

HETEROCYCLES, Vol. 92, No. 8, 2016, pp.1462 - 1467. © 2016 The Japan Institute of Heterocyclic Chemistry  
Received, 12th April, 2016, Accepted, 31st May, 2016, Published online, 20th June, 2016  
DOI: 10.3987/COM-16-13482

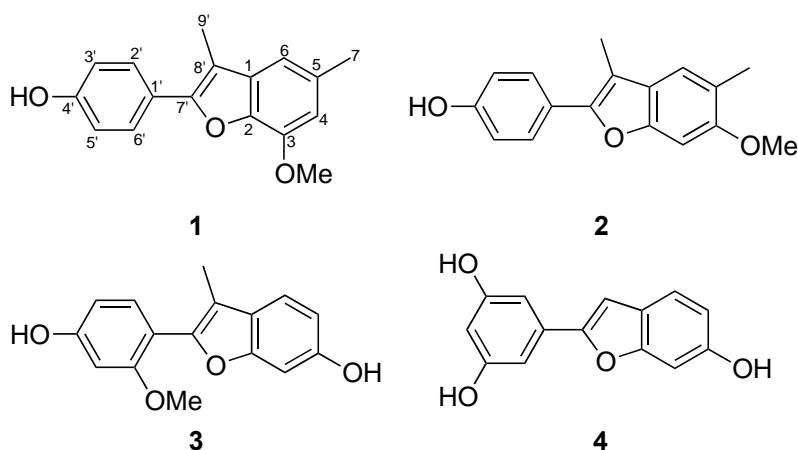
## TWO NEW 2-ARYLBENZOFURANS FROM THE STEMS OF *NICOTIANA TABACUM* AND THEIR BIOACTIVITIES

Pei-Song Yang,<sup>1,2</sup> Wei Zhang,<sup>2</sup> Xiao-Feng Shen,<sup>2</sup> Xin-Lin Wang,<sup>2</sup> Chao Li,<sup>2</sup>  
Xiao-Wei Gong,<sup>2</sup> Xu-Dong Zheng,<sup>2</sup> Dong-Lai Zhu,<sup>2\*</sup> and Jia-Qiang Wang<sup>2,\*</sup>

<sup>1</sup> School of Chemical Science and Technology, Yunnan University, Kunming 650091, P.R. China; <sup>2</sup> Key Laboratory of Tobacco Chemistry of Yunnan Province, China Tobacco Yunnan Industrial Co., Ltd, Kunming 650231, P.R. China; E-mail: 16594939@qq.com; jszxtg\_2015@163.com

**Abstract** – Two new 2-arylbenzofurans, 4-(7-methoxy-3,5-dimethyl-benzofuran-2-yl)phenol (**1**), 4-(6-methoxy-3,5-dimethylbenzofuran-2-yl)phenol (**2**), together with two known 2-arylbenzofurans (**3** and **4**) were isolated from the stems of *Nicotiana tabacum*. Their structures were determined by means of HRESIMS and extensive 1D and 2D NMR spectroscopic studies. Compounds **1-4** were tested for their anti-tobacco mosaic virus (TMV) activities and cytotoxicity activities. The results showed that compound **2** exhibited high anti-TMV activities with inhibition rates of 36.4% at the concentration of 20  $\mu\text{M}$ . This rate is higher than that of positive control. Compounds **1-4** showed moderate-to-weak cytotoxicities against some tested human tumor cell lines with  $\text{IC}_{50}$  values in the range of 2.2–8.4  $\mu\text{M}$ .

*Nicotiana tabacum*, tobacco is an important economic crop.<sup>1,2</sup> Its leaves are used as a raw material for the tobacco industry, aerial plant as an insecticide, and also as anesthetic, diaphoretic, sedative, and emetic agents in Chinese folklore medicine.<sup>1-3</sup> In previous literatures, many new bioactive compounds, such as, sesquiterpenes,<sup>4-6</sup> alkaloids,<sup>7,8</sup> lignans,<sup>9,10</sup> flavonoids,<sup>11-14</sup> phenylpropanoids,<sup>15,16</sup> chromanones,<sup>17,18</sup> biphenyls,<sup>19</sup> phenolic amides,<sup>20</sup> isocoumarins,<sup>21</sup> were isolated from *N. tabacum*. The stems of *N. tabacum* are the main by-product in tobacco planting and are normally used as organic fertilizer. The multipurpose utilization of the roots and stems of *N. tabacum* is an interesting topic and has attracted more and more attentions.<sup>16,21</sup> In continuing efforts to utilize *N. tabacum* and identify bioactive natural products, the phytochemistry investigation of the stems of Yunyan 201 (a variety of *N. tabacum*) led to the isolation of two new (**1** and **2**) and two known 2-arylbenzofurans (**3** and **4**). This paper deals with the isolation, structural elucidation, and their bioactivities of these compounds.



**Figure 1.** The structures of 2-arylbenzofurans from stems of *Nicotiana tabacum*

A 95% aq. MeOH extract prepared from the stems of tobacco was subjected repeatedly to column chromatography on silica gel and preparative HPLC to afford two new 2-arylbenzofurans, 4-(7-methoxy-3,5-dimethylbenzofuran-2-yl)phenol (**1**) and 4-(6-methoxy-3,5-dimethylbenzofuran-2-yl)phenol (**2**), together with two known 2-arylbenzofurans (**3** and **4**). The structures of the compounds **1-4** were as shown in **Figure 1**, and the  $^1\text{H}$  and  $^{13}\text{C}$  NMR data of **1** and **2** were listed in Table 1. The known compounds, compared with literature, were identified as 2-[(2'-methoxy-4'-hydroxy)phenyl]-3-methyl-6-hydroxybenzofuran (**3**)<sup>22</sup> and moracin M (**4**).<sup>23</sup>

Compound **1** was obtained as a yellow gum.

Its molecular formula was determined as  $\text{C}_{17}\text{H}_{16}\text{O}_3$  by HRESIMS ( $m/z$  291.0990  $[\text{M}+\text{Na}]^+$ ; calcd  $\text{C}_{17}\text{H}_{16}\text{NaO}_3$  for 291.0997). Its UV spectrum showed maximum absorption at 210, 270 and 309 nm. The IR absorptions at 3415, 1620, 1487, 1436  $\text{cm}^{-1}$  indicated the presence of hydroxy and aromatic rings. The  $^1\text{H}$ ,  $^{13}\text{C}$  NMR and DEPT spectra of **1** (**Table 1**) displayed 17 carbon and 16 proton signals, respectively, corresponding to a 1,2,3,5-tetrasubstituted phenyl ring (C-1~C-6, H-4 and H-6), a 1,4-substituted phenyl ring (C-1'~C-6',

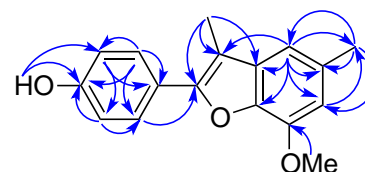
**Table 1.**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data of compounds **1** and **2** (in  $\text{C}_5\text{D}_5\text{N}$ )

No.	<b>1</b>		<b>2</b>	
	$\delta_{\text{C}}$	$\delta_{\text{H}}$ (m, <i>J</i> , Hz)	$\delta_{\text{C}}$	
1	126.7 s		118.1 s	
2	144.7 s		148.6 s	
3	151.4 s		101.3 d	7.02 s
4	112.3 d	7.37 (d) 1.8	155.6 s	
5	134.5 s		120.2 s	
6	116.8 d	7.73 (d) 1.8	121.5 d	7.33 s
7	24.9 q	2.31 s	18.6 q	2.40 s
1'	122.5 s		122.7 s	
2',6'	129.8 d	8.00 (d) 8.6	130.0 d	7.98 (d) 8.6
3',5'	116.3 d	6.78 (d) 8.6	116.5 d	6.79 (d) 8.6
4'	157.9 s		157.6 s	
7'	152.7 s		152.1 s	
8'	111.2 s		111.1 s	
9'	9.1 q	2.14 s	9.3 q	2.17 s
3-OMe	56.1 q	3.86 s		
4-OMe			56.4 q	3.84 s
Ar-OH		10.81 s		10.90 s

H<sub>2</sub>-2',6', and H<sub>2</sub>-3',5'), two methyl group (C-7 and C-9'; H<sub>3</sub>-7 and H<sub>3</sub>-9'), two olefin carbons (C-7' and C-8'), one methoxy group ( $\delta_{\text{C}}$  56.1 q;  $\delta_{\text{H}}$  3.86 s), and one phenolic hydroxy group ( $\delta_{\text{H}}$  10.81). Its  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectroscopic data [two phenyl rings, two olefin carbons (C-7' and C-8'), and methyl group (C-9' and H<sub>3</sub>-9')] were similar to those of known compound (**3**), which suggested that **1** should be a

3-methyl-2-arylbenzofuran.<sup>22</sup> In addition, the 3-methyl-2-arylbenzofuran skeleton of **1** was also assigned by the HMBC correlations (**Figure 2**) of H<sub>3</sub>-9' with C-1, C-7', and C-8'; of H<sub>2</sub>-2',6' with C-7'; and of H-6 with C-8'. The position of the methyl group (C-7) at C-5 of the arylbenzofuran was elucidated by the HMBC correlations from H<sub>3</sub>-7 to C-4, C-5, and C-6; from H-4 to C-7; and from H-6 to C-7. The HMBC correlations from the methoxy proton ( $\delta_{\text{H}}$  3.86) to C-3 ( $\delta_{\text{C}}$  151.4) supported the methoxy group located at C-3. The phenolic hydroxy group located at C-4' was supported by the HMBC correlation of phenolic hydroxy proton ( $\delta_{\text{H}}$  10.81) with C-4' and C-3',5'. Thus, the structure of **1** was established as 4-(7-methoxy-3,5-dimethylbenzofuran-2-yl)phenol.

Compound **2** was also obtained as yellow gum and it gave an [M+Na]<sup>+</sup> peak at *m/z* 291.0998 in HRESIMS, consistent with a molecular formula of C<sub>17</sub>H<sub>16</sub>O<sub>3</sub>. The <sup>1</sup>H and <sup>13</sup>C spectra data of **2** was very similar to these of **1** (see **Table 1**). The



major differences were due to the substituent position variations on the tetrasubstituted phenyl ring (C-1~C-6). The methyl group (C-7) located at C-5 of was supported by the HMBC correlations from H<sub>3</sub>-7 to C-4, C-5, and C-6; from H-4 to C-7; and from H-6 to C-7. The HMBC correlations from the methoxy proton ( $\delta_{\text{H}}$  3.84) to C-4 ( $\delta_{\text{C}}$  155.6) supported the methoxy group located at C-4. The phenolic hydroxy group located at C-4' was supported by the HMBC correlation of phenolic hydroxy proton ( $\delta_{\text{H}}$  10.90) with C-4' and C-3',5'. Thus, the structure of

4-(6-methoxy-3,5-dimethylbenzofuran-2-yl)phenol (**2**) was established.

Compounds **1-4** were tested for their anti-tobacco mosaic

**Table 2.** TMV Infection inhibition activities of compounds **1-4**

Compounds	Inhibition rate (%)	IC <sub>50</sub> ( $\mu\text{M}$ )	Compounds	Inhibition rates (%)	IC <sub>50</sub> ( $\mu\text{M}$ )
<b>1</b>	25.3 $\pm$ 3.5	62.5	<b>4</b>	21.2 $\pm$ 3.0	78.3
<b>2</b>	36.4 $\pm$ 3.2	31.6	ningnanmycin	31.5 $\pm$ 3.1	36.3
<b>3</b>	20.6 $\pm$ 2.8	71.6			

All results are expressed as mean  $\pm$  SD; n = 3 for all groups.

virus (TMV) activities. The anti-TMV activities were tested by half-leaf method, using ningnanmycin (a commercial product for plant disease in China, with inhibition rate of 32.2%) as a positive control.<sup>24,25</sup> The results showed that compounds **2** exhibited high anti-TMV activities with inhibition rates of 36.4% at the concentration of 20  $\mu\text{M}$ . This rate is higher than that of positive control. The other compounds also showed potential activities with inhibition rates in the range of 20.6%~25.3 % at the concentration of 20  $\mu\text{M}$ , respectively.

The cytotoxicities of compounds **1-4** were also tested using a previously reported procedure.<sup>26,27</sup> The cytotoxic abilities against five human tumor cell lines (NB4, A549, SHSY5Y, PC3, and MCF7) by MTT-assay were summarized in **Table 3**. The results revealed that compounds **1-4** showed moderate-to-weak inhibitory activities against some tested human tumor cell lines with IC<sub>50</sub> values in the range of 2.2–8.4  $\mu\text{M}$ .

## EXPERIMENTAL

**General.** UV spectra were obtained using a Shimadzu UV-2401A spectrophotometer. A Tenor 27 spectrophotometer was used for scanning IR spectroscopy with KBr pellets. 1D and 2D NMR spectra were recorded on DRX-500 spectrometers with TMS as internal standard, and the chemical shifts ( $\delta$ ) were expressed in ppm. HRESIMS was performed on an API QSTAR time-of-flight spectrometer and a VG Autospec-3000 spectrometer, respectively. Preparative HPLC was performed on a Shimadzu LC-8A preparative liquid chromatograph with a ZORBAX PrepHT GF (21.2 mm  $\times$  25 cm, 7  $\mu$ m) column or a Venusil MP C<sub>18</sub> (20 mm  $\times$  25 cm, 5  $\mu$ m) column. Column chromatography was performed with Si gel (200–300 mesh, Qing-dao Marine Chemical, Inc., Qingdao, China). The fractions were monitored by TLC, and spots were visualized by heating Si gel plates sprayed with 5% H<sub>2</sub>SO<sub>4</sub> in EtOH.

**Table 3.** Cytotoxic activity of compounds 1–4

Compounds	Cell lines and IC <sub>50</sub> ( $\mu$ M)				
	NB4	A549	SHSY5Y	PC3	MCF7
<b>1</b>	3.8	5.9	>10	6.4	>10
<b>2</b>	2.8	4.5	5.2	2.2	3.4
<b>3</b>	7.3	>10	>10	6.8	8.4
<b>4</b>	>10	5.5	>10	>10	6.0
<b>Taxol</b>	0.03	0.02	0.05	0.05	0.05

NB4, human leukemia cell; A549, carcinomic human alveolar basal epithelial cell; SHSY5Y, human neuroblastoma cell; PC3, Human prostate cancer cell; MCF7, Human breast adenocarcinoma cell.

**Plant material.** The stems of *N. tabacum* L (tobacco stems) was collected from Lijiang County, Yunnan Province, P. R. China, in September 2014. The tobacco variety is Yunyan-201, which had widely cultivated in China. The identification of the plant material was verified by Prof. H. W. Yang (School of Tobacco, Yunnan Agriculture University).

**Extraction and Isolation.** The air-dried and powdered tobacco stems (3.8 kg) were extracted with 95% MeOH, and the extract was partitioned between EtOAc. The EtOAc-soluble materials (63.8 g) were applied to silica gel (200–300 mesh) column chromatography, eluting with CHCl<sub>3</sub>/MeOH gradient system (9:1, 8:2, 7:3, 6:4, 5:5, 4:6) to give six fractions A–F. Further separation of fraction A (9:1, 13.6 g) by silica gel column chromatography, eluted with CHCl<sub>3</sub>/Me<sub>2</sub>CO (8:2-2:1) yielded mixtures A1–A7. Fraction A2 (7:3, 0.67 g) was subjected to silica gel column chromatography using petroleum ether/acetone, and then semi-preparative HPLC (48% MeOH/H<sub>2</sub>O, flow rate 12 mL/min) to give **1** (12.5 mg) and **2** (10.8 mg). Fraction A3 (6:4, 1.64 g) was subjected to silica gel column chromatography using petroleum ether/acetone, and then semi-preparative HPLC (44% MeOH/H<sub>2</sub>O, flow rate 12 mL/min) to give **3** (14.2 mg) and **4** (13.1 mg).

**Anti-TMV Assays.** The anti-TMV activity was tested using the half-leaf method,<sup>24,25</sup> and Ningnanmycin (2% water solution), a commercial product for plant disease in China, was used as positive control.

**Cytotoxicity Assay.** The cytotoxicity tests for the isolates were performed by against NB4, A549, SHSY5Y, PC3, and MCF7 tumor cell lines by MTT-assay (with Taxol as the positive control).<sup>26,27</sup>

**4-(7-Methoxy-3,5-dimethylbenzofuran-2-yl)phenol (1)**: C<sub>17</sub>H<sub>16</sub>O<sub>3</sub>, obtained as a yellow gum; UV (MeOH)  $\lambda_{\max}$  (log  $\epsilon$ ) 210 (4.10), 270 (3.15), 309 (3.52) nm; IR (KBr)  $\nu_{\max}$  3415, 1620, 1487, 1436, 1379, 1158, 1069, 865, 793 cm<sup>-1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR (500 and 125 MHz, in C<sub>5</sub>D<sub>5</sub>N, see Table-1; ESIMS  $m/z$  (positive ion mode) 291 [M+Na]<sup>+</sup>; HRESIMS (positive ion mode)  $m/z$  291.0990 [M+Na]<sup>+</sup> (calcd C<sub>17</sub>H<sub>16</sub>NaO<sub>3</sub> for 291.0997).

**4-(6-Methoxy-3,5-dimethylbenzofuran-2-yl)phenol (2)**: C<sub>17</sub>H<sub>16</sub>O<sub>3</sub>, obtained as a yellow gum; UV (MeOH)  $\lambda_{\max}$  (log  $\epsilon$ ) 210 (4.06), 268 (3.26), 306 (3.57) nm; IR (KBr)  $\nu_{\max}$  3420, 1618, 1482, 1440, 1382, 1153, 1062, 880, 764 cm<sup>-1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR (500 and 125 MHz, in C<sub>5</sub>D<sub>5</sub>N, see Table-1; ESIMS  $m/z$  (positive ion mode) 291 [M+Na]<sup>+</sup>; HRESIMS (positive ion mode)  $m/z$  291.0998 [M+Na]<sup>+</sup> (calcd C<sub>17</sub>H<sub>16</sub>NaO<sub>3</sub> for 291.0997).

## ACKNOWLEDGEMENTS

This research was supported by the National Natural Science Foundation of China (No. 21562049, No. 31360081 and No. 31400303), and the Applied Fundamental Foundation of Yunnan Province (No. 2014FB163, No. 2015FB162).

## REFERENCES

1. The Editorial Committee of the Administration Bureau of Flora of China, 'Flora of China,' Vol. 67, Beijing Science and Technology Press, Beijing, 2005.
2. A. Rodgman and T. A. Perfetti, 'The Chemical Components of Tobacco and Tobacco Smoke,' CRC Press, Taylor and Francis Group, Boca Raton, Florida, 2008.
3. A. Fuchs, W. Slobbe, P. C. Mol, and M. A. Posthumus, *Phytochemistry*, 1983, **22**, 1197.
4. S. Z. Shang, W. Zhao, J. G. Tang, X. M. Xu, H. D. Sun, J. X. Pu, Z. H. Liu, M. M. Miao, Y. K. Chen, and G. Y. Yang, *Fitoterapia*, 2016, **108**, 1.
5. X. Feng, Xin, J. S. Wang, J. Luo, and L. Y. Kong, *J. Asian Nat. Prod. Res.*, 2010, **12**, 252.
6. G. Y. Yang, W. Zhao, Y.-K. Chen, Z.-Y. Chen, Q.-F. Hu, and M.-M. Miao, *Asian J. Chem.*, 2013, **25**, 4932.
7. G. H. Kong, Y. P. Wu, W. Li, Z. Y. Xia, Q. Liu, K. M. Wang, P. He, R. Z. Zhu, X. X. Si, and G. Y. Yang, *Heterocycles*, 2016, **92**, 331.
8. X. C. Wei, S. C. Sumithran, A. G. Deaciuc, H. R. Burton, L. P. Bush, L. P. Dwoskin, and P. A. Crooks, *Life Sci.*, 2005, **78**, 495.
9. Y. K. Chen, X. S. Li, G. Y. Yang, Z. Y. Chen, Q. F. Hu, and M. M. Miao, *J. Asian Nat. Prod. Res.*, 2012, **14**, 450.
10. X. M. Gao, X. S. Li, X. Z. Yang, H. X. Mu, Y. K. Chen, G. Y. Yang, and Q. F. Hu, *Heterocycles*, 2012, **85**, 147.

11. M. M. Miao, L. Li, Q. P. Shen, C. B. Liu, Y. K. Li, T. Zhang, F. M. Zhang, P. He, K. M. Wang, R. Z. Zhu, Y. K. Chen, and G. Y. Yang, [Fitoterapia, 2015, 103, 260.](#)
12. Z. Y. Chen, J. L. Tan, G. Y. Yang, M. M. Miao, Z. Y. Chen, and T. F. Li, [Phytochem. Lett., 2012, 5, 233.](#)
13. Y. K. Li, Y. L. Zhao, N. J. Xiang, L. Yang, F. Wang, G. Y. Yang, and Z. Y. Wang, [Heterocycles, 2014, 89, 2771.](#)
14. Y. Wang, C. B. Liu, Q. P. Shen, F. M. Zhang, P. He, Z. H. Liu, H. B. Zhang, X. D. Yang, M. M. Miao, and G. Y. Yang, [Heterocycles, 2015, 91, 1198.](#)
15. H. Q. Leng, J. X. Chen, Y. Hang, Y. X. Duan, G. Y. Yang, Y. K. Chen, Y. D. Guo, Q. F. Hu, and M. M. Miao, [Chem. Nat. Compd., 2014, 49, 1028.](#)
16. J. L. Tan, Z. Y. Chen, G. Y. Yang, M. M. Miao, Y. K. Chen, and T. F. Li, [Heterocycles, 2011, 83, 2381.](#)
17. D. R. Mou, W. Zhao, T. Zhang, L. Wan, G. Y. Yang, Y. K. Chen, Q. F. Hu, and M. M. Miao, [Heterocycles, 2012, 85, 2485.](#)
18. G. Y. Yang, W. Zhao, T. Zhang, Y. X. Duan, Z. H. Liu, M. M. Miao, and Y. K. Chen, [Heterocycles, 2014, 89, 183.](#)
19. S. Z. Shang, W. X. Xu, P. Lei, W. Zhao, J. G. Tang, M. M. Miao, H. D. Sun, J. X. Pu, Y. K. Chen, and G. Y. Yang, [Fitoterapia, 2014, 99, 35.](#)
20. S. Z. Shang, Y. X. Duan, X. Zhang, J. X. Pu, H. D. Sun, Z. Y. Chen, M. M. Miao, G. Y. Yang, and Y. K. Chen, *Phytochem. Lett.*, 2014, 7, 413.
21. S. Z. Shang, W. X. Xu, L. Li, J. G. Tang, W. Zhao, P. Lei, M. M. Miao, H. D. Sun, J. X. Pu, Y. K. Chen, and G. Y. Yang, [Phytochem. Lett., 2015, 11, 53.](#)
22. Q. Wang, S. Ji, S. W. Yu, H. X. Wang, X. H. Lin, T. T. Ma, X. Qiao, C. Xiang, M. Ye, and D. A. Guo, [Fitoterapia, 2013, 85, 35.](#)
23. P. Wang, J. Xu, Q. Wang, S. X. Feng, T. Chen, and C.L. Zhang, *J. Chin. Mater. Med.*, 2013, 38, 1531.
24. M. Zhou, K. Zhou, X.-M. Gao, Z.-Y. Jiang, J.-J. Lv, Z.-H. Liu, G.-Y. Yang, M.-M. Miao, C.-T. Che, and Q.-F. Hu, [Org. Lett., 2015, 17, 2638.](#)
25. M. Zhou, M.-M. Miao, G. Du, S.-Z. Shang, W. Zhao, Z.-H. Liu, G.-Y. Yang, C.-T. Che, Q.-F. Hu, and X.-M. Gao, [Org. Lett., 2014, 16, 5016.](#)
26. Q. F. Hu, B. Zhou, J. M. Huang, Z. Y. Jiang, X. Z. Huang, L. Y. Yang, X. M. Gao, G. Y. Yang, and C. T. Che, [J. Nat. Prod., 2013; 76, 1866.](#)
27. Q. F. Hu, B. Zhou, Y. Q. Ye, Z. Y. Jiang, X. Z. Huang, Y. K. Li, G. Du, G. Y. Yang, and X. M. Gao, [J. Nat. Prod., 2013, 76, 1854.](#)