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FOUR NEW COMPOUNDS FROM *POUZOLZIA ZEYLANICA* (L.) BENN.

VAR. *MICROPHYLLA*

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Abstract – Two new stilbenes, pouzolignan D (**1**) and K (**2**), and two new norlignans, pouzolignan L (**3**) and M (**4**), together with four known flavonoids, rhamnocitrin (**5**), rhamnetin (**6**), isorhamnetin (**7**) and quercetin (**8**), were isolated from the aerial parts of *Pouzolzia zeylanica* (L.) Benn. var. *microphylla* (Wedd.) W. T. Wang. Their structures were elucidated by spectroscopic methods, including UV, IR, HR-ESI-TOF-MS, 1D and 2D NMR experiments.

INTRODUCTION

Pouzolzia zeylanica (L.) Benn. var. *microphylla* (Wedd.) W. T. Wang (Urticaceae), one of the four medicinal plants in the genus *Pouzolzia*, is extensively distributed in Japan, India, Malaysia, Indonesia, Australia and South China.^{1,2} It has been traditionally used for the treatment of gangrenous ulcers, sores, boils, diarrhea, syphilis, gonorrhoea, etc.³⁻⁵ A series of compounds including flavonoids, lignans, norlignans and triterpenoids have been isolated from this species and the genus *Pouzolzia*.⁶⁻¹¹ Previously, we reported the pharmacological activities of the extracts from *Pouzolzia zeylanica* var. *microphylla*, investigating its anti-inflammatory and analgesic effects,¹² therapeutic effects on mouse subcutaneous abscess¹³ and skin ulcers in rats.¹⁴ As part of our continuing study of this plant, two new stilbenes, pouzolignan D (**1**) and K (**2**), and two new norlignans, pouzolignan L (**3**) and M (**4**), along with four known flavonoids (**5-8**) (Figure 1) were obtained. Herein, we reported the isolation, structure elucidation of the four new compounds.

The ^1H NMR spectrum (Table 1) showed two 1,3,4-trisubstituted phenyl rings signals (ABX system) at δ_{H} 7.31 (1H, d, $J = 2.0$ Hz, H-2'''), 7.19 (1H, dd, $J = 8.5, 2.0$ Hz, H-6'''), 6.98 (1H, d, $J = 8.5$ Hz, H-5''') and 6.78 (1H, d, $J = 1.5$ Hz, H-2'), 6.72 (1H, dd, $J = 8.0, 1.5$ Hz, H-6'), 6.61 (1H, d, $J = 8.0$ Hz, H-5'), one 1,3,5-trisubstituted phenyl ring signal (AB₂ system) at δ_{H} 6.10 (2H, d, $J = 2.0$ Hz, H-2'', 6''), 6.01 (1H, t, $J = 2.0$ Hz, H-4'') and three methoxy groups signals at δ_{H} 3.87, 3.83, 3.68 (each 3H, s). Since three phenyl rings accounted for 12 out of 13 degrees of unsaturation, the molecule was tetracyclic. NMR and HSQC spectra showed the presence of four methines signals at δ_{H} 2.52 (1H, m, H-4) with δ_{C} 57.0 (C-4), δ_{H} 3.62 (1H, m, H-3) with δ_{C} 53.7 (C-3), δ_{H} 4.90 (1H, d, $J = 8.5$ Hz, H-5) with δ_{C} 81.8 (C-5), δ_{H} 5.20 (1H, d, $J = 7.5$ Hz, H-2) with δ_{C} 83.4 (C-2), which indicated the existence of a tetrahydrofuran ring, and another methylene signal at δ_{H} 3.80 (2H, m, H-6) with δ_{C} 61.3 (C-6) were recognized.

The NMR data suggested that compound **1** had the same fundamental skeleton as the known compound cestrumoside.¹⁵ The difference between them was **1** had three phenyl rings but cestrumoside had two, and their substituents on the phenyl rings were different. Comparison of the NMR data of **1** with those of cestrumoside showed that C-3 (δ_{C} 53.7) was shifted downfield 20.3 ppm, which suggested another phenyl ring attached on C-3 of the tetrahydrofuran ring. And it was confirmed by the HMBC correlations between H-3 and C-1'' and C-2'', 6''. The position of the hydroxy and methoxy groups in compound **1** were established by extensive analysis of the HMBC spectra (Figure 2). The relative *trans* configuration between the methine protons at C-3 and C-4, C-4 and C-5, and *cis* configuration between the methine protons at C-2 and C-3, C-2 and C-5 were established by the 2D NOESY spectral analysis. Thus, the structure of **1**, named pouzolignan D, was elucidated as 5-(3,4-dimethoxyphenyl)-2-(4-hydroxy-3-methoxyphenyl)-3-(3,5-dihydroxyphenyl)-4-(hydroxymethyl)tetrahydrofuran.

Compound **2** was obtained as a brown tabular crystal (CHCl_3 -MeOH 1:1) and had a molecular formula of $\text{C}_{30}\text{H}_{28}\text{O}_9$ established by HR-ESI-TOF-MS ($[\text{M}+\text{Na}]^+$, m/z 555.1608, $[\text{M}-\text{H}]^-$, m/z 531.1740), corresponding to 17 degrees of unsaturation. The IR spectrum showed the presence of hydroxy group (3351 cm^{-1}), aromatic ring ($1605, 1516$ and 1459 cm^{-1}). In the ^1H NMR spectrum (Table 1), two sets of aromatic rings signals at δ_{H} 6.85 (2H, d, $J = 1.5$ Hz, H-2', 2'''), 6.80 (2H, dd, $J = 8.5, 1.5$ Hz, H-6', 6'''), 6.76 (2H, d, $J = 8.5$ Hz, H-5', 5''') and 6.11 (4H, d, $J = 2.0$ Hz, H-2'', 6'', 2''', 6'''), 6.09 (2H, t, $J = 2.0$ Hz, H-4'', 4''') were recognized, revealing a 1,3,4-trisubstituted phenyl ring (ABX system) and a 1,3,5-trisubstituted phenyl ring (AB₂ system), respectively, as well as one methoxy group signal at δ_{H} 3.80 (6H, s, 3', 3'''-OMe). Moreover, the ^{13}C NMR and DEPT spectra showed ten olefinic carbon signals at

Table 1. NMR spectral data for compounds **1**, **2**
 (δ in ppm, J in Hz. 500 MHz for ^1H NMR and 125 MHz for ^{13}C NMR)

position	1 ^a		position	2 ^b	
	δ_{C}	δ_{H}		δ_{C}	δ_{H}
2	83.4	5.20 (1H, d, 7.5)	2(5)	88.6	5.22 (2H, dd, 6.5, 3.0)
3	53.7	3.62 (1H, m)	3(4)	63.3	3.44 (2H, dd, 6.5, 3.0)
4	57.0	2.52 (1H, m)			
5	81.8	4.90 (1H, d, 8.5)			
6	61.3	3.80 (2H, m)			
1'	131.4		1' (1''')	133.9	
2'	110.7	6.78 (1H, d, 1.5)	2' (2''')	110.4	6.85 (2H, d, 1.5)
3'	146.5		3' (3''')	146.6	
4'	145.0		4' (4''')	148.3	
5'	113.8	6.61 (1H, d, 8.0)	5' (5''')	115.5	6.76 (2H, d, 8.5)
6'	119.3	6.72 (1H, dd, 8.0, 1.5)	6' (6''')	119.4	6.80 (2H, dd, 8.5, 1.5)
1''	144.3		1'' (1''')	141.1	
2'' (6'')	108.1	6.10 (2H, d, 2.0)	2'' (6'', 2''', 6''')	107.3	6.11 (4H, d, 2.0)
3'' (5'')	157.7		3'' (5'', 3''', 5''')	158.9	
4''	100.2	6.01 (1H, t, 2.0)	4'' (4''')	101.8	6.09 (2H, t, 2.0)
1'''	134.5				
2'''	110.6	7.31 (1H, d, 2.0)			
3'''	149.3				
4'''	148.7				
5'''	111.7	6.98 (1H, d, 8.5)			
6'''	118.7	7.19 (1H, dd, 8.5, 2.0)			
3'-OMe	55.1	3.68 (3H, s)	3' (3''')-OMe	55.8	3.80 (6H, s)
3'''-OMe	55.4	3.87 (3H, s)			
4'''-OMe	55.2	3.83 (3H, s)			

^a in CD_3COCD_3 , ^b in CD_3OD .

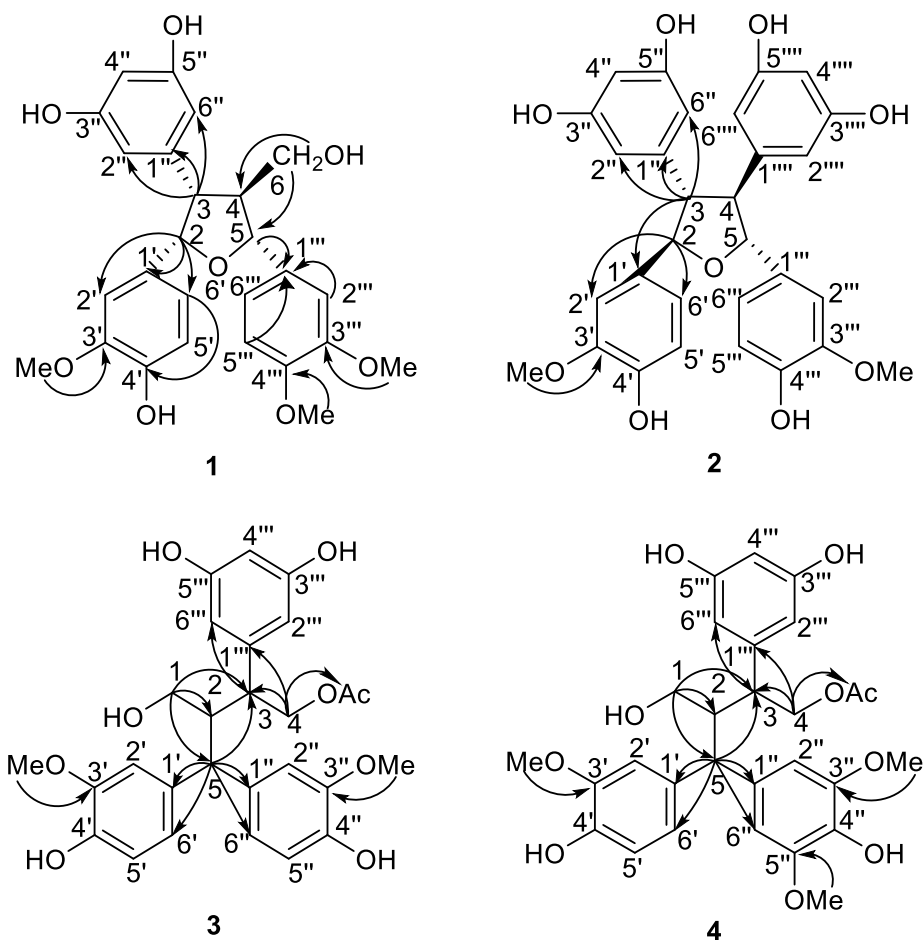


Figure 2. Selected HMBC correlations (H→C) of compounds 1-4

δ_C 158.9 (C-3'', 5'', 3''', 5'''), 148.3 (C-4', 4'''), 146.6 (C-3', 3'''), 141.1 (C-1'', 1'''), 133.9 (C-1', 1'''), 119.4 (C-6', 6'''), 115.5 (C-5', 5'''), 110.4 (C-2', 2'''), 107.3 (C-2'', 6'', 2''', 6'''), 101.8 (C-4'', 4'''), two tertiary methyl carbon signals at δ_C 88.6 (C-2, 5), 63.3 (C-3, 4) and one methoxy group signal at δ_C 55.8 (3', 3'''-OMe).

Furthermore, comparing the molecular formula $C_{30}H_{28}O_9$ and 17 degrees of unsaturation of compound **2** with its fewer 1H NMR and ^{13}C NMR signals, it indicated that this molecule had symmetrical elements and overlapping carbon signals with four phenyl rings and the integration of hydrogen in the 1H NMR spectrum should be doubled. Thus, compound **2** had four phenyl rings which accounted for 16 units of the 17 degrees of unsaturation and the molecule was pentacyclic. Moreover, the 1H NMR, ^{13}C NMR and HSQC spectra revealed four methines signals at δ_H 3.44 (2H, dd, $J = 6.5, 3.0$ Hz, H-3, 4) with δ_C 63.3 (C-3, 4) and δ_H 5.22 (2H, dd, $J = 6.5, 3.0$ Hz, H-2, 5) with δ_C 88.6 (C-2, 5), which suggested the presence of a tetrahydrofuran ring with symmetry substituents. From the above-mentioned analysis and HMBC correlations (Figure 2), it suggested that the four phenyl groups were linked to the four carbons of

tetrahydrofuran ring, respectively. It showed **2** was similar to compound **1** and had a symmetrical structure. Confirming evidence was obtained from the HMBC correlations of H-2, 5 with C-3, 4, C-2', 2''', C-6', 6''' and H-3, 4 with C-1', 1''', C-1'', 1''', C-2'', 6'', 2''', 6'''. The location of the methoxy group at C-3', 3''' were proved by HMBC correlation between methoxy protons (δ_{H} 3.80) and C-3', 3''' (δ_{C} 146.6) and NOESY interactions of methoxy protons with H-2', 2'''. The relative *trans* configuration between the methine protons at C-2, 5 and C-3, 4 were established by the weak interactions between H-2, 5 and H-3, 4 in the 1D and 2D NOESY spectra. Moreover, its basic skeleton is similar to the known (2*S*,3*R*,4*R*,5*S*)-2,3,4,5-tetrakis(4-methoxyphenyl)tetrahydrofuran¹⁶ and tricuspidatol-A.¹⁷ The ¹H NMR data of former compound showed the tetrahydrofuran signals at δ_{H} 5.26 (2H, dd, $J = 6.3, 2.7$ Hz) and 3.52 (2H, dd, $J = 6.3, 2.7$ Hz), while the data of tricuspidatol-A at δ_{H} 5.25 (2H, dd, $J = 5.0, 1.5$ Hz) and 3.50 (2H, dd, $J = 5.0, 1.5$ Hz), which both resemble the data of compound **2**. Thus, on the basis of 2D NOESY of **2**, we confirmed the relative stereochemistry structures of **2** in Figure 1. Consequently, the structure of **2**, named pouzolignan K, was characterized as 2,5-bis(4-hydroxy-3-methoxyphenyl)-3,4-bis(3,5-dihydroxyphenyl)tetrahydrofuran.

Compound **3** was obtained as white powder, the molecular formula of C₂₇H₃₀O₉ was determined from the HR-ESI-TOF-MS ($[\text{M}+\text{Na}]^+$, m/z 521.1786), suggesting 13 degrees of unsaturation. The UV spectrum showed absorption maxima at 275 and 281 nm in methanol, and the IR spectrum showed absorption bands at 3362, 1720, 1601, 1512 and 1462 cm⁻¹, indicating the presence of hydroxy group, carbonyl group, and aromatic ring. The ¹H NMR and ¹³C NMR spectrum of **3** (Table 2) along with analysis of the DEPT spectra displayed 27 carbon signals and 30 proton signals. In the ¹H-NMR spectrum of **3** (Table 2), two 1, 3, 4-trisubstituted protons (ABX system) at δ_{H} 6.94 (1H, d, $J = 2.0$ Hz, H-2'), 6.90 (1H, dd, $J = 2.0, 8.0$ Hz, H-6'), 6.82 (1H, d, $J = 8.0$ Hz, H-5'), and 6.76 (1H, d, $J = 2.0$ Hz, H-2''), 6.65 (1H, dd, $J = 8.0, 2.0$ Hz, H-6''), 6.63 (1H, d, $J = 8.0$ Hz, H-5''), one 1, 3, 5-trisubstituted phenyl protons (AB₂ system) at δ_{H} 6.17 (2H, d, $J = 2.0$ Hz, H-2''', 6'''), 6.19 (1H, t, $J = 2.0$ Hz, H-4'''), three methine protons at δ_{H} 2.84-2.88 (1H, m, H-2), 3.13-3.17 (1H, m, H-3), 3.37 (1H, d, $J = 12.0$ Hz, H-5), two oxymethylene protons at δ_{H} 3.31-3.37 (2H, m, H-1), 4.51-4.59 (2H, m, H-4), two methoxyl protons at δ_{H} 3.90, 3.78 (each 3H, s), as well as an acetyl methyl protons at δ_{H} 1.94 (3H, s) were recognized. DEPT experiment revealed that **3** had ten quaternary carbons, twelve tertiary carbons, two methylenes and three methyl groups (Table 2). In the HMBC spectrum, correlations of H-5 (δ_{H} 3.37) with C-1' (δ_{C} 137.9), C-6' (δ_{C} 121.9), C-1'' (δ_{C} 137.3), and C-6'' (δ_{C} 121.1), proved that the two 1, 3, 4-trisubstituted phenyl rings were both linked to C-5.

Table 2. NMR spectral data for compounds **3**, **4** and pouzolignan F
(in CD₃OD, δ in ppm, J in Hz. 500 MHz for ¹H NMR and 125 MHz for ¹³C NMR)

position	3		4		pouzolignan F ^a	
	δ_C	δ_H	δ_C	δ_H	δ_C	δ_H
1	63.2	3.31-3.37 (2H, m)	63.2	3.28-3.35 (2H, m)	65.6	3.72 (1H, dd, 11.5, 4.5) 4.05 (1H, dd, 11.5, 4.0)
2	46.7	2.84-2.88 (1H, m)	46.7	2.82-2.89 (1H, m)	42.3	3.12-3.14 (1H, m)
3	46.5	3.13-3.17 (1H, m)	46.5	3.16-3.19 (1H, m)	48.9	3.01-3.04 (1H, m)
4	69.7	4.51-4.59 (2H, m)	69.7	4.53-4.72 (2H, m)	65.6	3.96-3.98 (1H, m) 3.84 (1H, d, 7.5)
5	53.2	3.37 (1H, d, 12.0)	53.7	3.36 (1H, d, 12.0)	53.5	3.65 (1H, d, 12.0)
1'	137.9		137.1		137.1	
2'	113.4	6.94 (1H, d, 2.0)	113.5	6.96 (1H, d, 2.0)	113.2	7.05 (1H, d, 2.0)
3'	149.3		149.3		149.3	
4'	146.0		146.1		146.0	
5'	116.6	6.82 (1H, d, 8.0)	116.6	6.83 (1H, d, 8.0)	116.2	6.82 (1H, d, 8.0)
6'	121.9	6.90 (1H, dd, 8.0, 2.0)	121.9	6.92 (1H, dd, 8.0, 2.0)	122.1	6.95 (1H, dd, 8.0, 2.0)
1''	137.3		137.2		136.7	
2''	112.4	6.76 (1H, d, 2.0)	105.9	6.48 (1H, s)	112.7	6.84 (1H, d, 2.0)
3''	149.0		149.4		148.9	
4''	145.8		134.9		145.8	
5''	116.3	6.63 (1H, d, 8.0)	149.4		116.2	6.65 (1H, d, 8.0)
6''	121.1	6.65 (1H, dd, 8.0, 2.0)	105.9	6.48 (1H, s)	121.5	6.70 (1H, dd, 8.0, 2.0)
1'''	142.4		142.4		142.9	
2''', 6'''	109.5	6.17 (2H, d, 2.0)	109.5	6.18 (2H, d, 2.0)	109.1	6.10 (2H, d, 2.0)
3''', 5'''	159.3		159.3		159.3	
4'''	102.3	6.19 (1H, t, 2.0)	102.3	6.19 (1H, t, 2.0)	102.2	6.19 (1H, t, 2.0)

3'-OMe	56.7	3.90 (3H, s)	56.7	3.90 (3H, s)	56.6	3.92 (3H, s)
3''-OMe	56.5	3.78 (3H, s)	56.9	3.79 (3H, s)	56.5	3.80 (3H, s)
5''-OMe			56.9	3.79 (3H, s)		
Ac	21.1	1.94 (3H, s)	21.1	1.94 (3H, s)	20.6	1.87 (3H, s)
	173.2		173.2		172.8	

^a data from 11.

Furthermore, H-3 (δ_{H} 3.13-3.17) with C-2''', 6''' (δ_{C} 109.5) indicated that the 1, 3, 5-trisubstituted phenyl ring was attached to C-3 as well (Figure 2).

The NMR data suggested that compound **3** had the uniform norlignan skeleton as pouzolignan F¹¹ which was reported in our earlier phytochemical investigations. Comparing the NMR data of **3** with those of pouzolignan F, C-4 (δ_{C} 69.7) was shifted downfield 4.1 ppm and C-3 (δ_{C} 46.5) was shifted upfield 2.4 ppm, as well as correlations between H-4 with C-1''', C-3, CH₃CO in HMBC, thus, the acetoxy group was attached to C-4. The position of the hydroxy and methoxy groups in compound **3** were established by extensive analysis of the HMBC spectra (Figure 2). Based on the above results, the structure of **3**, named pouzolignan L, was established as 1-hydroxy-3-(3,5-dihydroxyphenyl)-2-[bis(4-hydroxy-3-methoxyphenyl)methyl]butyl acetate.

Compound **4** was obtained as white powder, the molecular formula of C₂₈H₃₂O₁₀ was assigned from the HR-ESI-TOF-MS ([M+Na]⁺, *m/z* 551.1889), suggesting 13 degrees of unsaturation. The ¹H and ¹³C NMR spectral data of **4** (Table 2) were similar to those of **3**, except for the presence of an additional methoxy group at δ_{H} 3.79 (3H, s) and δ_{C} 56.9, and the 1, 3, 4, 5-tetrasubstituted phenyl signals at δ_{H} 6.48 (2H, s, H-2'', 6'') with δ_{C} 105.9 (C-2'', 6''). The location of the methoxy group at C-5'' was confirmed by the HMBC correlation (Figure 2). Hence, the structure of **4**, named pouzolignan M, was identified as 1-hydroxy-3-(3,5-dihydroxyphenyl)-2-[4-hydroxy-3-methoxyphenyl-(4-hydroxy-3,5-dimethoxyphenyl)]butyl acetate.

The known compounds were identified as rhamnocitrin (**5**), rhamnetin (**6**),¹⁸ isorhamnetin (**7**)¹⁹ and quercetin (**8**),²⁰ by comparing the spectroscopic data with those reported in the literature values.

EXPERIMENTAL

General UV spectra were obtained using a Shimadzu UV-2450 spectrophotometer. IR spectra were recorded on a Nicolet 6700 FT-IR spectrometer. NMR spectra were run on a Bruker AVANCE III 500

spectrometer with TMS as internal standard, and the chemical shifts (δ) were expressed in ppm. HR-ESI-TOF-MS was performed on an API QSTAR time-of-flight spectrometer. TLC was performed on precoated silica gel GF254 plates from Yantai Jiangyou Company. Silica gel 100-200, 200-300 and 300-400 mesh from Qingdao Haiyang Chemical Company, YWG-C₁₈ (50-70 μ m) from Tianjin Boruijianhe Chromatography Technology Company and Sephadex LH-20 from Amersham Biosciences were used for column chromatography. Preparative RP-C₁₈ HPLC was performed by a Shimadzu LC-6AD series instrument with Shim-Park RP-C₁₈ column (20 \times 200 mm i.d.).

Plant material The aerial parts of *P. zeylanica* var. *microphylla* were collected in Guangzhou, Guangdong province of China on January, 2012 and identified by Professor Ji-Zhu Liu, Guangdong Pharmaceutical University. A voucher specimen (No. 20120113) has been deposited in the Lab of Traditional Chinese Medicine Chemistry, Guangdong Pharmaceutical University.

Extraction and Isolation The air-dried and powdered aerial parts of *P. zeylanica* var. *microphylla* (5 kg) were extracted with petroleum ether (150 L) under reflux for two times (1 h per time). After the solvent on the plants was volatilized completely, the plants were extracted with EtOAc (150 L) under reflux for three times (1.5 h per time), which was evaporated under reduced pressure to yield a residue (80 g). The EtOAc extract was subjected to silica gel (100-200 mesh) column chromatography, eluted with petroleum ether-EtOAc (50:1 \rightarrow 1:1), EtOAc, CH₂Cl₂-MeOH (1:1), MeOH to yield six fractions, Fr.A-F. Fr.D (13.4 g) was resubjected by silica gel (200-300 mesh) column chromatography, eluted with CHCl₃-MeOH (50:1 \rightarrow 5:1) to yield Fr.D1-D5. Fr.D1 was separated by Sephadex LH-20, eluted with CHCl₃-MeOH (1:1) and purified by RP-C₁₈ column (50-70 μ m), using 70% MeOH as the eluent to afford **2** (7.5 mg). Fr.D2 was subjected to sephadex LH-20, eluted with CHCl₃-MeOH (1:1) to yield four fractions, and the second fraction (0.3 g) was purified by RP-C₁₈ column (50-70 μ m), eluted with 70% MeOH to afford **1** (17 mg). Fr.E (4 g) was resubjected to silica gel (200-300 mesh) column chromatography, eluted with CH₂Cl₂-MeOH (25:1 \rightarrow 8:1) to yield Fr.E1-E4. Fr.E2 was separated repeatedly by silica gel (300-400 mesh) column chromatography, eluted with CH₂Cl₂-MeOH (17:1) and finally by preparative RP-C₁₈ HPLC, eluted with MeOH-H₂O-HCO₂H (23:1:0.5) to afford **3** (14 mg). Fr.E3 was further fractionated repeatedly by silica gel (300-400 mesh) column chromatography, eluted with CH₂Cl₂-MeOH (14:1), then was separated by Sephadex LH-20, eluted with CHCl₃-MeOH (3:2) to afford **4** (14 mg). Fr.F (14 g) was

submitted to silica gel (200-300 mesh) column chromatography, eluted with CHCl₃-MeOH (20:1→1:1) to afford Fr.F1-F6. Fr.F2 (0.5 g), Fr.F4 (1.0 g) and Fr.F5 (0.3 g) were subjected to Sephadex LH-20, eluted with MeOH to yield compounds **5** (15 mg), **6** (15 mg), **7** (18 mg) and **8** (25 mg), respectively.

Pouzolignan D (1) : Brown amorphous powder; $[\alpha]_D^{20} +34.0$ (*c* 0.1, MeOH); IR (KBr) ν_{\max} 3360, 2940, 2840, 1608, 1520, 1455, 1347, 1270, 1150, 820 cm⁻¹; UV (MeOH) λ_{\max} nm (log ϵ) 274 (4.02); HR-ESI-TOF-MS *m/z* 467.1729 [M-H]⁻ (calcd 467.1733 for C₂₆H₂₇O₈⁻), ¹H and ¹³C NMR data (see Table 1).

Pouzolignan K (2): Brown tabular crystals (CHCl₃-MeOH 1:1); $[\alpha]_D^{20} +71.4$ (*c* 0.1, MeOH); IR (KBr) ν_{\max} 3351, 1605, 1516, 1459, 1435, 1273, 1238, 1156, 1124, 1031, 1001 and 921 cm⁻¹; UV (MeOH) λ_{\max} nm (log ϵ) 280 (4.13); HR-ESI-TOF-MS *m/z* 555.1608 [M+Na]⁺, 531.1740 [M-H]⁻ (calcd 555.1611 for C₃₀H₂₈NaO₉⁺, 531.1743 for C₃₀H₂₇O₉⁻), ¹H and ¹³C NMR data (see Table 1).

Pouzolignan L (3): white powder; $[\alpha]_D^{20} +29.7$ (*c* 0.1, MeOH); UV (MeOH) λ_{\max} nm (log ϵ) 275 (3.25); IR (KBr) ν_{\max} 3362, 2942, 1720, 1601, 1512, 1462, 1365, 1128 and 849 cm⁻¹, HR-ESI-TOF-MS *m/z* 521.1786 [M+Na]⁺ (calcd 521.1788 for C₂₇H₃₀NaO₉⁺), ¹H and ¹³C NMR data (see Table 2).

Pouzolignan M (4): white powder; $[\alpha]_D^{20} +36.1$ (*c* 0.1, MeOH), UV (MeOH) λ_{\max} nm (log ϵ) 275(3.34); IR (KBr) ν_{\max} 3362, 2941, 1721, 1602, 1514, 1462, 1366, 1126 and 849 cm⁻¹; HR-ESI-TOF-MS *m/z* 551.1889 [M+Na]⁺ (calcd 551.1893 for C₂₈H₃₂NaO₁₀⁺) ¹H and ¹³C NMR data (see Table 2).

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