

HETEROCYCLES, Vol. 104, No. 9, 2022, pp. 1661 - 1667. © 2022 The Japan Institute of Heterocyclic Chemistry  
Received, 3rd July, 2022, Accepted, 15th July, 2022, Published online, 20th July, 2022  
DOI: 10.3987/COM-22-14713

## NEW ANTI-TMV ISOCHROMENES FROM *NICOTIANA TABACUM*-DERIVED ENDOPHYTIC FUNGUS *ASPERGILLUS VERSICOLOR*

Qiu-Fen Hu,<sup>1</sup> Ling-Fang Zhang,<sup>1</sup> Guang-Hai Zhang,<sup>2</sup> Mei-Fen Bao,<sup>3</sup> Yin-Ke Li,<sup>1</sup> Dong Miao,<sup>2</sup> Yu-Ping Wu,<sup>2</sup> Gang Du,<sup>1\*</sup> and Guang-Hui Kong<sup>2\*</sup>

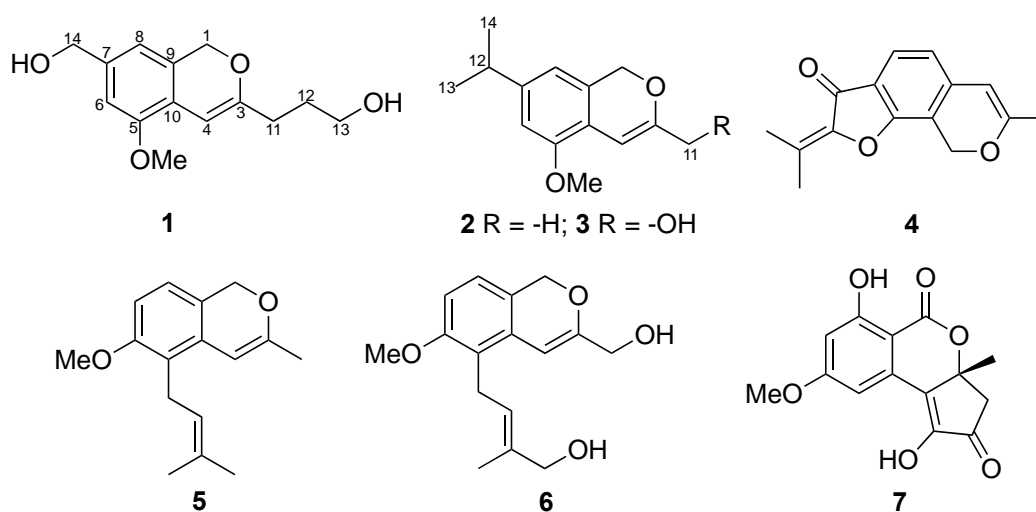
<sup>1</sup>Key Laboratory of Chemistry in Ethnic Medicinal Resources, Yunnan Minzu University, Kunming 60500, P. R. China. E-mail: 18372454@qq.com; <sup>2</sup> Yunnan Academy of Tobacco Agricultural Sciences, Kunming, Yunnan 650031, P. R. China, E-mail: 13908776036@163.com. <sup>3</sup>Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650201, P. R. China.

**Abstract** – Three new isochromenes, versicolols G-I (**1-3**), together with four known analogues (**4-7**) were isolated from the fermentation products of a *Nicotiana tabacum*-derived endophytic fungus *Aspergillus versicolor*. Their structures were elucidated by spectroscopic methods, including extensive <sup>1</sup>H, <sup>13</sup>C, and 2D-NMR techniques. Compounds **1-3** were also tested for their anti-tobacco mosaic virus (anti-TMV) activities, and the results revealed that compound **1** exhibited high anti-TMV activity with inhibition rate of 41.6%, and this rate are higher than that of positive control. Compounds **2** and **3** also showed potential anti-TMV activities with inhibition rates of 22.8 and 26.5%, respectively.

Endophytic fungi are prospective producers of an abundant source of bioactive compounds with potential for exploitation in a wide variety of medical areas.<sup>1,2</sup> Among them, the *Aspergillus* is an important fungal genus containing economically important species, as well as pathogenic species of animals and plants.<sup>3,4</sup> They also can produce a number of structurally complicated molecules with various biological activities.<sup>5,6</sup> In our previous works, some bioactive metabolites, such as, diterpenoids,<sup>7</sup> alkaloids,<sup>8</sup> butyrolactones,<sup>9-11</sup> isocoumarins,<sup>12-14</sup> and the like, had been isolated from the genus of this fungus. Particularly, some isocoumarins, such as oryzaeins B<sup>14</sup> with inhibition rate of 30.6%, versicoumarins A<sup>15</sup> with inhibition rate of 28.6%, versicolols D<sup>16</sup> with inhibition rate of 24.6%, have potential anti-tobacco mosaic virus (anti-TMV) activities.

The isocoumarins are an important class of naturally occurring coumarins isomers with a reversed lactone moiety, which are abundantly distributed in natural sources and are being extracted from different plants,

molds, lichens, fungi and bacteria strains.<sup>17</sup> As one of the characteristic components of *Aspergillus* fungi, isochromenes also attracts much attention from chemists and biologists due to their diverse structures and biological properties.<sup>18,19</sup> With the aim of continuously explore bioactive metabolites from *Aspergillus* fungi species, an investigations were carried out on the cultures of the endophytic fungi *Aspergillus versicolor* obtained from *Nicotiana tabacum*. As a result, three new isochromenes (**1-3**), together with four known analogues (**4-7**) were obtained. This paper describes the elucidation of the structures of these three new compounds, and a preliminary evaluation of their anti-TMV activities.

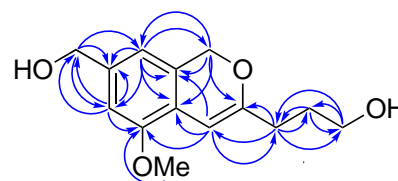


**Figure 1.** The structures of isochromenes and their analogues from fungus *A. versicolor*

The whole culture broth from the fermentation products of *A. versicolor* was extracted with EtOAc. The extract was subjected repeatedly to column chromatography on silica gel, MCI, RP-18 and preparative HPLC to afford compounds **1-7**, including three new isochromenes, versicolols G-I (**1-3**), along with four known analogues (**4-7**). The structures of compounds **1-7** were shown in Figure 1, and the NMR data of **1-3** were listed in Table 1. The known compounds, ustusorane B (**4**),<sup>20</sup> versicolols B (**5**),<sup>14</sup> oryzaein D (**6**),<sup>13</sup> and altenusin D (**7**)<sup>21</sup> were identified by the comparison of their spectroscopic data with literatures.

Compound **1** was obtained as a pale-yellow gum. Its molecular formula  $C_{14}H_{18}O_4$  was determined by its positive HRESIMS showed a peak at  $m/z$  273.1012  $[M+Na]^+$  (calcd  $C_{14}H_{18}NaO_4$  for 273.1103), indicating seven degrees of unsaturations. The IR spectrum

showed the absorption bands for hydroxyl ( $3410\text{ cm}^{-1}$ ) and aromatic group ( $1616$ ,  $1560$ , and  $1462\text{ cm}^{-1}$ ). The UV spectrum exhibited absorption bands at 215, 270, and 338 nm, also suggested the existence of aromatic chromophore. The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectrum of **1** (Table 1) showed the presence of a 1,2,3,5-tetrasubstituted benzene ring (C-5~C-10, H-6, and H-8), a hydroxypropyl group<sup>14</sup> (C-11~C-13,



**Figure 2.** Key HMBC (  $\curvearrowright$  ) correlations of **1**

H<sub>2</sub>-11, H<sub>2</sub>-11~H<sub>2</sub>-13'), a pair of double bond (C-3, C-4, and H-4), an oxidized methylene carbon (C-1 and H<sub>2</sub>-1), a hydroxymethyl group (C-14 and H<sub>2</sub>-14), and a methoxy group ( $\delta_C$  55.8 q and  $\delta_H$  3.77 s). According to above NMR data, the double bonds and oxidized methylene carbon should be incorporated with benzene ring to form an isochromene ring to support the seven degrees of unsaturations in the molecule.<sup>13</sup> In addition, the existence of isochromene core was supported by the HMBC correlations (Figure 2) from H-1 to C-3, C-8, C-9, C-10, from H-4 to C-3, C-5, C-9, C-10, and from H-8 to C-1, and the existence 3-hydroxypropyl group was also supported by the HMBC correlations from H<sub>2</sub>-13 to C-11, C-12, from H<sub>2</sub>-11 to C-12, C-13, and from H<sub>2</sub>-12 to C-11, C-13.

**Table 1.** <sup>1</sup>H NMR and <sup>13</sup>C NMR data for compounds **1-3**

No.	Compound <b>1</b>		Compound <b>2</b>		Compound <b>3</b>	
	$\delta_C$ (m)	$\delta_H$ (m, J, Hz)	$\delta_C$ (m)	$\delta_H$ (m, J, Hz)	$\delta_C$ (m)	$\delta_H$ (m, J, Hz)
1	75.6 t	5.21 s	74.6 t	5.19 s	75.2 t	5.20 s
3	155.5 s		152.7 s		150.9 s	
4	96.7 d	5.84 s	96.4 d	5.84 s	96.8 d	5.87 s
5	156.9 s		156.9 s		156.5 s	
6	110.3 d	6.76 (d) 1.8	110.4 d	6.73 (d) 1.8	110.5 d	6.73 (d) 1.8
7	140.3 s		144.3 s		144.4 s	
8	117.7 d	6.82 (d) 1.8	118.6 d	6.89 (d) 1.8	118.5 d	6.90 (d) 1.8
9	136.3 s		135.3 s		135.0 s	
10	119.5 s		119.3 s		119.3 s	
11	31.5 t	2.26 (t) 7.6	20.7 q	1.93 s	62.2 t	4.33 s
12	28.4 t	1.62 m	31.9 d	3.06 m	31.9 d	3.04 m
13	64.2 t	3.55 (t) 6.6	23.1 q	1.52 (d) 6.8	23.1 q	1.53 (d) 6.8
14	66.5 t	4.65 s	23.1 q	1.52 (d) 6.8	23.6 q	1.53 (d) 6.8
-OMe	55.8 q	3.77 s	55.9 q	3.79 s	55.9 q	3.76 s

Since the isochromene skeleton was determined, the positions of substituents (hydroxypropyl, hydroxymethyl, and methoxy groups) also can be determined by further analysis of its HMBC data (Figure 2). The HMBC correlations from the H<sub>2</sub>-11 to C-3, C-4, from H-4 to C-11 established that the 3-hydroxypropyl group was located at C-3. The methoxy group located at C-5 was clearly indicated by the HMBC correlations from methoxy proton ( $\delta_H$  3.77 s) to C-5. Furthermore, the hydroxymethyl located at C-7 can also be determined by the HMBC correlations from H<sub>2</sub>-14 to C-6, C-7, C-8, and from H-6 and H-8 to C-7. On the basis of above evidence, the structure of **1** was established as 3-(7-(hydroxymethyl)-5-methoxy-1*H*-isochromen-3-yl)propan-1-ol, and gave the trivial name of versicolol G.

7-Isopropyl-5-methoxy-3-methyl-1*H*-isochromene (**2**) was also obtained as pale-yellow gum with a molecular formula of  $C_{14}H_{18}NaO_2$ , according to the ion peak of ( $m/z$  241.1210,  $[M+Na]^+$ ) in the HRESIMS. The UV and IR spectra of **2** were highly similar to those of **1**. The chemical shift differences in NMR resulted from the disappearance of 3-hydroxypropyl and hydroxymethyl groups, and appearance of an isopropyl<sup>22</sup> (C-12~C-14, H-12, H<sub>6</sub>-13,14) and methyl group (C-11 and H<sub>3</sub>-11) in **2**. These changes indicated that the hydroxypropyl and hydroxymethyl groups in **1** were converted into isopropyl and methyl groups in **2**. The HMBC correlation from H-12 to C-6, C-7, C-8, from H<sub>3</sub>-11 to C-3, C-4, from H-4 to C-11 also supported the isopropyl group located at C-7, and the methyl group located at C-3, respectively. In addition, the positions of methoxy groups can also be determined by the HMBC correlations between the methoxy proton ( $\delta_H$  3.79 s) and C-5. The structure of **2** was therefore defined, and given the trivial name of versicolol H.

(7-Isopropyl-5-methoxy-1*H*-isochromen-3-yl)methanol (**3**) was obtained as a pale-yellow gum and showed a quasi-molecular ion at  $m/z$  257.1148  $[M+Na]^+$  in the HRESIMS (calcd  $m/z$  257.1154), corresponding to the molecular formula of  $C_{14}H_{18}O_3$ . The <sup>1</sup>H and <sup>13</sup>C NMR spectra of **3** were also highly similar to those of **2**. The obvious chemical shift differences resulted from the methyl group in **2** being replaced by a hydroxymethyl group in **3**. The position of the hydroxymethyl group at C-3 can be determined by the HMBC correlations from H<sub>2</sub>-11 to C-3, C-4, from H-4, to C-11. In addition, the positions of isopropyl and methoxy groups can also be determined by further analysis of their HMBC correlations. Thus, the structure of **3** was determined as shown, and given the trivial name of versicolol I. Since certain of the isocoumarin derivatives exhibit potential anti-virus activities.<sup>12,13,15,16,23</sup> Compounds **1-3** were evaluated for their anti-TMV activities. The anti-TMV activities were tested by half-leaf method,<sup>8,24</sup> using 20  $\mu$ M of ningnanmycin as a positive control. The results showed that compound **1** exhibited high anti-TMV activity with an inhibition rate of 41.6% at the concentration of 20  $\mu$ M, and this rate is higher than that of the positive control. Compounds **2** and **3** also showed potential activities with inhibition rates of 22.8 and 26.5% at the concentration of 20  $\mu$ M, respectively.

## EXPERIMENTAL

**General Experimental Procedures.** UV spectra were obtained using a Shimadzu UV-1900 spectrophotometer. A Bio-Rad FTS185 spectrophotometer was used for scanning IR spectra. <sup>1</sup>H, <sup>13</sup>C, and 2D-NMR spectroscopic data were recorded on a DRX-500 NMR spectrometer with TMS as internal standard. ESIMS and HRESIMS analyses were measured on Agilent 1290 UPLC/6540 Q-TOF mass spectrometer. Chemical shifts ( $\delta$ ) are expressed in ppm with reference to the TMS signal. Semi-preparative HPLC was performed on an Agilent 1260 preparative liquid chromatograph with Zorbax PrepHT GF C<sub>18</sub> (2.12 mm×25 cm) or Venusil MP C<sub>18</sub> (2.0 mm×25 cm) columns. Column

chromatography was performed using silica gel (200-300 mesh, Qing-dao Marine Chemical, Inc., Qingdao, China), Sephadex LH-20 (Sigma-Aldrich, Inc, USA), or MCI gel (75-150  $\mu\text{m}$ , Mitsubishi Chemical Corporation, Tokyo, Japan). Column fractions were monitored by TLC visualized by spraying with 5%  $\text{H}_2\text{SO}_4$  in ethanol and heating.

**Fungal material.** The culture of *Aspergillus versicolor* YNCA-20-25 was isolated from the leave of cigar tobacco, collected from Yingjiang County, Dehong Prefecture, Yunnan Province, in 2020. The strain was identified by one of authors (Dr. Gang Du) based on the analysis of the ITS sequence. It was cultivated at room temperature for 7 days on potato dextrose agar at 28 °C. Agar plugs were inoculated into 250 mL Erlenmeyer flasks each containing 100 mL potato dextrose broth and cultured at 28 °C on a rotary shaker at 180 rpm for five days. large scale fermentation was carried out in 100 Fernbach flasks (500 mL) each containing 300 mL of medium (glucose 5%; peptone 0.15%; yeast 0.5%;  $\text{KH}_2\text{PO}_4$  0.05%;  $\text{MgSO}_4$  0.05% in 1 L of deionized water; pH 6.5 before autoclaving). Each flask was inoculated with 5.0 mL of cultured broth and incubated at 27 °C for 20 days.

**Extraction and Isolation.** The whole culture broth of *A. versicolor* was extracted four times with EtOAc (4 $\times$ 10 L) at room temperature and filtered. The crude extract (108 g) was applied to silica gel column chromatography, eluting with a  $\text{CHCl}_3$ -MeOH gradient system (9:1, 8:2, 7:3, 6:4, 5:5). Five fractions were obtained from the silica gel column and individually decolorized on MCI gel to yield fractions A-E. The further separation of fraction A (9:1, 12.5 g) by silica gel column chromatography, eluted with  $\text{CHCl}_3$ -acetone (9:1, 8:2, 7:3, 6:4, 1:1) again, yielded mixtures A1-A5. Fraction A2 (8:2, 2.68 g) was subjected to RP-18 column chromatography and HPLC (MeOH/ $\text{H}_2\text{O}$  50:50-80:20 gradient) to give **4** (21.8 mg) and **5** (18.0 mg). Fraction A3 (7:3, 2.87 g) was subjected to RP-18 column chromatography and HPLC (MeOH/ $\text{H}_2\text{O}$  40:60-75:25 gradient) to give **2** (23.5 mg) and **3** (20.2 mg), and **6** (18.5 mg). The further separation of fraction B (8:2, 9.26 g) by silica gel column chromatography, eluted with  $\text{CHCl}_3$ -acetone (8:2, 7:3, 6:4, 1:1), yielded mixtures B1-B4. Fraction B3 (6:4, 2.38 g) was subjected to RP-18 column chromatography and HPLC (MeOH/ $\text{H}_2\text{O}$  30:70-40:60 gradient) to give **1** (18.2 mg) and **7** (15.8 mg).

**Anti-TMV Assays.** The anti-TMV activities were tested using the half-leaf method according our previous literature,<sup>8,24</sup> and 20  $\mu\text{M}$  of Ningnanmycin ( $\text{C}_{16}\text{H}_{25}\text{N}_7\text{O}_8$ , CAS#: 156410-09-2, a commercial new cytosine nucleoside peptide antibiotics for plant viral diseases in China, with inhibition rate of 34.2%) was used as a positive control. The virus was inhibited by mixing with the solution of tested compounds (20  $\mu\text{M}$  in DMSO). After 30 min, the mixture was inoculated on the left side of the leaves of *N. glutinosa*, whereas the right side of the leaves was inoculated with the mixture of DMSO solution and the virus as control. The local lesion numbers were recorded 3-4 days after inoculation. Three repetitions were conducted for each compound. The inhibition rates were calculated according to the formula:

$$\text{inhibition rate (\%)} = [(C-T) / C] \times 100\%$$

where C is the average number of local lesions of the control and T is the average number of local lesions of the treatment.

**3-(7-(Hydroxymethyl)-5-methoxy-1H-isochromen-3-yl)propan-1-ol (1):** C<sub>14</sub>H<sub>18</sub>O<sub>4</sub>, Obtained as a pale yellow gum; UV (MeOH)  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 215 (3.84), 270 (3.68), 338 (3.57) nm; IR (KBr)  $\nu_{\text{max}}$  3410, 3052, 2938, 2870, 1648, 1616, 1560, 1462, 1373, 1241, 1166, 1067, 832 cm<sup>-1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR (500 and 125 MHz, in CDCl<sub>3</sub> see Table 1; ESIMS  $m/z$  (positive ion mode) 273 [M+Na]<sup>+</sup>; HRESIMS (positive ion mode)  $m/z$  273.1012 [M+Na]<sup>+</sup> (calcd C<sub>14</sub>H<sub>18</sub>NaO<sub>4</sub> for 273.1103).

**7-Isopropyl-5-methoxy-3-methyl-1H-isochromene (2):** C<sub>14</sub>H<sub>18</sub>O<sub>2</sub>, Obtained as a pale yellow gum; UV (MeOH)  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 215 (3.79), 264 (3.64), 332 (3.52) nm; IR (KBr)  $\nu_{\text{max}}$  3062, 2943, 2865, 1646, 1615, 1554, 1470, 1384, 1239, 1156, 1062, 809 cm<sup>-1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR (500 and 125 MHz, in CDCl<sub>3</sub> see Table 1; ESIMS  $m/z$  (positive ion mode) 241 [M+Na]<sup>+</sup>; HRESIMS (positive ion mode)  $m/z$  241.1210 [M+Na]<sup>+</sup> (calcd C<sub>14</sub>H<sub>18</sub>NaO<sub>2</sub> for 241.1204).

**(7-Isopropyl-5-methoxy-1H-isochromen-3-yl)methanol (3):** C<sub>14</sub>H<sub>18</sub>O<sub>3</sub>, Obtained as a pale yellow gum; UV (MeOH)  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 215 (4.01), 266 (3.73), 334 (3.67) nm; IR (KBr)  $\nu_{\text{max}}$  3392, 3057, 2938, 2862, 1614, 1550, 1463, 1376, 1245, 1159, 1058, 835 cm<sup>-1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR (500 and 125 MHz, in CDCl<sub>3</sub> see Table 1; ESIMS  $m/z$  (positive ion mode) 257 [M+Na]<sup>+</sup>; HRESIMS (positive ion mode)  $m/z$  257.1148 [M+Na]<sup>+</sup> (calcd C<sub>14</sub>H<sub>18</sub>NaO<sub>3</sub> for 257.1154).

## ACKNOWLEDGEMENTS

This work was financially supported by the National Natural Science Foundation of China (No. 81860624), the China Tobacco Monopoly Bureau Grants and Yunnan Provincial Tobacco Monopoly Bureau Grants (110202103018, 2022530000241002), and the Foundation of Yunnan Innovative Research Team (2019HC020).

## REFERENCES AND NOTES

1. R. K. Tenguria, F. N. Khan, and S. Quereshi, *World J. Sci. Technol.*, 2011, **1**, 127.
2. S. Techaoei, C. Jirayuthcharoenkul, K. J. Kom, T. Dumrongphuttidecha, and W. Khobjai, *Saudi J. Biol. Sci.*, 2020, **27**, 2883.
3. F. Bongomin, C. R. Batac, M. D. Richardson, and D. W. Denning, *Mycopathologia*, 2018, **183**, 485.
4. J. M. Restrepo-Florez, A. Bassi, and M. R. *Int. Biodeterior. Biodegradation*, 2014, **88**, 83.
5. S. Skanda and B. S. Vijayakumar, *Curr. Microbiol.*, 2021, **78**, 1317.
6. M. Y. Deng, X. Chen, Z. Y. Shi, and S. S. Xie, *Fitoterapia*, 2021, **151**, 104882.

7. W. G. Wang, L. Q. Du, S. L. Sheng, A. Li, Y. P. Li, G. G. Cheng, G. P. Li, G. L. Sun, Q. F. Hu, and Y. Matsuda, *Org. Chem. Front.*, 2019, **6**, 571.
8. M. Zhou, M. M. Miao, G. Du, X. N. Li, S. Z. Shang, W. Zhao, Z. H. Liu, G. Y. Yang, C. T. Che, Q. F. Hu, and X. M. Gao, *Org. Lett.*, 2014, **16**, 5016.
9. K. Zhou, L. Zhu, X. L. Wang, T. D. Zhang, Y. D. Wang, W. Dong, B. K. Ji, H. Y. Yang, G. Du, Q. F. Hu, and M. Zhou, *Chem. Nat. Compd.*, 2016, **52**, 591.
10. L. Yuan, W. Z. Huang, K. Zhou, Y. D. Wang, W. Dong, G. Du, X. M. Gao, Y. H. Ma, and Q. F. Hu, *Nat. Prod. Res.*, 2015, **29**, 1914.
11. M. Zhou, G. Du, H. Y. Yang, C. F. Xia, J. X. Yang, Y. Q. Ye, X. M. Gao, X. N. Li, and Q. F. Hu, *Planta Med.*, 2015, **81**, 235.
12. G. Du, Z. C. Wang, Y. K. Yang, H. M. Yang, H. Y. Yang, M. Zhou, Y. Q. Ye, X. M. Li, and Q. F. Hu, *Heterocycles*, 2015, **91**, 1996.
13. M. Zhou, K. Zhou, P. He, K. M. Wang, R. Z. Zhu, Y. D. Wang, W. Dong, G. P. Li, H. Y. Yang, Y. Q. Ye, G. Du, X. M. Li, and Q. F. Hu, *Planta Med.*, 2016, **82**, 414.
14. M. Zhou, J. Lou, Y. K. Li, Y. D. Wang, K. Zhou, B. K. Ji, W. Dong, X. M. Gao, G. Du, and Q. F. Hu, *Arch. Pharm. Res.*, 2017, **40**, 32. 1.
15. Y. Q. Ye, C. F. Xia, J. X. Yang, Y. Qin, M. Zhou, X. M. Gao, G. Du, H. Y. Yang, X. M. Li, and Q. F. Hu, *Phytochem. Lett.*, 2014, **10**, 215.
16. Q. F. Hu, H. H. Xing, Y. D. Wang, Z. H. Yu, K. L. Yan, K. Zhou, W. Dong, M. Zhou, H. Y. Yang, D. L. Zhu, and G. Du, *Chem. Nat. Compd.*, 2017, **53**, 436.
17. D. Engelmeier, F. Hadacek, O. Hofer, G. Lutz-Kutschera, M. Nagl, G. Wurz, and H. Greger, *J. Nat. Prod.*, 2004, **67**, 19.
18. A. Saeed, *Eur. J. Med. Chem.*, 2016, **116**, 290.
19. S. Pal, V. Chatare, and M. Pal, *Curr. Org. Chem.*, 2011, **15**, 782.
20. Z. Y. Lu, Y. Wang, C. D. Miao, P. P. Liu, K. Hong, and W. M. Zhu, *J. Nat. Prod.*, 2009, **72**, 1761.
21. Y. Y. Liu, Y. N. Wu, R. Zhai, Z. M. Liu, X. S. Huang, and Z. G. She, *RSC Adv.*, 2016, **6**, 72127.
22. G. Y. Yang, J. M. Dai, Q. L. Mi, Z. J. Li, X. M. Li, J. D. Zhang, J. Wang, Y. K. Li, W. G. Wang, M. Zhou, and Q. F. Hu, *Phytochemistry*, 2022, **198**, 113137.
23. S. Z. Shang, W. X. Xu, L. Li, J. G. Tang, W. Zhao, P. Lei, M. M. Miao, H. D. Sun, J. X. Pu, Y. K. Chen, and G. Y. Yang, *Phytochem. Lett.*, 2015, **11**, 53.
24. Q. F. Hu, B. Zhou, J. M. Huang, X. M. Gao, L. D. Shu, G. Y. Yang, and C. T. Che, *J. Nat. Prod.*, 2013, **76**, 292.