text{C}}$  63.4,  $\delta_{\text{H}}$  3.57 (1H, dd,  $J = 9.0, 2.8$  Hz), 4.36 (1H, dd,  $J = 9.0, 2.8$  Hz), CH<sub>2</sub>-18]. In addition, the chemical shift of C-17 ( $\delta_{\text{C}}$  143.0) was obvious different from those in ajmalicine ( $\delta_{\text{C}}$  154.5) and akuammigine ( $\delta_{\text{C}}$  154.8). At the same time, the molecular weight of **1** is odd, suggesting that it contains three nitrogen atoms, which is one more than those of ajmalicine and akuammigine.

**Table 1.**  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR spectroscopic data of **1**

| No. | $\delta_{\text{C}}^{\text{a)}$ | $\delta_{\text{H}}^{\text{b)}$  |
|-----|--------------------------------|---|
| 2   | 138.1                          | -   |
| 3   | 50.8                           | 4.16 (1H, m)  |
| 5   | 52.4                           | $\alpha$ 3.17 (1H, m)<br>$\beta$ 2.89 (1H, m)   |
| 6   | 22.6                           | $\alpha$ 2.98 (1H, m)<br>$\beta$ 2.69 (1H, m)   |
| 7   | 107.4                          | -   |
| 8   | 128.4                          | -   |
| 9   | 118.4                          | 7.37 (1H, d, $J = 7.8$ Hz)  |
| 10  | 119.7                          | 6.96 (1H, t, $J = 7.8$ Hz)  |
| 11  | 121.7                          | 7.02 (1H, t, $J = 7.8$ Hz)  |
| 12  | 111.9                          | 7.27 (1H, d, $J = 7.8$ Hz)  |
| 13  | 138.1                          | -   |
| 14  | 31.1                           | $\alpha$ 2.16 (1H, m)<br>$\beta$ 1.95 (1H, m)   |
| 15  | 24.3                           | 3.13 (1H, m)  |
| 16  | 101.9                          | -   |
| 17  | 143.0                          | 7.52 (1H, s)  |
| 18  | 63.4                           | $\alpha$ 3.57 (1H, dd, $J = 9.0, 2.8$ Hz)<br>$\beta$ 4.36 (1H, dd, $J = 9.0, 2.8$ Hz) |
| 19  | 84.8                           | 4.74 (1H, d, $J = 5.2$ Hz)  |
| 20  | 36.7                           | 2.39 (1H, m)  |
| 21  | 63.8                           | 3.98 (1H, ddd, $J = 9.7, 7.3, 2.5$ Hz)  |
| 22  | 170.5                          | -   |
| 23  | 51.3                           | 3.35 (3H, s)  |

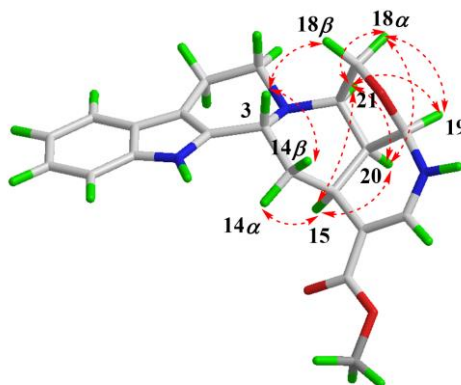
<sup>a)</sup> Measured at 175 MHz. <sup>b)</sup> Measured at 700 MHz.

The  $^1\text{H}$ - $^1\text{H}$  COSY spectrum of **1** revealed the presence of three spin systems as shown in Figure 2. In the HMBC spectrum, the correlations between H-15 and C-19, between H-19 and C-17/C-18/C-21, between H-18 and C-20/C-21/C-19 suggested the presence of a  $\text{CH}_2$  adjacent to C-21. According to the downfield chemical shifts of  $\text{CH}_2$ -18/ $\text{CH}$ -19, upfield chemical shift of CH-17, and the molecular weight information, it can be inferred that  $\text{CH}_2$ -18 and CH-19 may be linked to oxygen atom and form a five-membered heterocyclic ring with CH-20 and CH-21, as well as CH-17 may be linked to nitrogen atom rather than oxygen. Based on the above evidence, the planar structure of **1** was assigned as shown in Figure 2.



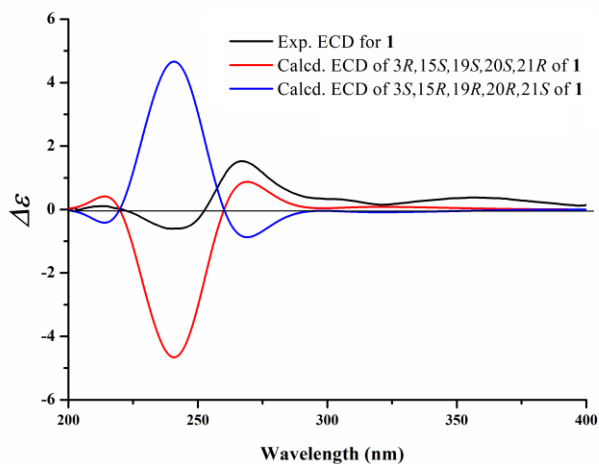
**Figure 2.** Key  $^1\text{H}$ - $^1\text{H}$  COSY and HMBC correlations of **1**

In the NOESY spectrum, the correlations between H-3 ( $\delta_{\text{H}}$  4.16) and H-14 $\beta$  ( $\delta_{\text{H}}$  1.95)/H-18 $\beta$  ( $\delta_{\text{H}}$  4.36), between H-15 ( $\delta_{\text{H}}$  3.13) and H-14 $\alpha$  ( $\delta_{\text{H}}$  2.16)/H-20 ( $\delta_{\text{H}}$  2.39)/H-21 ( $\delta_{\text{H}}$  3.98), between H-19 ( $\delta_{\text{H}}$  4.74) and H-18 $\alpha$  ( $\delta_{\text{H}}$  3.57)/H-21 ( $\delta_{\text{H}}$  3.98), between H-20 ( $\delta_{\text{H}}$  2.39) and H-15 ( $\delta_{\text{H}}$  3.13)/H-21 ( $\delta_{\text{H}}$  3.98) indicated the relative configuration of **1** as shown in Figure 3. However, the orientation of H-20 could not be deduced directly from insufficient evidence in NOESY spectrum. In order to determine the orientation of H-20 and further confirmed the structure of **1**, models of the *s*-isomer and *r*-isomer (Figure S1) of **1** were investigated by quantum chemical calculation. The calculation  $^{13}\text{C}$  NMR chemical shifts of **1** (*s*-isomer) at the B3LYP/6 31 G (2d, p) level with the PCM in  $\text{CD}_3\text{OD}$  agreed well with the experimental data (Figure S2), a correlation coefficient ( $R^2$ ) of 0.9992 (Figure S3), indicating the structure with relative configuration of **1** shown in Figure 3 was rational.



**Figure 3.** Key NOESY correlations of **1**

To determine the absolute configuration of **1**, ECD curves for the two possible enantiomers (3*R*, 15*S*, 19*S*, 20*S*, 21*R*-**1** and 3*S*, 15*R*, 19*R*, 20*R*, 21*S*-**1**) were calculated using the TD-DFT method. As shown in Figure 4, the experimental ECD spectrum of **1** showed a negative cotton effect at 243 nm and positive cotton effects at 265 nm, which were similar to the calculated one for the isomer with 3*R*, 15*S*, 19*S*, 20*S* and 21*R* configuration. Therefore, the absolute stereochemistry of **1** was assigned as 3*R*, 15*S*, 19*S*, 20*S* and 21*R*.



**Figure 4.** Experimental and calculated ECD spectra of **1**

Uncanidine A (**1**) was evaluated for its NO inhibitory effects in LPS-stimulated RAW 264.7 cells. The results showed that **1** could inhibit the generation of NO from RAW 264.7 cells stimulated by LPS.

## EXPERIMENTAL

**General experimental procedures.** Optical rotations were determined on a JASCO P-2000 polarimeter. IR and UV spectra were measured on Nicolet 5700 and TU-1901 UV/Vis spectrophotometers, respectively. NMR spectra were recorded on a Bruker Avanced spectrometer at 700 MHz and 400 MHz, with TMS as an internal standard. HR-ESI-MS was acquired on a Waters Xevo-G2-S Q-TOF mass spectrometer. CD spectra were recorded with a Chirascan V100 spectropolarimeter. For column chromatography (CC), silica gel (200-300 mesh) was obtained from Qingdao Ocean Chemical Plant of Qingdao, China, and Sephadex LH-20 was obtained from Pharmacia Company of Switzerland. Semi-preparative HPLC analysis was carried out on an Agilent 1260 series system equipped with a diode array detector. TLC silica gel GF<sub>254</sub> plates were purchased from Qingdao Ocean Chemical Plant and were visualized under a UV lamp at 254 nm or by spraying the Dragendorff's reagent.

**Plant material.** The hook-bearing branches of *Uncaria rhynchophylla* (Miq.) Miq. ex Havil. (10.0 kg) were collected by Yanrong Zhang (Anguo Tongde Medicinal Materials, Hebei, China) from Hunan, China in September 2018, and were identified by Professor Yaojun Yang (Beijing University of Chinese

Medicine). A voucher specimen was deposited in the School of Traditional Chinese Materia Medica, Beijing University of Chinese Medicine.

**Extraction and isolation.** The powdered hook-bearing branches of *Uncaria rhynchophylla* (10.0 kg) were extracted with 90% EtOH (3 × 2 h). The extract was suspended in H<sub>2</sub>O (6 L) and acidified with 10% HCl to pH 2, which was further partitioned with EtOAc to remove the neutral components. The aqueous layer was then basified with ammonia to pH 10 and re-extracted with CH<sub>2</sub>Cl<sub>2</sub> to obtain a total alkaloid fraction (9.2 g). The alkaloid fraction was separated continuously by column chromatography over silica gel, Sephadex LH-20, ODS and HPLC to yield **1** (2.5 mg). Extraction and isolation of **2-15** see Supporting Information.

**Uncanidine A (1):** yellow amorphous powder;  $[\alpha]_D^{25} + 66.7$  (*c* 0.06, MeOH); UV (MeOH)  $\lambda_{\max}$  (log  $\epsilon$ ): 210 (4.27), 254 (2.89) nm; IR (KBr)  $\nu_{\max}$ : 3267, 2921, 2850, 1707, 1620, 1547, 1518, 1454, 1383, 1300, 1261, 1213, 1066, 918, 874, 703, 755 cm<sup>-1</sup>; HRESIMS *m/z*: 366.1813 [M+H]<sup>+</sup> (Calcd for C<sub>21</sub>H<sub>24</sub>N<sub>3</sub>O<sub>3</sub><sup>+</sup>, 366.1812); CD (MeCN,  $\Delta\epsilon$ )  $\lambda_{\max}$  243 (-1.19), 267 (+2.31), 300 (+0.50); <sup>1</sup>H NMR (CD<sub>3</sub>OD, 700 MHz) and <sup>13</sup>C NMR data (CD<sub>3</sub>OD, 175 MHz) are shown in Table 1.

**In vitro anti-inflammatory Assay.** The inflammatory assay was employed to evaluate the activity of **1** related to the release of NO from macrophage cells as the literature reported.<sup>21,22</sup> Detailed experiments and results see Supporting Information.

## ACKNOWLEDGEMENTS

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